

Nov 02, 2023

🌐 Two-step method for isolation of inactivated CD4+ T-cells from human blood mononuclear cells

DOI

dx.doi.org/10.17504/protocols.io.36wgq3mxolk5/v1



Anna Esman¹, Michael Vinokurov¹

¹Central Research Institute of Epidemiology

Anna Esman: <https://www.researchgate.net/profile/Anna-Esman>

Michael Vinokurov: <https://www.researchgate.net/profile/Michael-Vinokurov-2>



Anna Esman

Central Research Institute of Epidemiology

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.36wgq3mxolk5/v1>

Protocol Citation: Anna Esman, Michael Vinokurov 2023. Two-step method for isolation of inactivated CD4+ T-cells from human blood mononuclear cells. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.36wgq3mxolk5/v1>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: October 24, 2023

Last Modified: November 02, 2023

Protocol Integer ID: 89818

Keywords: CD4+ cells , magnetic separation , untouched human CD4+ T cells, CD4+ inactivated cells, CD4+ T-cells, cells from human blood mononuclear cell, human blood mononuclear cell, inactivated cell, inactivated cd4, obtaining human cd4, human cd4, magnetic tube separator, step method for isolation, obtaining cd4, cell

Funders Acknowledgements:







Effect of DNA methylation in HIV-1 positive individuals on viral reservoir reactivation

Grant ID: 122053000056-2

Abstract

1. Obtaining human CD4+ T cells
2. Obtaining CD4+ inactivated cells







using

1.  Dynabeads™ Untouched™ Human CD4 T Cells Kit **ThermoFisher Catalog #11346D**
2.  CELLection™ Biotin Binder Kit **ThermoFisher Catalog #11533D**
3.  CD71 (Transferrin Receptor) Monoclonal Antibody (OKT9 (OKT-9)) **eBioscience Catalog #14-0719-82**
4.  CD25 Monoclonal Antibody (BC96), Biotin, eBioscience **eBioscience Catalog #13-0259-82**
5.  HLA-DR Monoclonal Antibody (LN3), Biotin **eBioscience Catalog #13-9956-82**
6.  CD69 Monoclonal Antibody (FN50), Biotin **eBioscience Catalog #13-0699-82**

and magnetic tube separator.



Materials

1.  Dynabeads™ Untouched™ Human CD4 T Cells Kit **Thermofisher Catalog #11346D**
2.  CELLection™ Biotin Binder Kit **Thermofisher Catalog #11533D**
3.  CD71 (Transferrin Receptor) Monoclonal Antibody (OKT9 (OKT-9)) **eBioscience Catalog #14-0719-82**
4.  CD25 Monoclonal Antibody (BC96), Biotin, eBioscience **eBioscience Catalog #13-0259-82**
5.  HLA-DR Monoclonal Antibody (LN3), Biotin **eBioscience Catalog #13-9956-82**
6.  CD69 Monoclonal Antibody (FN50), Biotin **eBioscience Catalog #13-0699-82**
7. Magnetic separator for 1.5 / 5 / 15 / 50 ml tubes
8. Mixer with tilt and rotation capabilities
9. PBS (Ca 2+ and Mg 2+ free)  PBS, pH 7.4 **Thermo Fisher Catalog #10010023**
10. 0.1% BSA
11.  2 millimolar (mM) EDTA
12.  Fetal Bovine Serum **Gibco - Thermo Fisher Scientific Catalog #26140079**
13.  RPMI-1640 **Pan-Eco Catalog #C310p**

.



Protocol materials

⊗ PBS, pH 7.4 **Thermo Fisher Catalog #10010023**

⊗ Dynabeads™ Untouched™ Human CD4 T Cells Kit **Thermofisher Catalog #11346D**

⊗ CD25 Monoclonal Antibody (BC96), Biotin, eBioscience **eBioscience Catalog #13-0259-82**

⊗ CD69 Monoclonal Antibody (FN50), Biotin **eBioscience Catalog #13-0699-82**

⊗ PBS, pH 7.4 **Thermo Fisher Catalog #10010023**

⊗ Fetal Bovine Serum **Gibco - Thermo Fisher Scientific Catalog #26140079**

⊗ Dynabeads™ Untouched™ Human CD4 T Cells Kit **Thermofisher Catalog #11346D**

⊗ CELLection™ Biotin Binder Kit **Thermofisher Catalog #11533D**

⊗ CD71 (Transferrin Receptor) Monoclonal Antibody (OKT9 (OKT-9)) **eBioscience Catalog #14-0719-82**

⊗ RPMI-1640 **Pan-Eco Catalog #C310p**

⊗ HLA-DR Monoclonal Antibody (LN3), Biotin **eBioscience Catalog #13-9956-82**

⊗ PBS, pH 7.4 **Thermo Fisher Catalog #10010023**

⊗ PBS, pH 7.4 **Thermo Fisher Catalog #10010023**

⊗ CD71 (Transferrin Receptor) Monoclonal Antibody (OKT9 (OKT-9)) **eBioscience Catalog #14-0719-82**

