

Sep 21, 2020

VEPextended

DOI

dx.doi.org/10.17504/protocols.io.bkhvkt66

Israel Aguilar Ordoñez¹

¹Instituto Nacional de Medicina Genómica (INMEGEN)

Whole genome variation...



Judith Ballesteros Villascán

Centro de Investigación y de Estudios Avanzados del IPN (Cin...

OPEN  ACCESS



DOI: dx.doi.org/10.17504/protocols.io.bkhvkt66

External link: <https://github.com/laguilaror/nf-VEPextended>

Protocol Citation: Israel Aguilar Ordoñez 2020. VEPextended. **protocols.io**

<https://dx.doi.org/10.17504/protocols.io.bkhvkt66>

Manuscript citation:

Aguilar-Ordoñez I, Pérez-Villatoro F, García-Ortiz H, Barajas-Olmos F, Ballesteros-Villascán J, González-Buenfil R, Fresno C, Garcíarrubio A, Fernández-López JC, Tovar H, Hernández-Lemus E, Orozco L, Soberón X, Morett E (2021) Whole genome variation in 27 Mexican indigenous populations, demographic and biomedical insights. PLoS ONE 16(4): e0249773. doi: [10.1371/journal.pone.0249773](https://doi.org/10.1371/journal.pone.0249773)

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: August 30, 2020

Last Modified: September 21, 2020

Protocol Integer ID: 41237

Abstract

'VEPextended' is a tool, implemented in Nextflow, that annotates called variants using Variant Effect Predictor (VEP) and additional plugins that implement functionalities, that are not included in variation API.

All steps described are mk modules of code that will be done automatically through Nextflow pipeline.

Guidelines

Installation

Download VEPextended from Github repository:

```
git clone https://github.com/Iaguilaror/nf-VEPextended.git
```

Compatible OS*:

- **Ubuntu 18.04.03 LTS**

* VEPextended may run in other UNIX based OS and versions, but testing is required.

Software Requirements:

Software

bcftools

NAME

Software

htslib

NAME

Software

ensembl-vep

NAME

Software

Nextflow

NAME

Software

Plan9

NAME

<https://github.com/9fans/plan9port>

SOURCE LINK



Materials

Pipeline Inputs

- A compressed vcf file with extension '.vcf.gz', which must have a TABIX index with .tbi extension, located in the same directory as the vcf file.

Note(s): INFO must contain AC

Example line(s):

```
##fileformat=VCFv4.2
##FILTER=
##GATKCommandLine.ApplyRecalibration.2=##INFO=
##INFO=
##contig=
##contig=
##contig=
##bcftools_viewCommand=view --compression-level 0 --output-type z --min-ac 1 --threads
1 ./test/data/sample.vcf.gz;
#CHROM POS ID REF ALT QUAL FILTER INFO
chr22 30000353 . G T . PASS
AC=107;AF_mx=0.708;AN=150;DP=883;nhomalt_mx=39
```

- *_dbSNP.vcf.gz: A reference dbSNP b152 file, that contains rsID of variants.

Dataset

dbSNPb152_GRCH38_for_GATK.vcf.gz NAME

<https://ftp.ncbi.nlm.nih.gov/snp/> LINK

- Pre-scored files for SNVs and InDels compressed and indexed provided from CADD. Availables in

Dataset

*_InDels.tsv.gz, *_SNVs.tsv.gz NAME

<https://cadd.gs.washington.edu/download> LINK

- Liftedover of gnomAD release 2.1.1 to GRCh38



Dataset

completegenome_gnomAD.vcf.bgz

NAME

<https://gnomad.broadinstitute.org/downloads>

LINK

- Coverages from gnomAD v 2.1.1

Dataset

gnomad.genomes.coverage.summary.bed.gz

NAME

<https://storage.googleapis.com/gnomad-public/release/2.1/coverage/genomes/gnomad.genomes.coverage.summary.tsv.bgz>

LINK

- GWAS association for SNVs (no haplotypes) compiled by iaguiar from GWAScat database at Spring 2019

Dataset

All_20180418_noINFO.GWAScatalog.vcf.gz

NAME

<https://www.ebi.ac.uk/gwas/docs/file-downloads>

LINK

- Coordinates for every pre-miRNA, mature miRNA and seed region from miRBase v22.

Dataset

miRBase.bed.gz

NAME

<ftp://mirbase.org/pub/mirbase/CURRENT/>

LINK

- _coverages.bed.gz: contains coverage of your sample.
- genotype - drug associations from PGKB

Dataset

var_drug_ann.tsv

NAME

<https://www.pharmgkb.org/downloads>

LINK



Before start

Test

To test VEPextended's execution using test data, run:

```
./runtest.sh
```

Your console should print the Nextflow log for the run, once every process has been submitted, the following message will appear:

```
=====  
VEP annotator: Basic pipeline TEST SUCCESSFUL  
=====
```

VEPextended results for test data should be in the following file:

```
test/results/_pos1_rejoin_chromosomes/sample.filtered.untangled_multiallelics.anno_dbS  
NP_vep.vcf.gz
```

Usage

To run VEPextended go to the pipeline directory and execute:

```
nextflow run vep-annotator.nf --vcffile [ --output_dir path to results ] [-resume]
```

For information about options and parameters, run:

```
nextflow run vep-annotator --help
```




Pre-processing

1 **Filter VCF**

Remove the variants that did not have any copy of the alternative allele.

Note

- a) Filter and remove when there was not an alternative allele in the VCF file.vcf.gz, to only conserve found variants.
- b) Compress the filtered file using one thread for compression.
- c) Make and index output file using the filtered and compressed file.

Dependencies:

Software

bcftools

NAME

2 **Extract chromosomes**

Extract variants per chromosome vcf file that have at least one variant.

