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## Western Blot

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**We use this protocol and it's working**

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## Abstract

Western Blot

## Troubleshooting



- 1 For protein isolation, cell pellets are resuspended in RIPA buffer with 1X protease inhibitors, sonicated, and centrifuged at 10,400xg for 10 minutes at 4°C.
- 2 Supernatant is collected and protein concentration is determined by Bradford assay.
- 3 Protein is mixed with 6x LSB (Sigma-Aldrich) and 4% DTT (Sigma-Aldrich) and heated at 95 °C for 5 minutes in a Thermomixer.
- 4 Samples are subjected to SDS-PAGE on 8 to 12% of polyacrylamide and samples are run with a PowerPac Basic (Bio-Rad Laboratories) at 90-120 Volts for 60-90 minutes in a Mini-Protean® 3 Cell (Bio-Rad Laboratories).
- 5 Gel is transferred at 350 Amperes for 120 minutes onto pre-activated polyvinylidene difluoride (PVDF) or Nitrocellulose membranes in a Mini-Protean® 3 Cell (BioRad Laboratories). Correct protein transfer is checked by red Ponceau S Solution (Panreac).
- 6 Membrane is rinsed 3 times with TBS with 0.1% of Tween20 and blocked at RT for 1 hour with 5% milk or 5% BSA and incubated with primary antibody at 4°C ON.
- 7 The day after, membrane is rinsed and incubated with secondary Horseradish Peroxidase (HRP)-labelled antibody at RT for 1 hour.
- 8 After rinsing, signal is developed with ECL™ start Western Blotting Detection Reagent (GE Healthcare Amersham™) and images are obtained using a ChemiDoc System (Bio-Rad Laboratories).
- 9 Protein amount is expressed as a ratio between the band intensity of the protein of interest and the loading control protein  $\beta$ -Actin or Glyceraldehyde-3- Phosphate Dehydrogenase (GAPDH).