

May 15, 2019

# **©** UC Davis - Temperature and activity by Telemetry

DOI

dx.doi.org/10.17504/protocols.io.ywtfxen



### John Rutledge<sup>1</sup>

<sup>1</sup>University of California, Davis

Mouse Metabolic Phenotyping Centers Tech. support email: info@mmpc.org



Lili Liang

# Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account





DOI: https://dx.doi.org/10.17504/protocols.io.ywtfxen

External link: <a href="https://mmpc.org/shared/document.aspx?id=282&docType=Protocol">https://mmpc.org/shared/document.aspx?id=282&docType=Protocol</a>

Protocol Citation: John Rutledge 2019. UC Davis - Temperature and activity by Telemetry. protocols.io

https://dx.doi.org/10.17504/protocols.io.ywtfxen



License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: March 07, 2019

Last Modified: May 15, 2019

Protocol Integer ID: 21171

Keywords: Temperature and activity by Telemetry, comprehensive assessment of circadian pattern, circadian pattern, core body temperature in multiple intervention, core body temperature, activity by telemetry summary, genetic mouse model, temperature, telemetry summary, physiol genomic, mouse model, telemeter, activity measure, single mouse

#### Abstract

#### Summary

Utilizing the TA-F10 (DSI) telemeter we can simultaneously collect temperature and activity measures in a single mouse. This can provide a comprehensive assessment of circadian patterns, activity levels, and core body temperature in multiple interventions (drug, altered fed access, etc.) or genetic mouse models. The following protocol describes the implantation and measurement parameters and intervals.

**Modified from:** Butz et al. Physiol Genomics. 2001 Mar 8;5(2):89-97.



### **Materials**

#### **MATERIALS**

- X Isoflurane
- x sterile cotton swabs
- 205016 9mm AutoClips (wound clips) Braintree Scientific Catalog #205016
- MikRon 9mm AuoClip Applicator Braintree Scientific Catalog #ACS APL
- Silk ligatures Braintree Scientific
- ₩ Vannas spring micro-scissors Fine Science Tools Catalog #15610-08
- annulation forceps Fine Science Tools Catalog #00608-11
- blunt dissecting Fine Science Tools Catalog #14018-14
- **X** veterinary adhesive
- Buprenex (buprenorphine hydrochloride)
- Dataguest ART data acquisition system Data Scientific International
- X TA-F10 telemeter Data Scientific International Catalog #TA-F10

### **Troubleshooting**

# Safety warnings



### **WARNING**

All blood components and biological materials should be handled as potentially hazardous. Follow universal precautions established by CDC when handling and disposing of infectious agents.



- 1 Mice are anesthetized with isoflurane (2.5%) and the flank/back was shaved and disinfected.
- 2 Implantation of transmitter subcutaneously along the right flank.
  - a. Through the same ventral neck incision, a subcutaneous pouch is formed for placement of the transmitter body along the animal's right flank.
  - b. Using a pair of blunt dissecting scissors (Fine Science Tools, catalog no. 14018-14), the skin is gently dissected free from underlying tissue starting at the right neck region and proceeding posteriorly to form a "pocket" along the right flank. It is important that the pocket be made sufficiently large to house the transmitter without unduly stretching the skin, because pressure necrosis could result.
  - c. Using a 1mL syringle with warmed saline, insert tip of syringe into the pocket for lubrication prior to inserting the transmitter.
  - d. The transmitter is slipped under the skin and down into the pocket along the flank as close to the right hindlimb as possible.
  - e. A small drop of veterinary adhesive is placed on the catheter in the right neck region to further secure the device.
  - f. The neck incision is closed using 1 or 2 wound clips and further sealed with tissue adhesive.
  - g. Mice are kept warm on a heating pad and monitored closely until fully recovered from anesthesia.
- Implantation of transmitter on the back. Alternatively, the transmitter body can be installed on the back at the midscapular region.
  - a. The upper back is shaved, and a horizontal incision was made.
  - b. The transmitter is placed in the opening and sutured to both the muscle and the skin (with the suture loop around the probe) via the suture rib on the probe as described.
  - c. A separate ventral neck incision is then made, the catheter is tunneled subcutaneously to the neck, and the catheterization procedure is carried out as described.
  - d. Mice are observed postoperatively as above.
- Buprenex (buprenorphine hydrochloride) 0.1 mg/kg will be administered subcutaneously at the termination of surgery to maintain analgesia.



- 5 Following full recovery from anesthesia, mice are returned to their home cages (placed atop telemetry receivers), where they continued to be monitored daily throughout the study for general condition, body weight, food and water intake, state of surgical wound healing, and any signs of morbidity.
  - a. Radiotelemeters are magnetically activated immediately upon return to the home cage, and temperature and locomotor activity are recorded continuously.
- 6 Radiotelemetry data are collected continuously (sampling every 5 min for 10-s intervals) and stored using the Dataquest ART data acquisition system (Data Sciences International).
  - a. MAP and HR data collected from the first 12 consecutive days following surgery were plotted as mean values over each hour.