Note

- a) Using the index of the compressed vcf file, list the chromosomes names.
- b) If there is at least one variant per chromosome, separate variants per chromosome.

Dependencies:

Software

bcftools

NAME

**Software**

htslib

NAME

3 Split chromosomes*Split in chunks a vcf file, keeping its format.***Note**

- a) Save the header of a vcf in a temporary file.
- b) Save the body of a vcf in a temporary file.
- c) Make chunks of the body of the vcf file.
- d) Add the header to each body chunk.

Dependencies:**Software**

bcftools

NAME

Core-processing**4 Untangle multiallelic***Split multiallelic sites.***Note**

- a) Separate multiallelic sites and conserve vcf format.
- b) Do not print bcftools version.

Dependencies:

**Software**

bcftools

NAME

5 Annotate rsID*Annotate rsID to each variant in ID column of a VCF.***Note**

- a) Make a Reference file with a define range.
- b) Compress input file.
- c) Annotate rsID in the compressed input file using a Reference.

Dependencies:**Software**

bcftools

NAME

Software

htslib

NAME

6 Vep Extended*Annotate variants with Variant Effector Predictor tool (VEP). For more information about, see [VEP](#)***Dependencies:**



Software

ensembl-vep

NAME

Pos-processing

7 Rejoin chromosomes

Concatenate annotated chunks in a single vcf file.

Dependencies:

Software

bcftools

NAME

Software

htslib

NAME

Final Output:

Expected result

VCF file with only variants of each chromosome from the input.

Example line(s):



```
##fileformat=VCFv4.2 #CHROM      POS      ID      REF      ALT      AC
AN      DP      AF_mx    nhomalt_mx    Allele  Consequence
IMPACT  SYMBOL  Gene     Feature_type  Feature BIOTYPE EXON
INTRON  HGVSc    HGVSp    cDNA_position CDS_position
Protein_position      Amino_acids    Codons    Existing_variation
DISTANCE              STRAND  FLAGS    VARIANT_CLASS  SYMBOL_SOURCE
HGNC_ID CANONICAL      TSL      APPRIS  CCDS      ENSP      SWISSPROT
TREMBL  UNIPARC  SOURCE  GENE_PHENO      SIFT      PolyPhen
DOMAINS HGVS_OFFSET      HGVSg    AF      AFR_AF    AMR_AF    EAS_AF
EUR_AF  SAS_AF  AA_AF    EA_AF    gnomAD_AF      gnomAD_AFR_AF
gnomAD_AMR_AF    gnomAD_ASJ_AF    gnomAD_EAS_AF    gnomAD_FIN_AF
gnomAD_NFE_AF    gnomAD_OTH_AF    gnomAD_SAS_AF    MAX_AF
MAX_AF_POPS      CLIN_SIG      SOMATIC PHENO      PUBMED    MOTIF_NAME
MOTIF_POS      HIGH_INF_POS    MOTIF_SCORE_CHANGE
GeneHancer_type_and_Genes      gnomADg gnomADg_AC      gnomADg_AN
gnomADg_AF      gnomADg_DP      gnomADg_AC_nfe_seu
gnomADg_AN_nfe_seu      gnomADg_AF_nfe_seu
gnomADg_nhomalt_nfe_seu gnomADg_AC_raw    gnomADg_AN_raw
gnomADg_AF_raw    gnomADg_nhomalt_raw    gnomADg_AC_afr
gnomADg_AN_afr    gnomADg_AF_afr    gnomADg_nhomalt_afr
gnomADg_AC_nfe_onf      gnomADg_AN_nfe_onf      gnomADg_AF_nfe_onf
gnomADg_nhomalt_nfe_onf gnomADg_AC_amr    gnomADg_AN_amr
gnomADg_AF_amr    gnomADg_nhomalt_amr    gnomADg_AC_eas
gnomADg_AN_eas    gnomADg_AF_eas    gnomADg_nhomalt_eas
gnomADg_nhomalt    gnomADg_AC_nfe_nwe      gnomADg_AN_nfe_nwe
gnomADg_AF_nfe_nwe      gnomADg_nhomalt_nfe_nwe gnomADg_AC_nfe_est
gnomADg_AN_nfe_est      gnomADg_AF_nfe_est
gnomADg_nhomalt_nfe_est gnomADg_AC_nfe    gnomADg_AN_nfe
gnomADg_AF_nfe    gnomADg_nhomalt_nfe      gnomADg_AC_fin
gnomADg_AN_fin    gnomADg_AF_fin    gnomADg_nhomalt_fin
gnomADg_AC_asj    gnomADg_AN_asj    gnomADg_AF_asj
gnomADg_nhomalt_asj      gnomADg_AC_oth    gnomADg_AN_oth
gnomADg_AF_oth    gnomADg_nhomalt_oth    gnomADg_popmax
gnomADg_AC_popmax      gnomADg_AN_popmax      gnomADg_AF_popmax
gnomADg_nhomalt_popmax gnomADg_cov      gwascatalog
gwascatalog_GWASC_trait gwascatalog_GWASC_pvalue
gwascatalog_GWASC_study clinvar clinvar_CLNDN    clinvar_CLNSIG
clinvar_GENEINFO      clinvar_CLNDISDB      miRBase
pharmgkb_drug    pharmgkb_drug_PGKB_annid
pharmgkb_drug_PGKB_gene pharmgkb_drug_PGKB_chem
pharmgkb_drug_PGKB_phencat chr22      16132524      .      G
C      27      156      1962    0.173    3      C
intergenic_variant      MODIFIER      .      .      .      .
.      .      .      .      .      .      .      .
```

chr	pos	ref	alt	SNV
chr22	16132524	G	T	chr22:g.16132524G>C
chr22	27840687	G	T	