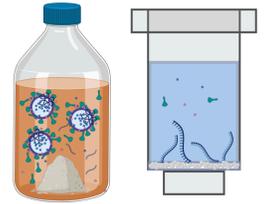


Aug 14, 2020

Version 2

V.2 - Direct wastewater RNA capture and purification via the "Sewage, Salt, Silica and SARS-CoV-2 (4S)" method V.2



DOI

<https://dx.doi.org/10.17504/protocols.io.bjr9km96>

Oscar Whitney¹, Basem Al-Shayeb², Alex Crits-Cristoph³, Mira Chaplin⁴, Vinson Fan¹, Hannah Greenwald⁴, Adrian Hinkle⁴, Rose Kantor⁴, Lauren Kennedy⁴, Anna Maurer¹, Robert Tjian⁵, Kara L. Nelson⁶, UC Berkeley Wastewater-based epidemiology consortium⁶

¹University of California, Berkeley, Tjian & Darzacq laboratory;

²University of California, Berkeley, Banfield & Doudna laboratory;

³University of California, Berkeley, Banfield laboratory; ⁴University of California, Berkeley, Nelson laboratory;

⁵University of California, Berkeley, HHMI; ⁶University of California, Berkeley

Coronavirus Method De...



Oscar N Whitney

University of California, Berkeley, Tjian & Darzacq Laborato...

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.bjr9km96>

Protocol Citation: Oscar Whitney, Basem Al-Shayeb, Alex Crits-Cristoph, Mira Chaplin, Vinson Fan, Hannah Greenwald, Adrian Hinkle, Rose Kantor, Lauren Kennedy, Anna Maurer, Robert Tjian, Kara L. Nelson, UC Berkeley Wastewater-based epidemiology consortium 2020. V.2 - Direct wastewater RNA capture and purification via the "Sewage, Salt, Silica and SARS-CoV-2 (4S)" method. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.bjr9km96>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: August 14, 2020

Last Modified: August 14, 2020

Protocol Integer ID: 40481

Keywords: SARS-CoV-2, Wastewater-based epidemiology, Direct capture, RNA extraction, COVID-19, existing wastewater rna purification method, wastewater rna purification method, direct wastewater rna capture, rna extraction, handling raw wastewater sample, rna, raw wastewater sample, wastewater, purification, sewage, extraction,

Abstract

This protocol describes the procedure of the "4S" (Sewage, Salt, Silica and SARS-CoV-2) method for SARS-CoV-2 RNA extraction from wastewater. Offering a highly efficient, modular and economical alternative to existing wastewater RNA purification methods, this procedure lowers the barrier to entry for SARS-CoV-2 wastewater-based epidemiology. This procedure is intended to be carried out in a BSL2+ laboratory space, with precautions when handling raw wastewater samples.

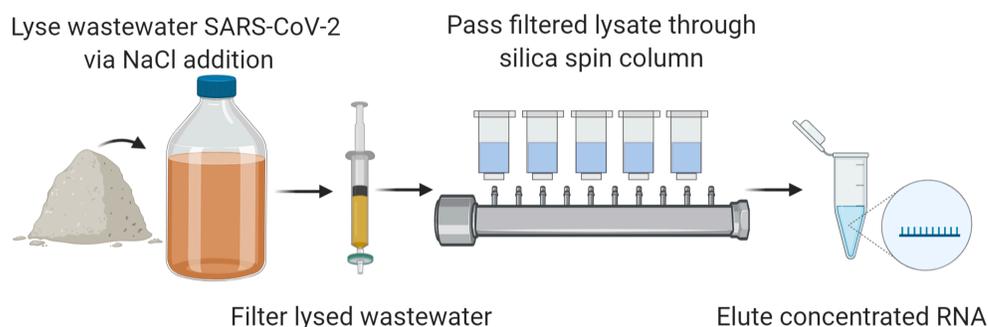


Image Attribution

Figures created with BioRender.com

Guidelines

DISCLAIMER – FOR INFORMATIONAL PURPOSES ONLY; USE AT YOUR OWN RISK

The protocol content here is for informational purposes only and does not constitute legal, medical, clinical, or safety advice, or otherwise; content added to [protocols.io](https://www.protocols.io) is not peer reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol should not substitute for independent professional judgment, advice, diagnosis, or treatment. Any action you take or refrain from taking using or relying upon the information presented here is strictly at your own risk. You agree that neither the Company nor any of the authors, contributors, administrators, or anyone else associated with [protocols.io](https://www.protocols.io), can be held responsible for your use of the information contained in or linked to this protocol or any of our Sites/Apps and Services.

