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Transcreener® pADPr PARP FP Assay Technical Manual

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We use this protocol and it's working

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Abstract

The Transcreener® pADPr PARP FP Assay is a biochemical HTS assay for measuring the production of poly(ADP-ribose) (pADPr) in poly(ADP-ribose) polymerase (PARP) reactions. The assay relies on pADPr Coupling Enzymes (CE) to convert pADPr into AMP, which is then detected using a far-red, competitive fluorescence polarization (FP) assay. As an example, the Transcreener pADPr PARP FP assay can be used to detect the activity of human PARP1, which uses NAD+ to form pADPr on itself, histones, and DNA repair proteins.

The Transcreener pADPr PARP FP assay is designed specifically for high throughput screening (HTS), with a single-addition, mix-and-read format. It is easy to integrate into automated HTS workflows, with outstanding reagent stability (deck stability > 16 hours, signal stability > 24 hours), robust detection of ADPR monomers released from pADPr over a range of 10 nM to 100 µM, and compatibility with commonly used multimode plate readers. Data quality is excellent (Z′ ≥ 0.7), and the assay uses a far-red tracer to minimize interference from fluorescent compounds and light scattering.

Introduction

1 The Transcreener® pADPr PARP FP Assay is a biochemical HTS assay for measuring the production of poly(ADP-ribose) (pADPr) in poly(ADP-ribose) polymerase (PARP) reactions (**Figure 1**). The assay relies on pADPr Coupling Enzymes (CE) to convert pADPr into AMP, which is then detected using a far-red, competitive fluorescence polarization (FP) assay. As an example, the Transcreener pADPr PARP FP assay can be used to detect the activity of human PARP1, which uses NAD+ to form pADPr on itself, histones, and DNA repair proteins.

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Key Applications:

- Screening for enzyme inhibitors/activators
- Generating dose response curves and IC_{50} values for inhibitors.
- Kinetic and mechanistic analyses.

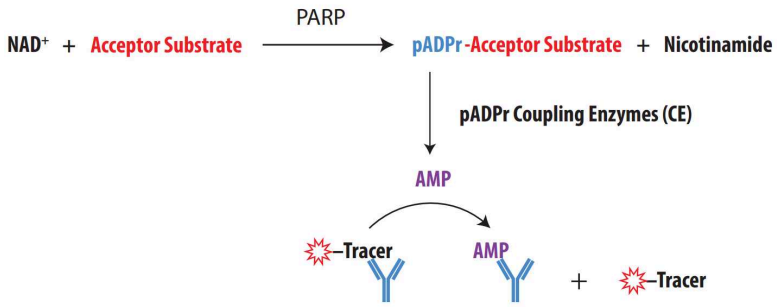


Figure 1. Schematic overview of the Transcreener pADPr PARP FP Assay. pADPr produced by the target PARP enzyme is converted to AMP in real time by the pADPr Coupling Enzymes. In the detection step, the PARP Enzyme and CE are quenched by EDTA and AMP displaces an Alexa Fluor® 633 tracer from the AMP^2/GMP^2 antibody, resulting in decreased fluorescence polarization.

Product Specifications

2

	Product	Quantity	Part #
	Transcreener® pADPr PARP FP Assay	1,000 assays*	3043-1K
		10,000 assays*	3043-10K

*The exact number of assays depends on the enzyme reaction conditions. The kits are designed for use with 384-well plates, using a 10 µL Enzyme Reaction and a 20 µL Complete Assay Volume.

IMPORTANT: Antibody centrifugation is required to remove aggregates that can disrupt data quality. Antibodies should be centrifuged at 10,000 x g for 10 minutes before use. Following centrifugation, pipet the solution needed from the top of the aliquot to ensure precipitate is not present in the detection reagents.

Storage

The pADPr Coupling Enzymes (CE) should be stored at -80°C; other reagents can be stored at –20°C. Though we have confirmed that the CE is stable up to 5 freeze-thaw cycles, we recommend aliquoting the reagents and snap-freezing for multiple uses to minimize loss of activity.

Use the reagents provided in this kit within 6 months from date of receipt.

