

Sep 08, 2020

DOI

dx.doi.org/10.17504/protocols.io.bk3qkymw



Jon Arizti-Sanz^{1,2}, Catherine A. Freije¹, Chloe K. Boehm¹, Sameed M. Siddiqui^{1,3}, Allen M. Goodman¹, Tinna-Solveig F. Kosoko-Thoroddsen¹, A'Doriann Y. Bradley¹, Jeremy Johnson¹, Pardis C. Sabeti^{1,4,5,6,7}, Cameron Myhryold^{1,8,7}

¹Broad Institute of Massachusetts Institute of Technology (MIT) and Harvard, Cambridge, MA 02142, USA.;

XPRIZE Rapid Covid Tes...



Catherine Freije

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

²Harvard-MIT Program in Health Sciences and Technology, 77 Massachusetts Ave., Cambridge, MA 02139, USA.;

³Computational and Systems Biology PhD Program, MIT, Cambridge, MA 02139, USA.;

⁴Harvard T.H. Chan School of Public Health, 677 Huntington Avenue, Boston, MA 02115, USA.;

⁵Department of Organismic and Evolutionary Biology, Harvard University, 26 Oxford Street, Cambridge, MA 02138, USA.;

⁶Howard Hughes Medical Institute, Chevy Chase, MD 20815, USA.;

⁷Massachusetts Consortium on Pathogen Readiness, Boston, MA, USA.;

⁸Department of Organismic and Evolutionary Biology, Harvard University, 26 Oxford Street, Cambridge, MA 02138, USA



OPEN ACCESS



DOI: https://dx.doi.org/10.17504/protocols.io.bk3qkymw

Protocol Citation: Jon Arizti-Sanz, Catherine A. Freije, Chloe K. Boehm, Sameed M. Siddiqui, Allen M. Goodman, Tinna-Solveig F. Kosoko-Thoroddsen, A'Doriann Y. Bradley, Jeremy Johnson, Pardis C. Sabeti, Cameron Myhrvold 2020. XPRIZE SHINE - Paper-based SARS-CoV-2 NP Test. **protocols.io** https://dx.doi.org/10.17504/protocols.io.bk3qkymw

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: September 08, 2020

Last Modified: September 08, 2020

Protocol Integer ID: 41808

Keywords: CRISPR, SARS-CoV-2, nucleic acid diagnostic, detection of sar, nasopharyngeal sample, based assay, collected nasopharyngeal sample, based sar, rna, cov, sar,

Disclaimer

The protocol and its content is for informational and academic purposes only. It does not constitute legal, medical, clinical, or safety advice, or otherwise. Content added to **protocols.io** is not peer reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol is not a substitute for independent professional judgment, advice, diagnosis, or treatment. Any action taken or not taken based on the protocol presented here is strictly at your own risk. You agree that none of the authors, contributors, administrators, nor anyone else associated with **protocols.io**, can be held responsible for your use of the information contained in or linked to this protocol.

Abstract

This protocol describes how to perform a SHINE paper-based assay to detect SARS-CoV-2 RNA from a self-collected nasopharyngeal sample. This protocol is intended for in-home use. All enzymatic components are provided as a single-test freeze-dried pellet for shelf-stable storage, and all steps of the protocol are performed at ambient temperature. The protocol presented here is an improved version of the method presented in Arizti-Sanz J*, Freije CA*, *et al.* Integrated sample inactivation, amplification, and Cas13-based detection of SARS-CoV-2. *bioRxiv* (2020).



Image Attribution

biorender.com

Materials

MATERIALS

- Nalgene™ Dropper Bottles with Control Dispensing Tip, 4mL, white Thermo Fisher Catalog #2750-9125
- X FastAmp® Viral and Cell Solution for Covid-19 Testing Solution B Catalog #4633
- Reagent Mix A (Paper-based SARS-CoV-2 resuspension mix)
- X Lyophilized Reagent Mix B (Paper-based SARS-CoV-2 detection mix)
- X HybriDetect Universal Lateral Flow Assay Kit Catalog #MGHD 1
- Main Inoculating Loops and Needles Sterile 10 uL VWR International (Avantor) Catalog #12000-810
- 🔀 Custom nasopharyngeal swab and collection tube

STEP MATERIALS

- Nalgene™ Dropper Bottles with Control Dispensing Tip, 4mL, white Thermo
- Reagent Mix A (Paper-based SARS-CoV-2 resuspension mix)
- X Lyophilized Reagent Mix B (Paper-based SARS-CoV-2 detection mix)
- Inoculating Loops and Needles Sterile 10 uL VWR International (Avantor) Catalog #12000-810
- X HybriDetect Universal Lateral Flow Assay Kit Catalog #MGHD 1
- FastAmp® Viral and Cell Solution for Covid-19 Testing Solution B Catalog #4633
- X Custom nasopharyngeal swab and collection tube



