VERSION 2

FEB 01, 2020

OPEN ACCESS

DOI:
dx.doi.org/10.17504/protocols.io.bb2wqife

Protocol Citation: Elizabeth Neumann, Jamie Allen, Maya Brewer, David Anderson, Mark De Caestecker, Danielle Gutierrez, Jeff Spraggins 2020. VU Biomolecular Multimodal Imaging Center (BIOMIC) kidney characterization pipeline for tissues collected through the Cooperative Human Tissue Network (CHTN). **protocols.io** <https://dx.doi.org/10.17504/protocols.io.bb2wqife>

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Protocol status: Working
 We use this protocol and it's working

Created: Feb 01, 2020

Last Modified: Apr 08, 2023

PROTOCOL integer ID:
 32566

Keywords: HuBMAP, BIOMIC, Vanderbilt, Multimodal Imaging, IMS, MxIF, Proteomics, Image Registration

VU Biomolecular Multimodal Imaging Center (BIOMIC) kidney characterization pipeline for tissues collected through the Cooperative Human Tissue Network (CHTN) V.2

Elizabeth Neumann¹, Jamie Allen¹, Maya Brewer¹, David Anderson¹, Mark De Caestecker², Danielle Gutierrez¹, Jeff Spraggins¹

¹Vanderbilt University; ²Division of Nephrology, Vanderbilt University Medical Center

VU Biomolecular Multimodal Imaging Center / Spraggins Research Group

Human BioMolecular Atlas Program (HuBMAP) Method Development

Jamie Allen
 Vanderbilt University

ABSTRACT

We aim to develop high resolution, chemically informative imaging methodologies for building an atlas of human organs, such as the kidney.

Scope:

Provide an overview of the methods used by the Vanderbilt Tissue Mapping Center as part of the Human Biomolecular Atlas Program (HuBMAP, NIH Common Fund) and contextualize individual protocols within our larger workflow.

- 1 Collection of post-surgical tissue.
 Collection: dx.doi.org/10.17504/protocols.io.7gehjte
- 2 Stabilize and freeze tissues.
 Freezing Tissue: dx.doi.org/10.17504/protocols.io.6wghfbw
- 3 Initial Rapid Pathology Assessment of Kidney Tissue
 Staining: dx.doi.org/10.17504/protocols.io.4qngvve
 Assessment: dx.doi.org/10.17504/protocols.io.9dph25n
- 4 Cryosection tissues into micrometer thick sections, alternating between thaw mounting onto indium tin-oxide and positively charged glass slides (proceed to step 4), or collecting several tissue sections within a microcentrifuge tube for proteomics analysis.
 Cryosectioning: dx.doi.org/10.17504/protocols.io.7ethjen

Proteomics: [dx.doi.org/10.17504/protocols.io.67nhhme](https://doi.org/10.17504/protocols.io.67nhhme)

- 5 Perform autofluorescence microscopy on all tissue sections before IMS (step 5) or MxIF analysis (step 9)
Autofluorescence: [dx.doi.org/10.17504/protocols.io.7e3hjgn](https://doi.org/10.17504/protocols.io.7e3hjgn)
- 6 Coat tissue sections with MALDI matrix for IMS analysis.
Matrix Application: [dx.doi.org/10.17504/protocols.io.4srgwd6](https://doi.org/10.17504/protocols.io.4srgwd6)
- 7 Perform high resolution IMS analysis of matrix coated tissue sections.
IMS: [dx.doi.org/10.17504/protocols.io.7gdhjs6](https://doi.org/10.17504/protocols.io.7gdhjs6)
- 8 Perform fluorescence microscopy to visualize laser ablation spots.
Post IMS AF: [dx.doi.org/10.17504/protocols.io.879hxr6](https://doi.org/10.17504/protocols.io.879hxr6)
- 9 Annotation of Lipids from IMS Data
Cal & Annotate: [dx.doi.org/10.17504/protocols.io.864hzgw](https://doi.org/10.17504/protocols.io.864hzgw)
- 10 Remove MALDI matrix and perform PAS staining.
PAS Staining: [dx.doi.org/10.17504/protocols.io.4qngvve](https://doi.org/10.17504/protocols.io.4qngvve)
- 11 Alternatively, MxIF can be performed after step 4.
Antibody labeling: [dx.doi.org/10.17504/protocols.io.667hnhn](https://doi.org/10.17504/protocols.io.667hnhn)
MxIF: [dx.doi.org/10.17504/protocols.io.665hhg6](https://doi.org/10.17504/protocols.io.665hhg6)
- 12 Registration of autofluorescence images from both IMS and MxIF sections allow for the direct correlation of the two orthogonal approaches.
Registration: <https://doi.org/10.1021/acs.analchem.8b02884>
- 13 RNA Assessment from tissue.
RNA extraction: [dx.doi.org/10.17504/protocols.io.86nhzde](https://doi.org/10.17504/protocols.io.86nhzde)