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 We use this protocol and it's working

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 21107

UC Davis - Non-Esterified Fatty Acids Protocol

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ABSTRACT

Summary:

The Wako enzymatic method relies upon the acylation of coenzyme A (CoA) by the fatty acids in the presence of added acyl-CoA synthetase (ACS). The acyl-CoA thus produced is oxidized by added acyl-CoA oxidase (ACOD) with generation of hydrogen peroxide, in the presence of peroxidase (POD) permits the oxidative condensation of 3-methy-N-ethyl-N(β-hydroxyethyl)-aniline (MEFA) with 4-aminoantipyrine to form a purple colored adduct which can be measured colorimetrically at 550 nm.

MATERIALS

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Calibrator **FUJIFILM Wako Pure Chemical Corporation Catalog #276-76491**

Reagents(999-34691 995-34791 991-34891 993-35191) **FUJIFILM Wako Diagnostic U.S.A. Catalog #999-34691,995-34791,991-34891,**

Microplate **Contributed by users**

Platereader **Contributed by users**

Reagent Preparation:

Reagent A – reconstitute Color Reagent A with Solvent A

Reagent B – reconstitute Color Reagent B with Solvent B

Note:

FUJIFILM Wako [RRID:SCR_013651](#)

Keywords: Non-Esterified
Fatty Acids, Wako enzymatic
method

1 Reconstitute Color Reagent A with 50 ml of Solvent A and Color Reagent B with Solvent B.

2 Add 5 µl of calibrator and sample to each well.

3 Add 200 µl of Reagent A to each well. Incubate at 37°C for 5 minutes. Read at 560 nm.

IMPORTANT: Make sure not to add any bubbles to the wells when dispensing reagents, this will interfere with reading in the platereader.

4 Add 100 µl of Reagent B to each well. Incubate at 37°C for 5 minutes. Read at 560 nm.

5 Subtract blank readings from final readings. The assay will be linear so the unknown samples can be calculated as $(\text{Sample Absorbance} \div \text{Calibrator Absorbance}) \times \text{Calibrator Concentration}$.