Materials Provided

2.1

	Component	B	C
	AMP^2/GMP^2 Antibody	1.26 mg/mL solution in PBS with 10% glycerol*	Sufficient antibody is included in the kit to complete 1,000 assays (Part # 3043-1K) or 10,000 assays (Part # 3043-10K).
	AMP^2/GMP^2 Alexa Fluor® 633 Tracer	800 nM solution in 2 mM HEPES (pH 7.5) containing 0.01% Brij-35	The final tracer concentration in the 20 µL Complete Assay is 4 nM. Sufficient tracer is included in the kit to complete 1,000 assays (Part # 3043-1K) or 10,000 assays (Part # 3043-10K).
	pADPr Coupling Enzymes (CE)	400X CE in 50 mM Tris-HCl (pH 7.5), 100 mM NaCl, 1 mM TCEP, 10% glycerol, 0.05% triton	Sufficient CE for 1,000 assays (Part # 3043-1K) or 10,000 assays (Part # 3043-10K) with coupling enzyme present in excess to ensure pADPr is completely converted to AMP.
	Stop & Detect Buffer B, 10X	200mM HEPES (pH 7.5), 400 mM EDTA, and 0.2% Brij-35	The Stop & Detect Buffer B components quench the PARP Enzyme and CE reactions by chelating Mg^{2+}. Therefore, it will work for any target enzyme, as long as the EDTA concentration is at least equimolar to the Mg^{2+}.
	NAD^+, 5 mM	5 mM in H_2O	Sufficient NAD^+ is included in the kit to complete 1,000 assays (Part # 3043-1K) or 10,000 assays (Part # 3043-10K).
	AMP, 5 mM	5 mM in H_2O	The AMP in this kit can be used to create a standard curve to convert mP values to AMP product formed.

*The exact concentration may vary from batch to batch. Please refer to the Certificate of Analysis for an accurate concentration.

Materials Required But Not Provided

2.2

	Component	Notes
	Ultrapure Nuclease Free Water	Some deionized water systems are contaminated with enzymes that can degrade both nucleotide substrates and products, reducing assay performance. Use nuclease free water such as: Invitrogen Part # AM9930

	Component	Notes
	Enzyme	The Transcreener® pADPr PARP FP Assay is designed for use with purified PARP enzymes. It is not recommended for cell lysates or impure enzyme preparations as contaminating enzymes, such as phosphatases or nucleotidases, can produce background signal and reduce the assay window.
	Acceptor Substrate	PARP1 can ADP-ribosylate itself and does not require addition of an additional acceptor substrate. If a separate acceptor substrate is used, it should be highly purified to avoid degradation of AMP by contaminating enzymes.
	Plate Reader	A multimode microplate reader configured to measure FP is required. Transcreener Assays have been validated on the following instruments: BioTek Synergy™2 and Synergy™4; BMG Labtech PHERAstar® Plus and CLARIOstar® Plus; Molecular Devices SpectraMax™ Paradigm; Perkin Elmer EnVision® and ViewLux; and Tecan Infinite® F500, Safire2™, and M1000. Full list of compatible plate readers and settings.
	Liquid Handling Devices	Use liquid handling devices that can accurately dispense submicroliter volumes into 384-well plates.
	Assay Plates	It is important to use assay plates that are entirely black with a nonbinding surface. We recommend Corning® 384-well plates (Cat. # 4514). The suggested plate has a square well top that enables easier robotic pipetting and a round bottom that allows good Z′ factors. It has a recommended working volume of 15–20 µL.

Note

Note: Contact BellBrook Labs Technical Service for suppliers and catalog numbers for buffer components, and additional information regarding setup of FP instruments.

Before You Begin

3 1. Read the entire protocol and note any reagents or equipment needed (see **Section 2.2**).
2. Check the plate reader and verify that it is compatible with the Transcreener AMP^2/GMP^2 Assays. **Full list of compatible plate readers and settings.**

Protocol

4 The Transcreener pADPr PARP FP Assay kit optimization follows a simple protocol. First, the instrument parameters are configured to ensure optimal signal detection (**Section 4.1**). Then, the Transcreener AMP^2/GMP^2 antibody concentration is selected for the desired dynamic range (**Section 4.2**). Next, an optimal enzyme concentration is determined to provide a good assay window while meeting initial velocity requirement (**Section 4.3**). Once the instrument parameters, antibody, and enzyme optimization are complete, the assay itself consists of a simple mix-and-read protocol (**Section 4.4**).

