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Yale SenNet Human TMC - H&E staining following Phenocycler-Fusion Imaging

Forked from Yale Murine TMC - H&E staining following Phenocycler-Fusion Imaging

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We use this protocol and it's working

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Abstract

This protocol was adapted from (Yale Murine TMC - H&E staining following Phenocycler-Fusion Imaging)
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Flow cell removal after Phenocycler-Fusion imaging and following H&E staining steps from Yale Pathology Tissue Services (YPTS). For FFPE samples, the tissue section is deparaffinized and rehydrated during the preprocessing steps for CODEX, hence these steps do not need to be repeated following theremoval of the flow cell

Troubleshooting

Removing flow cell following CODEX imaging

1 Remove slides from storage buffer.

2 Immerse slides to xylene or HistoClear overnight. However, an incubation of 20 minutes should suffice. 10m

3 Separate flow cell from the slide using a razorblades.

4 Rinse the tissue sample with DI water using at least 1L of DI water to ensure thorough rinsing.

5 Next, wash the tissue sample slide in 1x PhenoCyclerTM Buffer without additive, 3 × 10 minutes.

6 If you are not performing H&E immediately, transfer the tissue sample slide(s) to the PhenoCycler Storage
buffer for storage.

Hematoxylin and eosin staining 11m 30s

7 Stain with hematoxylin for 5 min. 5m

8 Rinse with water.

9 Clarifier for 30 seconds. 30s

10 Rinse with water.

11 Bluing for 1 min. 1m

12 Rinse with water.

13 Stain with eosin 5 min. 5m

14 Immerse slides in graded ethanol and clear slide in xylene.

15 Add coverslip with resin mounting medium.

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