⊗ CD25 Monoclonal Antibody (BC96), Biotin, eBioscience **eBioscience Catalog #13-0259-82**

⊗ HLA-DR Monoclonal Antibody (LN3), Biotin **eBioscience Catalog #13-9956-82**

⊗ CD69 Monoclonal Antibody (FN50), Biotin **eBioscience Catalog #13-0699-82**

⊗ Dynabeads™ Untouched™ Human CD4 T Cells Kit **Thermofisher Catalog #11346D**

⊗ Dynabeads™ Untouched™ Human CD4 T Cells Kit **Thermofisher Catalog #11346D**

⊗ BD Vacutainer® CPT™ Mononuclear Cell Preparation Tube - Sodium Heparin **BD Biosciences Catalog #362753**

⊗ PBS, pH 7.4 **Thermo Fisher Catalog #10010023**

⊗ HLA-DR Monoclonal Antibody (LN3), Biotin **eBioscience Catalog #13-9956-82**

⊗ CD69 Monoclonal Antibody (FN50), Biotin **eBioscience Catalog #13-0699-82**

⊗ Dynabeads™ Untouched™ Human CD4 T Cells Kit **Thermofisher Catalog #11346D**

⊗ CELLection™ Biotin Binder Kit **Thermofisher Catalog #11533D**


⊗ CD71 (Transferrin Receptor) Monoclonal Antibody (OKT9 (OKT-9)) **eBioscience Catalog #14-0719-82**


⊗ CD25 Monoclonal Antibody (BC96), Biotin, eBioscience **eBioscience Catalog #13-0259-82**

⊗ BD Vacutainer® CPT™ Mononuclear Cell Preparation Tube - Sodium Heparin **BD Biosciences Catalog #362753**

Troubleshooting

Before start

 Sample


 BD Vacutainer® CPT™ Mononuclear Cell Preparation Tube - Sodium Heparin **BD Biosciences Catalog #362753**

•




Negative depletion of CD4+ T-cells

1 **Preparation of PBMC** (peripheral blood mononuclear cells)

1.1 Collect at least  5 mL of human whole blood to the



BD Vacutainer® CPT™ Mononuclear Cell Preparation Tube - Sodium Heparin **BD Biosciences Catalog #362753**

1.2 Store tube upright at  Room temperature until centrifugation.

Note

To ensure proper barrier formation, blood samples should be centrifuged within 2 hours of blood collection. Centrifugation more than 2 hours after specimen collection may cause incomplete barrier formation.

1.3  1500-1800 rcf, 18-25°C in a horizontal rotor (swing-out head) for a minimum of




00:15:00

15m

1.4 Aspirate approximately half of the plasma without disturbing the cell layer.

Note

Mononuclear cells and platelets will be in a whitish layer just under the plasma layer

1.5 Collect cell layer with a Pasteur Pipette and transfer to a  15 mL size conical centrifuge tube with cap.

Note

Collection of cells immediately following centrifugation will yield best results



Note

An alternative procedure for recovering the separated mononuclear cells is to resuspend the cells into the plasma by inverting the unopened BD Vacutainer CPT Tube gently 5 to 10 times. This is the preferred method for storing or transporting the separated sample for up to 24 hours after centrifugation. To collect the cells, open the BD Vacutainer CPT Tube and pipette the entire contents of the tube above the gel into a separate vessel.

Expected result




Separation of PBMC from whole blood.

2 Preparation of the *Isolation buffer*

 PBS, pH 7.4 Thermo Fisher Catalog #10010023 supplemented with 0.1% BSA and  2 millimolar (mM) EDTA.

3 Preparation of magnetic particles

3.1 Resuspend the

 Dynabeads™ Untouched™ Human CD4 T Cells Kit Thermofisher Catalog #11346D in the vial (i.e. vortex for >  00:00:30 or tilt and rotate for  00:05:00)

5m 30s

3.2 Transfer the desired volume of

 Dynabeads™ Untouched™ Human CD4 T Cells Kit Thermofisher Catalog #11346D to a tube.

3.3 Add the same volume of *Isolation Buffer*, or at least 1 mL and resuspend.

3.4 Place the tube in a magnet for 00:01:00 and discard the supernatant.

1m


3.5 Remove the tube from the magnet and resuspend the washed



 Dynabeads™ Untouched™ Human CD4 T Cells Kit Thermofisher Catalog #11346D



in the same volume of Isolation Buffer as the initial volume of Dynabeads®.