Materials

MATERIALS

 Tris

 EDTA

 Sodium Chloride **Catalog #PubChem CID: 5234**

 Microcentrifuge

 Ethanol

 Zymo III-P column **Zymo Research Catalog #C1040-5**

 EZ-Vac Vacuum Manifold **Zymo Research Catalog #S7000**

 Durapore® Membrane Filter 5.0 µm **Merck MilliporeSigma (Sigma-Aldrich) Catalog #SVLP04700**

 Magnetic Funnel 300mL 47mm **Pall Catalog #4242**

 Bovilis Coronavirus Calf Vaccine **Merck Animal Health Catalog #16445**

 Swinnex Filter Holder **Merck MilliporeSigma (Sigma-Aldrich) Catalog #SX0004700**

 ZymoPURE Elution Buffer **Zymo Research Catalog #D4200-7-30**

STEP MATERIALS

 Durapore® Membrane Filter 5.0 µm **Merck MilliporeSigma (Sigma-Aldrich) Catalog #SVLP04700**

 Swinnex Filter Holder **Merck MilliporeSigma (Sigma-Aldrich) Catalog #SX0004700**

 Magnetic Funnel 300mL 47mm **Pall Catalog #4242**

 EZ-Vac Vacuum Manifold **Zymo Research Catalog #S7000**

 Zymo III-P column **Zymo Research Catalog #C1040-5**

 ZymoPURE Elution Buffer **Zymo Research Catalog #D4200-7-30**

 TE buffer

 Bovilis Coronavirus Calf Vaccine **Merck Animal Health Catalog #16445**

Protocol materials

 ZymoPURE Elution Buffer **Zymo Research Catalog #D4200-7-30**

 Swinnex Filter Holder **Merck MilliporeSigma (Sigma-Aldrich) Catalog #SX0004700**

 Zymo III-P column **Zymo Research Catalog #C1040-5**

 Ethanol

 Durapore® Membrane Filter 5.0 µm **Merck MilliporeSigma (Sigma-Aldrich) Catalog #SVLP04700**

 Tris

 EDTA

 Magnetic Funnel 300mL 47mm **Pall Catalog #4242**

 Sodium Chloride **Catalog #PubChem CID: 5234**

 EZ-Vac Vacuum Manifold **Zymo Research Catalog #S7000**

 Bovilis Coronavirus Calf Vaccine **Merck Animal Health Catalog #16445**

 TE buffer

 Microcentrifuge

Troubleshooting

Safety warnings

-  Wastewater is intrinsically hazardous, so we advise handling wastewater samples in a biosafety cabinet in a BSL2+ laboratory space.

Before start

We developed this procedure to provide a highly efficient, economical and rapid method for extraction of SARS-CoV-2 RNA from wastewater. Using this procedure at the University of California Berkeley, we have captured and quantified SARS-CoV-2 and pepper mild mottle virus (PMMoV) present in a variety of San Francisco Bay Area raw wastewater influent samples and samples collected upstream of wastewater treatment plants. Results may vary depending on wastewater sample type and laboratory setting.

This procedure relies on vacuum column processing, which can be performed with a vacuum manifold and vacuum pump or central vacuum line. In our laboratory, this procedure yields concentrated and purified wastewater RNA in less than 3 hours.

In our laboratory, this purification method enables the detection of SARS-CoV-2 N and E gene RNA as well as PMMoV RNA via RT-qPCR probe-mediated detection. Depending on sample origin, we are able to recover an average of 35 ng RNA/mL of purified wastewater sample (min = 9.33 ng/mL, max = 95 ng/mL).

Preparing RNA wash buffers

- 1 Prepare 1 L each of two wash buffers - Wash buffer #1 (4S-WB1) and #2 (4S-WB2), for later use during cleanup of RNA bound to silica columns.

1.1 4S-WB1 composition:

Reagent	Original molarity/%	Final molarity/%	Volume per liter of buffer
NaCl	5 M	1.5 M	300 mL
Ethanol	100%	20%	200 mL
TRIS pH 7.2	1 M	10 mM	10 mL
Pure water (MilliQ or distilled)	NA	NA	490 mL

Add 490 mL water to sterile bottle

Add 300 mL of 5 Molarity (M) NaCl

Add 200 mL of 100 % volume Ethanol

Add 10 mL of 1 Molarity (M) pH 7.2 TRIS

Agitate to fully mix buffer solution

1.2 4S-WB2 composition:

Reagent	Original molarity/%	Final molarity/%	Volume per liter of buffer
NaCl	5 M	100 mM	20mL
Ethanol	100%	80%	800mL
TRIS pH 7.2	1 M	10 mM	10mL
Pure water (MilliQ or distilled)	NA	NA	170mL

Add 170 mL water to sterile bottle

Add 20 mL of 5 Molarity (M) NaCl

Add 800 mL of 100 % volume Ethanol

Add 10 mL of 1 Molarity (M) pH 7.2 TRIS

Agitate to fully mix buffer solution

Sample preparation, RNA preservation and particle lysis

- 2 Obtain a  40 mL wastewater sample in a sterile sample collection tube. Maintain at  4 °C during transport to the lab.

Note

Sodium chloride and TE buffer (Go to step 4) can be added to sample immediately after collection. Our unpublished analysis demonstrates that Sodium chloride & TE buffer preserve RNA present in wastewater.

