

# True-Nuclear™ Transcription Factor Staining Protocol for 5 mL Tubes V.4

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## Guidelines

### General Tips and FAQ:

*What are some improvements in the True-Nuclear™ Buffer compared to the Foxp3 Fix/Perm (Cat. No. 421403) and the Nuclear Factor Fixation and Permeabilization Buffer Sets (Cat. No. 422601)?*

- Less background for nuclear staining (e.g.- Foxp3) staining, offering improved signal/noise ratio for better population distinction
- Minimal effects on surface staining, especially those stained using tandem fluorophores such as APC/FIRE™ 750

*I am observing high background staining. What can I do to reduce it?*

- Longer fixation time – as noted in the fixation step, a longer fixation period can help ameliorate high background

*Is the True-Nuclear™ Buffer set recommended for intracellular cytokine staining?*


- No: The permeabilization conditions required for staining transcription factors using True-Nuclear™ (or Foxp3 Fix/Perm and Nuclear Factor Fixation and Permeabilization Buffer Sets) is much harsher than those recommended for staining cytokines (i.e.- cells become more porous). This can lead to cytokine leakage upon permeabilization, resulting in loss of signal. For staining intracellular cytokines, we recommend using the **Fixation (Cat. No. 420801)** and **Permeabilization Wash Buffers (Cat. No. 421002)** and following our intracellular staining protocol, which can be found [here](#).


*Any known nuclear targets not suited for detection using the True-Nuclear™ buffer?*

- Ki-67: You can find our recommended protocol for Ki-67 staining [here](#).

## Materials

### MATERIALS

 Cell Staining Buffer **BioLegend Catalog #420201**

 True-Nuclear™ Transcription Factor Buffer Set **BioLegend Catalog #424401**

### STEP MATERIALS

 Cell Staining Buffer **BioLegend Catalog # 420201**



- 1 Perform cell surface staining as described in BioLegend's Cell Surface Immunofluorescence Staining Protocol.
- 2 Add 1mL of the True-Nuclear™ 1X Fix Concentrate to each tube, vortex and incubate at room temperature in the dark for 45-60 minutes. If necessary, the protocol can be suspended at this point. After discarding supernatant, re-suspend cells in CytoLast™ Buffer (Cat. No. [422501](#)) or equivalent. Samples can be stored at 4°C for 12-18 hours, protected from light and plastic-wrapped to protect buffer evaporation.
- 3 Without washing, add 2mL of the True-Nuclear™ 1X Perm Buffer to each tube.
- 4 Centrifuge tubes at 300-400xg at room temperature for 5 minutes, and discard the supernatant.
- 5 Add 2mL of the True-Nuclear™ 1X Perm Buffer to each tube.
- 6 Centrifuge tubes at 300-400xg at room temperature for 5 minutes, and discard the supernatant.
- 7 Resuspend the cell pellet in 100µL of the True-Nuclear™ 1X Perm Buffer.
- 8 Add the appropriate amount of fluorochrome conjugated antibody diluted in True-Nuclear™ 1X Perm Buffer for detection of intracellular antigen(s) to each well and incubate in the dark at room temperature for at least 30 minutes.
- 9 Add 2mL of the True-Nuclear™ 1X Perm Buffer to each tube.
- 10 Centrifuge tubes at 300-400 x g at room temperature for 5 minutes, and discard the supernatant.
- 11 Add 2mL of cell staining buffer (Cat. No. [420201](#)).
- 12 Centrifuge tubes at 300-400xg at room temperature for 5 minutes, and discard the supernatant.
- 13 Resuspend in 0.5mL cell staining buffer then acquire the tubes on a flow cytometer.