The assay procedure was developed for a 384-well format, using a 10 µL Enzyme Reaction and 20 µL Complete Assay volume when the plates are read (**Figure 2**). The use of different densities or reaction volumes will require changes in reagent quantities (see

Section 7.3 for example reaction volumes).

Note: The tracer concentration remains constant at 4 nM in the 20 µL Complete Assay regardless of changes to other reaction conditions.

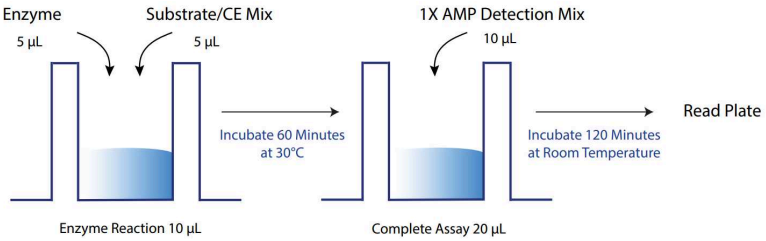


Figure 2. An outline of the Assay Procedure. The target Enzyme Reaction is run in the presence of CE, so that pADPr is converted to AMP in real time. After the Enzyme Reaction incubation is complete, AMP detecting reagents (Transcreener AMP^2/GMP^2 Antibody and Tracer) are added along with EDTA to quench the Enzyme Reaction.

Set Up the Instrument

4.1 Becoming familiar with ideal instrument settings for FP is essential to the success of the Transcreener®
pADPr PARP FP Assay.

4.1.1 Verify That the Instrument Measures FP

Ensure that the instrument is capable of measuring FP (not simply fluorescence intensity) of the AMP^2/GMP^2 AlexaFluor 633 Tracer.

Note

Note: A complete list of instruments and instrument-specific application notes can be found online at:
https://www.bellbrooklabs.com/technicalresources/instrument-compatibility

Contact BellBrook Labs Technical Service if you have questions about settings and filter sets for a specific instrument.

4.1.2 Define the Maximum mP Window for the Instrument

Measuring high (tracer + antibody) and low (free tracer) FP will define the maximum assay window of your specific instrument. Prepare High and Low FP Mixtures in quantities sufficient to perform at least 6 replicates for each condition.

Use AMP^2/GMP^2 Alexa Fluor® 633 Tracer at 4 nM in a 20 µL Complete Assay. This mimics the 2-fold dilution when adding an equal volume of detection mix to an Enzyme Reaction.

High FP Mixture

	Component	As Provided	Final Concentration in 20 µL Complete Assay	Example: 25 Assays	Your Numbers
	AMP^2/GMP^2 Antibody	1.26 mg/mL*	18 µg/mL	8.6 µL**	

	Component	As Provided	Final Concentration in 20 µL Complete Assay	Example: 25 Assays	Your Numbers
	AMP^2/GMP^2 Alexa Fluor 633 Tracer	800 nM	4 nM	3 µL	
	Stop & Detect Buffer B	10X	0.5X	30 µL	
	Water			558.4 µL	
	Total			**600.0 µL**	

*Please note AMP^2/GMP^2 Antibody concentration varies by lot number. This is an example and should be adjusted based on stock concentration accordingly.
**Pipetting small sample volumes accurately requires the correct equipment and proper technique. An extra dilution step may be required to ensure accuracy.

Low FP Mixture

	Component	As Provided	Final Concentration in 20 µL Complete Assay	Example: 25 Assays	Your Numbers
	AMP^2/GMP^2 Alexa Fluor 633 Tracer	800 nM	4 nM	3 µL	
	Stop & Detect Buffer B	10X	0.5X	30 µL	
	Water			567 µL	
	Total			**600.0 µL**	

4.1.3 Measure the FP
Subtract the Low FP Mixture readings from the corresponding High FP Mixture readings. The difference between the low and high FP values should be >100 mP.