- 3 Resuspend dry bovine coronavirus stock (Bovilis Coronavirus Calf Vaccine) in  2 mL of PBS. Dilute this resuspended stock into PBS at a dilution of 1:10 ( 100 µL of stock into  900 µL PBS). Spike 50uL of diluted bCoV into the wastewater sample as a recovery efficiency control. Agitate sample to fully mix bCoV or other spiked-in controls with the wastewater sample.

 Bovilis Coronavirus Calf Vaccine **Merck Animal Health Catalog #16445**

Note

Other recovery controls can be used instead of bCoV. Some candidates include Phi6 bacteriophage and coronavirus OC43. In addition, purified RNAs can be used to quantify the extraction efficiency of "free RNA".

- 4 Add  9.5 g of sodium chloride to  40 mL wastewater sample.
 Make  7.2 TE buffer ( 1 Molarity (M) TRIS,  100 millimolar (mM) EDTA).
 Add  400 µL of TE buffer to  40 mL wastewater sample.

Note

Here, NaCl lyses lipid-protein envelopes, denatures proteins and disrupts RNA-protein interactions. EDTA inhibits the enzymatic degradation of RNA by RNases present in wastewater and TRIS provides optimal buffering conditions for nucleic acids.

- 4.1 Agitate sample until all NaCl dissolves in the wastewater. Vortex or shake sample for  00:00:30 to promote lysis.



Raw wastewater containing NaCl, TRIS & EDTA.



- 5 (OPTIONAL) Heat inactivate sample at  70 °C for  00:30:00 . Our unpublished analyses have shown that this step will not affect SARS-CoV-2 RNA enrichment and detection.
- 6 Filter the sample through a 5-um PVDF filter via syringe filtration or funnel top vacuum.





Syringe filter setup: Wastewater is filtered through a 47-mm reusable filter membrane holder.

⊗ Durapore® Membrane Filter 5.0 μm Merck MilliporeSigma (Sigma-Aldrich) Catalog #SVLP04700

⊗ Swinnex Filter Holder Merck MilliporeSigma (Sigma-Aldrich) Catalog #SX0004700

⊗ Magnetic Funnel 300mL 47mm Pall Catalog #4242



Wastewater filtering through a 5-um PVDF filter in a Pall filter holder.

Direct RNA extraction (RNA Binding, Washing, Eluting)

- 7 Aliquot 40 mL filtrate into two 20 mL aliquots. Add 20 mL of 70 % volume ethanol to each 20 mL sample filtrate aliquot.



Filtered sample before ethanol addition. Filtrate should be semi-clear.

- 7.1 Agitate sample to mix ethanol and wastewater lysate.
- 8 Attach Zymo III-P (or other) silica spin column to a vacuum manifold. Vacuum the full  80 mL volume (both aliquots) of wastewater lysate & ethanol through the spin column.

Note

Commercial silica spin columns vary in their silica membrane packing tightness, changing the flow rate of lysed wastewater. We advise the use of the Zymo III-P column to avoid column clogging issues, but columns such as the Qiagen RNeasy, QIAamp Mini Spin and Zymogen II-CR can act as substitutes, depending on vacuum strength and sample particulate content. Large-format "maxiprep" style columns are also able to purify wastewater RNA, but require a large volume RNA elution up to 20mL (Step 13) and a downstream precipitation-concentration step (Isopropanol precipitation, see [companion protocol](#), Step 12) .



Passing lysed & filtered samples through Zymo III-P columns for direct RNA capture.

 EZ-Vac Vacuum Manifold Zymo Research Catalog #S7000

 Zymo III-P column **Zymo Research Catalog #C1040-5**

9 Vacuum  5 mL wash buffer #1 (4S-WB1) through the silica spin column.

10 Vacuum  10 mL wash buffer #2 (4S-WB2) through the silica spin column.

RNA elution

11 Detach silica spin column from vacuum manifold, remove any attached reservoirs/funnels and place column into a 1.5-mL centrifugation-compatible flowthrough collection tube.

12 Centrifuge silica spin column in tube at  10000 x g, 4°C, 00:02:00 to remove any residual 4S-WB2 present in the column.

12.1 Discard the collection tube and place silica column into a new 1.5-mL centrifugation-compatible flowthrough collection tube.

13 Pre-warm  200 μ L of ZymoPURE elution buffer or  200 μ L  8 TE buffer per RNA sample to  50 °C in a heat block, waterbath or incubator.

 ZymoPURE Elution Buffer **Zymo Research Catalog #D4200-7-30**

 TE buffer

13.1 Add  200 μ L of pre-warmed elution buffer to each silica spin column. Incubate the elution buffer and column + collection tube assembly in a heat block or incubator warmed to  50 °C for  00:10:00 .

13.2 Spin at  10000 x g, 37°C, 00:05:00 to elute RNA from the column. The flowthrough present in the collection tube contains the purified RNA.

Storage

- 14 The eluted RNA is now ready for downstream analysis. Store RNA at  4 °C for same-day use or freeze at  -80 °C for later use and storage.