4 Isolation Procedure



4.1 Transfer  100 µL (5×10^7) PBMC in *Isolation Buffer* to a tube.

4.2 Add  20 µL  PBS, pH 7.4 Thermo Fisher Catalog #10010023



4.3 Add  20 µL of Antibody Mix

Note

Contains mouse IgG antibodies towards human CD8, CD14, CD16 (specific for CD16a and CD16b), CD19, CD36, CD56, CDw123 and CD235a (Glycophorin A)

4.4 Mix well and incubate for  00:20:00 at  2-8 °C



20m

4.5 Wash the cells by adding  2 mL *Isolation Buffer*. Mix well by tilting the tube several times and  350 x g, 2-8°C, 00:08:00 . Discard the supernatant.

8m

4.6 Resuspend the cells in  100 µL *Isolation Buffer*.

4.7 Add  100 µL pre-washed Dynabeads.

4.8 Incubate for  00:15:00 at  Room temperature with gentle tilting and rotation

15m


4.9 Add  1 mL *Isolation Buffer*.

Note


When working with lower cell volumes, never use less than 1 mL *Isolation Buffer*




4.10 Resuspend the bead-bound cells thoroughly by pipetting >10 times using a pipette with a narrow tip opening. Avoid foaming.

4.11 Place the tube in the magnet for  00:02:00 . Transfer the supernatant containing the untouched human CD4+ T cells, to a new larger tube.


2m

4.12 Add  2 mL *Isolation Buffer* to the tube containing the Dynabeads and resuspend the bead-bound cells by pipetting as described in step 4.10.

4.13 Place the tube in the magnet for  00:02:00 and then combine the two supernatants.

2m

Note

To remove residual beads; place the tube in the magnet for  00:02:00 and transfer cells to a new tube.





Expected result

The supernatant contains negatively isolated human CD4+ T-cells.


Negative depletion of inactivated CD4+ T-cells

46m 30s

5 Preparing buffers for operations

- *Buffer 1:*  PBS, pH 7.4 Thermo Fisher Catalog #10010023 supplemented with 0.1% bovine serum albumin (BSA),  7.4
- *Buffer 2:*  PBS, pH 7.4 Thermo Fisher Catalog #10010023 with 0.1% BSA and 0.6% sodium citrate or  2 millimolar (mM) EDTA.

6 Prepare Release Buffer

1. For each vial of freeze-dried DNase I, transfer  300 μ L from the Releasing Buffer Component II to each tube of Releasing Buffer Component I (DNase I).

**Note**

Dissolve the enzyme gently. Vigorous mixing will destroy the enzyme.

2. Aliquot the reconstituted Release Buffer into suitable portions. Store at -20°C . Thaw immediately before use and keep on ice once thawed. Thawed Release Buffer can be re-frozen once.

7 Wash Dynabeads

7.1 Resuspend the Dynabeads® in the vial (i.e. vortex for > 00:00:30 or tilt and rotate for 00:05:00. Transfer the desired volume of Dynabeads® to a tube (25 μL for one sample). 5m 30s

7.2 Add the same volume of *Buffer 1*, or at least 1 mL and resuspend.

7.3 Place the tube in a magnet for 00:01:00 and discard the supernatant. 1m

7.4 Remove the tube from the magnet and resuspend the washed Dynabeads® in the same volume of *Buffer 1* as the initial volume of Dynabeads®.

8 Isolate Cells (Labeling Cells with Biotinylated Antibodies)

8.1 Add ~ 10 μg primary biotinylated antibody to 1 mL cell suspension and mix

⊗ CD71 (Transferrin Receptor) Monoclonal Antibody (OKT9 (OKT-9)) **eBioscience Catalog #14-0719-82**



⊗ CD25 Monoclonal Antibody (BC96), Biotin, **eBioscience Catalog #13-0259-82**

⊗ HLA-DR Monoclonal Antibody (LN3), Biotin **eBioscience Catalog #13-9956-82**

⊗ CD69 Monoclonal Antibody (FN50), Biotin **eBioscience Catalog #13-0699-82**


8.2 Incubate for 00:10:00 at $2-8^{\circ}\text{C}$ 10m



8.3 Wash the cells by adding  2 mL *Buffer 2* and  350 x g, 00:08:00 . Discard the supernatant.


8m

8.4 Suspend the cells in  4 mL *Buffer 2*.


8.5 Add  25 µL pre-washed and resuspended Dynabeads

8.6 Incubate for  00:20:00 at  2-8 °C with gentle tilting and rotation.

20m

8.7 Place the tube in the magnet for  00:02:00 . Transfer the supernatant containing the inactivated human CD4+ T-cells to a new larger tube.

2m

8.8 Add  4 mL *Buffer 2* to the tube containing the Dynabeads and repeat step 8.7

8.9 Combine the two supernatants.

Expected result

The resulting supernatant contains the inactivated human CD4+ T cells.

Protocol references

[Dynabeads™ Untouched™ Human CD4 T Cells Kit](#)

[CELlection™ Biotin Binder Kit](#)