Note

Caution: Contact BellBrook Labs Technical Service for assistance if the assay window is <100 mP.

Calculate the Optimal AMP^2/GMP^2 Antibody Concentration

4.2 The sensitivity of the Transcreener pADPr PARP FP Assay, and thus the dynamic range, is determined by the concentration of the Transcreener AMP^2/GMP^2 antibody: a lower concentration of antibody increases the sensitivity, higher antibody results in lower sensitivity. Assuming that PARP reactions will be run under initial velocity conditions (≤ 10% consumption of NAD^+), as is typical for almost all applications, the concentration of NAD^+ used establishes the desired dynamic range. For example, if 100 µM NAD^+ is used in the PARP reaction, then the desired dynamic range is 1-10 µM AMP. The optimal Transcreener AMP^2/GMP^2 Antibody concentration for the 1X AMP Detection Mix can be calculated based on the concentration of NAD^+ in the Enzyme Reaction:

[Ab] (µg/mL) = 0.3274 x [NAD+] (µM) + 4.24

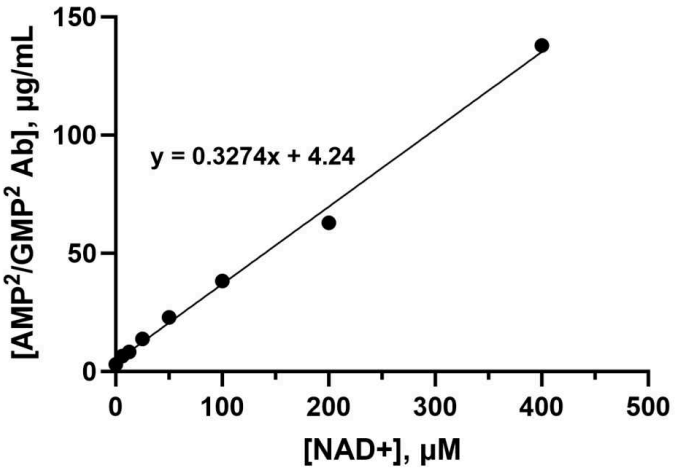


Figure 3. Relationship between NAD^+ concentration in the Enzyme Reaction and the Optimal Transcreener AMP^2/GMP^2 Antibody Concentration for detection under initial velocity conditions.

The antibody concentration can be calculated using the equation: [Ab] (µg/mL) = 0.3274 x [NAD+] (µM) + 4.24. This equation, though not shown, is valid for NAD^+ concentrations as high as 1000 µM, provided that the Enzyme Reaction and the 1X AMP Detection Mix are equivalent in volume. For example, if you are using 10 µM NAD^+ in a 10 µL Enzyme Reaction, the optimal AMP^2/GMP^2 Antibody concentration in the 10 µL 1X AMP Detection Mix would be (0.327 × 10) + 4.24 = 7.51 µg/mL. In the 20 µL Complete Assay, the optimal AMP^2/GMP^2 Antibody concentration would be half the concentration in the 1X AMP Detection Mix, or 3.75 µg/mL for this example.

Please see **Section 7.2** for more information about optimizing the AMP^2/GMP^2 Antibody concentration for cases in which the concentration calculated is not yielding the results that you require.

Optimize the Enzyme Concentration

4.3 Perform an enzyme titration to identify the optimal enzyme concentration for the Transcreener pADPr PARP FP Assay. Use enzyme buffer conditions and substrate concentrations that are optimal for your enzyme and experimental goals. If a compound screen is planned, you should include the library solvent at its final assay concentration. Run your enzymatic reaction at its requisite temperature and time period.

