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Version 2

Western Blot V.2

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Protocol status: Working

We use this protocol and it's working

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Abstract

Western Blot using the iBind Automated Western Systems

Materials

10x RIPA buffer (Millipore 20-188)

Protease inhibitor cocktail (AbCAM, ab271306)

Qubit Protein BR Assay kit (A50668)

Qubit4



Collecting cells from the plate

15m

- 1 This protocol is written for a 6-well plate, please adjust the medium/buffer volume based on plate size.
- 2 Aspirate the medium in the wells and wash once with pre-cold DPBS. Add 1 mL of cold DPBS to each well, and collect the cells with cell lifter.
- 3 Centrifuge the tubes at 300g for 5 minutes. Discard the supernatant. If the WB is not performed immediately, the pellet can be stored at -80°C.

10m



5m



Preparing Western Blot lysate with RIPA buffer

55m

- 4 Prepare 1x RIPA buffer to lysis the cells. Use 10x RIPA buffer (Millipore 20-188) to prepare 1X RIPA buffer (2mL RIPA buffer + 18mL H₂O).
- 5 Add protease inhibitor cocktail (1x RIPA buffer + Protease inhibitor cocktail (1:100). At this step, 1x RIPA buffer with inhibitor can be aliquoted and frozen down at -20°C.
- 6 Add 100 ul RIPA buffer to the cell pellet and re-suspend the cells. Leave the RIPA lysate on ice for 30 minutes. Pipetting to mix every 10 minutes until the lysate is clear.
- 7 Centrifuge the lysate for 10 minutes at 15000g at 4°C (important that it is done at 4°C). Transfer the supernatant to a new 1.5mL Eppendorf tube. Make sure that the step is performed on ice. The supernatant will be your samples for the Western Blot.
- 8 At this step, cell lysis can be stored at -80°C for long-term storage.

5m

5m



30m



15m



Measuring Protein concentration Using Qubit Protein BR Assay

40m

- 9 Use the Qubit Protein BR Assay kit (A50668) and Qubit4 to measure the protein concentration. In the 0.5mL Qubit Invitrogen tubes, prepare a Blank, Standard 1, Standard 2, and samples according to the following, ensuring that the order added is Sample (or Standard), followed by the BR buffer, followed by the kit reagent:

20m



Blank: Add 170uL of BR buffer + 30uL of Reagent



Standard 1: Add 20uL of S1 + 150uL BR Buffer + 30uL Reagent
Standard 2: Add 20uL of S2 + 150uL BR Buffer + 30uL Reagent
Sample: Add 2uL of Sample + 168uL of BR Buffer + 30uL of Reagent

- 10 Vortex each of the blank, standards, and samples, and leave at room temperature for 10 minutes. Measure the protein concentration.

20m



Sample Preparation and Boiling

5m

- 11 In a separate 1.5mL Eppendorf tube, prepare a 1:9 ratio of 4x Laemmli Sample Buffer to 2-mercaptoethanol. Prepare enough for 34uL of this mix per sample.
- 12 Add 34uL of the loading mix to 100uL of lysis sample, and boil the samples for 3-5 minutes. At this point, prior to boiling, samples can be stored long-term at -80°C.

5m

II

Gel Preparation and Running

1h 20m

- 13 After boiling samples, prepare 1X running buffer: mix 100mL of 10X Tris/Glycine/SDS Buffer with 900mL of Milli-Q water.
- 14 Prepare mini-protein TGX stain-free gels (4-20%), in the BIO-RAD running cassette, and add running buffer.
- 15 Load samples onto wells depending on protein concentration (up to 20uL/well). Add 8uL of loading marker dye to each end.
- 16 Run the gel at 100V for 10 minutes, and then at 150V for 30-40 minutes.

5m

5m

10m

1h

Transfer with Trans-Blot Turbo Transfer System

1h 30m

- 17 Transfer gel to PVDF membrane using the Trans-Blot Turbo Mini 0.2 um PVDF Transfer Packs. Transfer for 30 minutes.
- 18 After the transfer, membranes are placed in iBind buffer. If alpha-synuclein and pS129-alpha-synuclein are to be blotted, fix the membrane with 4% PFA in PBS for 10 minutes and wash with PBS for 30 minutes before blotting.

30m

1h



Protein Immunodetection with iBind Automated Western Systems

8h 40m

- 19 Following transfer, prepare 1X iBind Solution by adding: 300uL of 100X Additive + 6mL iBind 5X buffer + 23.7mL H₂O to a 50mL falcon tube. 10m
- 20 Prepare primary antibodies, secondary antibodies and washing buffer according to iBind kit protocol. Add them to iBind Automated Western Systems and run overnight. 8h

- 21 Detect the signal with Max Western ECL and image with Biorad ChemiDoc Imaging system 30m