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Vascular perfusion of mice V.2

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Protocol status: Working

We use this protocol and it's working

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Abstract

Summary:

This protocol describes the procedure for perfusion of mice prior to organ or tissue harvesting.

Diabetic Complications:



Cardiovascular



Retinopathy



Neuropathy



Nephropathy





Uropathy

Materials

| Reagents and Materials | Quantity Required |
|-----------------------------------|--------------------------|
| Perfusion Apparatus | 1 |
| Perfusion Bottles | 2 |
| Small diameter tubing (< 1cm) | |
| butterfly 23 gauge ¾ | 1 |
| Ketamine | 3 ml |
| Xylazine | 1 ml |
| Plastic Tray | 1 |
| Phosphate Buffered Saline, pH 7.4 | 4 |
| PFA | |
| Sucrose | |
| 70% alcohol | |
| Scissors | |
| Cotton swabs | |



1 **Preparation (see figure below for setup)**

- ◆ Fix tubing (small diameter tubes, below 1 cm) to the bottles: 1 bottle for PFA, 1 bottle for sucrose
- ♦ Fill perfusion solutions in the bottles: solutions must be freshly prepared and filtered (using 0.22 µm filter)
- ◆ Adjust perfusion pressure to 150-160 mm Hg
- ◆ Fix needle at the end of tubing after valve: butterfly 23 gauge 3/4
- ◆ Remove any air from all tubing!!
- ◆ Anesthetic: ketamine-xylazine mix (150/10 mg/kg=0.1 ml/20 g BW)
- ◆ Mix 3 ml ketamine 100mg/ml + 1 ml xylazine 20mg/ml + 6 ml sterile saline
- ♦ Fix the anesthetized animal in a plastic tray
- ♦ Solution #1: 4 % PFA in PBS, ph 7.4
- ♦ Solution #2: 18% Sucrose in PBS, ph 7.4

2 **Perfusion:**

- ◆ Swab mouse with 70% alcohol to wet the fur, cut open abdominal skin by a longitudinal incision
- ◆ Remove skin and gut to expose abdomal aorta and Vena cava
- ♦ Clean the aort abdominalis and vena cava from connective tissue and fat with cotton swabs
- ♦ Clamp aorta abdominalis and vena cava in the area of the iliac arteries
- ◆ Insert the butterfly 23 gauge 3/4 needle in the Aorta below the renal arteries
- ♦ Immediately cut open the Vena cava with small and sharp scissors: this should allow a rapid drain of the perfusion solutions!
- ◆ Watch the needle carefully during the entire perfusion!!!



- ◆ Start perfusion for 3 minutes with PFA, pH 7.4 at 37°C
- ♦ Switch valve to Solution #2:
- ♦ Without any interruption of pressure/flow perfuse for 5 minutes with sucrose, 7.4 at 37°C
- ◆ Stop perfusion, remove butterfly
- ◆ Cut out kidneys at renal hilum for further processing
- ◆ Before starting to perfuse the next animal rinse the tubing and needle extensively

3 **Schematic of custom-made perfusion system**