4.3.1 Enzyme Titration Considerations

For detection of inhibitors at single concentration or in dose response mode, we recommend selecting an enzyme concentration that produces a 50–80% change in FP signal (EC_{50} to EC_{80}) (see **Figure 4**) and an assay window of at least 100 mP. This will result in initial velocity conditions, which correspond to the linear phase of the reaction (after conversion of mP values to AMP formed). The EC_{50} is provided by common graphing programs; the EC_{80} enzyme concentration can be calculated from the EC_{50}, as follows:

$$EC_X = (X \div (100 - X))^{(1 \div |\text{hillslope}|)} \times EC_{50}$$

4.3.2 Performing an Enzyme Assay

1. Prepare Assay Buffer with the required components for the Enzyme Reaction.
2. Dilute the target PARP Enzyme to 2X the desired starting concentration.
3. Add 10 µL of the PARP Enzyme to well 1 (including replicates).
4. Add 5 µL of Assay Buffer to wells 2-11, DO NOT add the Assay Buffer to well 1.
5. Transfer 5 µL from well 1 to well 2 and mix by pipetting, then transfer 5 µL from well 2 to well 3 and mix by pipetting; repeat this serial dilution process until well 11 has received PARP Enzyme. Well 12 is to be used as a blank and should not include enzyme.

IMPORTANT: After mixing the last well (11) in the dilution series, remove 5 µL from that well only and discard, so that all the wells contain 5 µL final volume.

6. Prepare a Substrate/CE Mix composed of the CE, NAD^+ and any other components necessary for enzymatic activity.

Note: Prepare the Substrate/CE Mix right before use to avoid substrate degradation. All components should be added at 2X the desired concentration in the 10 µL Enzyme Reaction. For example, CE concentration is 2X in the Substrate/CE Mix and 1X in the 10 µL Enzyme Reaction.

7. Start the Enzyme Reaction by adding 5 µL of the Substrate/CE Mix in each well. It is recommended to incubate the Enzyme Reaction at 30°C for 60 minutes.
8. Prepare 1X AMP Detection Mix with an appropriate AMP^2/GMP^2 Antibody concentration based on the NAD^+ concentration in the Enzyme Reaction. The following table contains an example preparation based on 100 µM NAD^+:

1X AMP Detection Mix - Add 10 µL Per Well

	Component	As Provided	Detection Mix Concentration	Final Concentration in 20 µL Complete Assay	Final Volume in AMP Detect Mix
	AMP^2/GMP^2 Antibody	1.26 mg/mL	36 µg/mL	18 µg/mL	285.7 µL
	AMP^2/GMP^2 Alexa Fluor 633 Tracer	800 nM	8 nM	4 nM	100 µL
	Stop & Detect Buffer B	10X	1X	0.5X	1000 µL
	Water				8,614.3 µL
	Total				**10,000 µL**

9. Following the incubation, add 10 µL of 1X AMP Detection Mix and mix gently on a plate shaker for 40 to 60 seconds.

Note: After 1X AMP Detection Mix is added to the Enzyme Reaction the final concentration of components in the 20 µL. Complete Assay will be 0.5X AMP Detection Mix (4 nM Alexa Fluor® 633 Tracer, 18 µg/mL AMP^2/GMP^2 Antibody, and 0.5X Stop & Detect Buffer B).

10. Incubate at room temperature (20–25°C) for 120 minutes to allow equilibration of assay components and measure FP.

Note

Note: This is an example of running an assay for HTS or to obtain a dose response. Your volumes and concentrations may vary. It is important to have a 1:1 ratio of Enzyme Mix and Detection Mix for the final assay readout.

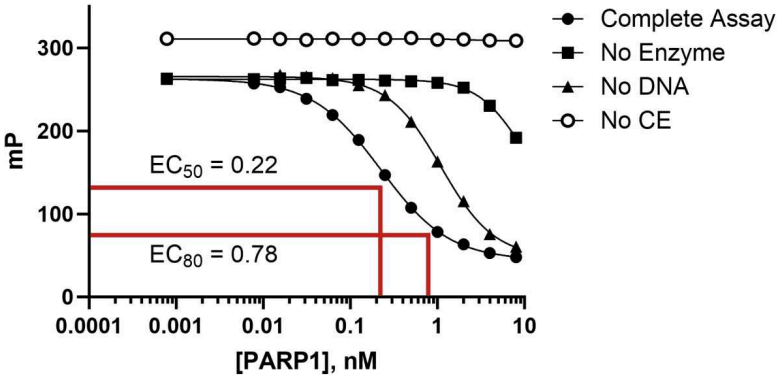


Figure 4. Enzyme titration curve. Example PARP1 Enzyme titration. The ideal range of enzyme concentrations is between EC_{50} and EC_{80}; the specific concentration may vary depending on the enzyme lot.

