

Aug 04, 2025

ZIKA Antiviral Immunofluorescence staining Protocol

DOI

<https://dx.doi.org/10.17504/protocols.io.14egn6kzyl5d/v1>

Briana McGovern^{1,2}

¹Icahn School of Medicine at Mount Sinai; ²ASAP Discovery Consortium

ASAP Discovery



Briana L McGovern

Icahn School of Medicine at Mount Sinai

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.14egn6kzyl5d/v1>

Protocol Citation: Briana McGovern 2025. ZIKA Antiviral Immunofluorescence staining Protocol. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.14egn6kzyl5d/v1>

License: This is an open access protocol distributed under the terms of the **[Creative Commons Attribution License](#)**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: September 25, 2024

Last Modified: August 04, 2025

Protocol Integer ID: 108375

Keywords: SARS-CoV-2, immunofluorescence, antiviral staining, Alexa Fluor 488, cell fixation, permeabilization, fluorescence microscopy, zika antiviral immunofluorescence, ic7c7 primary antibody, zika, infected cells for quantification, viral infection, primary antibody, secondary antibody, conjugated secondary antibody, immunofluorescence, labeled infected cell, using ic7c7, staining protocol, fixed sar

Funders Acknowledgements:

National Institutes of Health/National Institute Of Allergy and Infectious Diseases (NIH/NIAID)

Grant ID: U19AI171399

Disclaimer

The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Abstract

This protocol describes immunofluorescence staining of fixed SARS-CoV-2 infected cells using IC7C7 primary antibody (produced in house against SARS-CoV-2 NP) and Alexa Fluor 488-conjugated secondary antibody. The procedure results in fluorescently labeled infected cells for quantification of viral infection.

Protocol materials

⊗ 0.1% Triton X-100-containing 1XPBS solution

⊗ Tween 20 **Bio-Rad Laboratories Catalog #170-6606-MSDS**

⊗ Goat anti-Rabbit IgG (H L) Cross-Adsorbed Secondary Antibody, Alexa Fluor™ 488 **Thermo Fisher Scientific Catalog #A-11008**

Safety warnings

⚠ Please be sure to wear proper Personal Protective Equipment (PPE) while performing this experiment.

Before start

Assay is performed under biosafety level 3 conditions.



Post-fix prep

- 1 Once you have the cells fixed with 10% PFA for minimum 24 hours you may handle the plates. 
- 2 Remove PFA from the wells and discard in the fume hood. 
- 3 Wash the wells 3x with 100ul PBS. 

Blocking

- 4 Add 100ul 1% PBS-BSA to the wells
- 5 Incubate for 1 hour at RT 

Permeabilization

- 6 Treat the wells with 100ul  0.1% Triton X-100-containing 1XPBS solution for 10-15 minutes.
- 7 Remove the triton, and wash 3x with 100ul 0.1%  Tween 20 **Bio-Rad Laboratories Catalog #170-6606-MSDS** 

Primary Antibody

- 8 Treat the wells with 50 ul primary flavivirus envelope protein antibody 4G2 (generated in house by Andrew.Duty@mssm.edu) at 0.001mg/ml (**1:1000**) in 1% BSA 

Incubate overnight at 4C. If urgent, incubate for 1-2 hours at RT
- 9 After incubation, wash 2x with 100ul 0.1%  Tween 20 **Bio-Rad Laboratories Catalog #170-6606-MSDS**



Secondary Antibody

- 10 Treat the wells with 50ul secondary antibody



Goat anti-Rabbit IgG (H L) Cross-Adsorbed Secondary Antibody, Alexa Fluor™
488 Thermo Fisher Scientific Catalog #A-11008

(1:1000) **and DAPI** in 1% BSA

Incubate overnight at 4C. If urgent, incubate 1-2 hours at RT

- 11 After incubation, wash 2x with 100ul 0.1%



Tween 20 Bio-Rad Laboratories Catalog #170-6606-MSDS

- 12 Add 100ul plain PBS to the wells for reading and storage