Protocol materials

- FastAmp® Viral and Cell Solution for Covid-19 Testing Solution B Catalog #4633
- Inoculating Loops and Needles Sterile 10 uL VWR International (Avantor) Catalog #12000-810
- 🔯 HybriDetect Universal Lateral Flow Assay Kit Catalog #MGHD 1
- Inoculating Loops and Needles Sterile 10 uL VWR International (Avantor) Catalog #12000-810
- X Custom nasopharyngeal swab and collection tube
- X HybriDetect Universal Lateral Flow Assay Kit Catalog #MGHD 1
- 🔯 FastAmp® Viral and Cell Solution for Covid-19 Testing Solution B Catalog #4633
- X Custom nasopharyngeal swab and collection tube
- Reagent Mix A (Paper-based SARS-CoV-2 resuspension mix)
- X Lyophilized Reagent Mix B (Paper-based SARS-CoV-2 detection mix)
- X Lyophilized Reagent Mix B (Paper-based SARS-CoV-2 detection mix)
- , Nalgene™ Dropper Bottles with Control Dispensing Tip, 4mL, white Thermo Fisher Catalog #2750-9125
- Nalgene™ Dropper Bottles with Control Dispensing Tip, 4mL, white Thermo
- Reagent Mix A (Paper-based SARS-CoV-2 resuspension mix)
- Custom nasopharyngeal swab and collection tube
- X FastAmp® Viral and Cell Solution for Covid-19 Testing Solution B Catalog #4633
- Reagent Mix A (Paper-based SARS-CoV-2 resuspension mix)
- X Lyophilized Reagent Mix B (Paper-based SARS-CoV-2 detection mix)
- Nalgene™ Dropper Bottles with Control Dispensing Tip, 4mL, white Thermo
- Inoculating Loops and Needles Sterile 10 uL VWR International (Avantor) Catalog #12000-810
- Mari HybriDetect Universal Lateral Flow Assay Kit Catalog #MGHD 1

Troubleshooting

Safety warnings

Please take care with potentially infectious sample material that does not come into contact with the provided viral lysis solution contained within the NP sample collection tube.



Before start

Download the HandLens application on the user-provided smart device (smartphone, tablet, etc.). Wash hands prior to starting the protocol.



Sample Collection and Viral Lysis

Open the nasopharyngeal (NP) collection tube and rotate the nasal swab (attached to the NP collection tube cap) 4 times around the inside of each nostril. Return the swab to the collection tube and cap the tube.

Nasopharyngeal collection tube contains necessary volume of FastAmp® Viral and Cell Solution.

- X FastAmp® Viral and Cell Solution for Covid-19 Testing Solution B Catalog #4633
- 🛭 Custom nasopharyngeal swab and collection tube
- Mix NP sample and FastAmp® Viral and Cell Solution by shaking the closed sample collection tube for 600000:10
- Wait 00:05:00, incubating sample at Room temperature, before proceeding to Step 4.

SARS-CoV-2 Detection

- Add the entire volume in the dropper bottle containing Reagent Mix A to the tube containing lyophilized Reagent Mix B. Cap the tube.
 - Nalgene™ Dropper Bottles with Control Dispensing Tip, 4mL, white **Thermo**Fisher Catalog #2750-9125

 - X Lyophilized Reagent Mix B (Paper-based SARS-CoV-2 detection mix)
- Mix Reagent Mix A and B by shaking for approximately 00:00:10.
- 6 Dip the inoculation loop into the sample collection tube (a small layer of liquid should be contained within the loop).
 - Inoculating Loops and Needles Sterile 10 uL VWR International (Avantor) Catalog #12000-810



- 7 Transfer liquid in inoculation loop to the Reagent Mix A and B tube by dipping the loop into tube and stirring for 00:00:05. Remove and discard the inoculation loop and cap the tube.
- 8 Mix the sample combined with Reagent Mix A and B by shaking for approximately 00:00:10.
- 9 Wait 01:30:00 , incubating sample at Room temperature , before proceeding to Step 10.

Paper-based Readout and Automated Analysis

- Open the Sample-Reagent Mix A and B tube and place the test strip into the liquid with the arrows on the test strip pointing upward and towards you.
 - ₩ HybriDetect Universal Lateral Flow Assay Kit Catalog #MGHD 1
- 11 Wait 00:05:00 with sample at Room temperature for visible horizontal bands to appear on the test strip.
- With the user-provided smart device such as a smartphone, open the HandLens application and select paper-based as the test type.
- Take a photo of the test strip, and select upload. The result of the test will appear on the smart device screen.