

May 14, 2019

UC Davis - HbA1c Protocol

DOI

dx.doi.org/10.17504/protocols.io.ynrfvd6



Peter Havel¹

¹University of California, Davis

Mouse Metabolic Phenotyping Centers
Tech. support email: info@mmpc.org



Lili Liang

OPEN  ACCESS



DOI: dx.doi.org/10.17504/protocols.io.ynrfvd6

External link: <https://mmpc.org/shared/document.aspx?id=89&docType=Protocol>

Protocol Citation: Peter Havel 2019. UC Davis - HbA1c Protocol. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.ynrfvd6>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: February 27, 2019

Last Modified: May 14, 2019

Protocol Integer ID: 20913

Keywords: Enzymatic HbA1c test,



Abstract

Summary:

Direct Enzymatic HbA1c test is an enzymatic assay in which lysed whole blood samples are subjected to extensive protease digestion with *Bacillus* sp protease. This process releases amino acids including glycated valines from the hemoglobin beta chains. Glycated valines then serve as substrates for specific recombinant fructosyl valine oxidase (FVO) enzyme, produced in *E. coli*. The recombinant FVO specifically cleaves N-terminal valines and produces hydrogen peroxide. This, in turn, is measured using a horseradish per-oxidase (POD) catalyzed reaction and a suitable chromagen.

The HbA1c concentration is expressed directly as %HbA1c by use of a suit-able calibration curve in which the calibrators have values for each level in %HbA1c.

Materials

MATERIALS

✕ Calibrator **Diazyme Catalog #DZ168A-CAL**

✕ Reagents **Diazyme Catalog #DZ168A-K**

✕ Microplate

✕ Platereader

Reagent Preparation:

Lysis Buffer – ready to use

R1a & R1b – mix together in 70:30 ratio

R2 – ready to use

- 1 Use 250 µl of lysis buffer to lyse 20 µl samples of whole blood and calibrators.

IMPORTANT: Make sure the samples are totally lysed. Any solid material floating around will interfere with reading in the platereader.

- 2 Mix R1a and R1b reagents together in a 70:30 ratio.

- 3 Add 25 µl of each calibrator and sample to each well.

- 4 Add 160 µl of reagent R1ab mix to each well. Incubate at 37°C for 5 minutes then read at 720 nm.

IMPORTANT: Make sure not to add any bubbles to the wells when dispensing reagents, this will interfere with reading in the platereader.

- 5 Add 70 µl of R2 to each well. Incubate at 37°C for 3 minutes then read at 720 nm.

- 6 Subtract blank readings from final readings. The assay will be linear so the unknown samples can be calculated as (Sample Absorbance ÷ Calibrator Absorbance) × Calibrator Concentration.