4.3.3 Enzyme Assay Controls

The enzyme reaction controls define the limits of the enzyme assay.

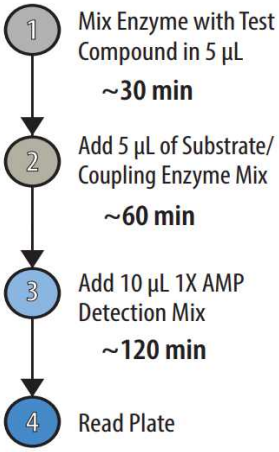
	Component	Notes
	No Inhibitor Control	This is a complete Enzyme Reaction with all detection components. It provides the maximum signal (minimal FP value) for an uninhibited enzyme reaction.
	No Enzyme Control	This contains all Enzyme Reaction components in the absence of the target enzyme. It provides the minimal signal (maximal FP value), mimicking 100% Inhibition.
	Minus-Substrate Control	This is an alternative to a No Enzyme control; it may be more appropriate for enzymes that produce some background signal that is not substrate-dependent.

Performing Single Compound Screening and Dose-Response Assays

4.4 **4.4.1 Experimental Samples**

Single Compound Screening and Dose-Response Assays follow the protocol listed in **Section 4.3.2**, with a slight change to the initial steps. Rather than performing a serial dilution of the target PARP Enzyme, a serial dilution of the test compound is performed. The target PARP Enzyme is then added to the test compounds; the total mixture volume should be 5 µL. The enzyme is added at a total concentration which yields the concentration calculated in **Section 4.3.1** after addition of the remaining components that make up the 10 µL Enzyme Reaction. We recommend mixing gently on a plate shaker for 40 to 60 seconds and preincubating the Enzyme Inhibitor Mix for the desired time (typically at least 15 minutes) at room temperature to allow equilibration of the E-I complex.

Note: Final concentration of test compounds should be based on the volume of the Enzyme Reaction. Compound volume added should be minimized to avoid signal interference caused by the compound or its solvent.



4.4.2 AMP Detection Controls

These controls are used to calibrate the FP plate reader and are added to wells that do not contain enzyme.

	Component	Notes
	Minus Antibody (Free Tracer) Control	This control contains 4 nM AMP^2/GMP^2 Tracer in the 20 µL Complete Assay without the AMP^2/GMP^2 Antibody and is set to low mP, typically between 20-50 mP depending on the instrument.
	Minus Tracer Control	This control contains the AMP^2/GMP^2 Antibody without the AMP^2/GMP^2 Tracer and is used as a sample blank for all wells. It contains the same AMP^2/GMP^2 Antibody concentration that will be used in the assay.

General Considerations

5

Endpoint Assay

5.1 The Transcreener pADPr PARP FP Assay is designed for endpoint readout. The Stop & Detect Buffer B contains EDTA, which quenches the PARP Enzyme and the CE by chelating Mg^{2+}.

Real-Time Assays

5.2 This assay can be performed in real time by eliminating stop reagents and including the 1X AMP Detection Mix components (antibody and tracer) in the Enzyme Reaction. However, this mode should only be used for relative activity comparisons, because the extended signal equilibration time precludes accurate quantitation of AMP in real time. A standard curve run under similar conditions (continuous mode) will help in extrapolating the mP to the amount of AMP product formed.

Reagent and Signal Stability

5.3 **5.3.1 Signal Stability**

The stability of the FP assay window at 10% substrate conversion was determined after the addition of the 1X AMP Detection Mix to the standard samples. The FP assay window at 10% substrate conversion remained constant (<10% Change) for at least 24 hours at room temperature (20–25°C). If you plan to read FP on the following day, seal the plates to prevent evaporation.

5.3.2 AMP Detection Mix Stability

The AMP Detection Mix is stable for at least 16 hours at room temperature (20–25°C). If you prepare the AMP Detection Mix more than 30 minutes before addition, store it on ice or at 4°C until needed to help decrease the equilibration time.

Frequently Asked Questions

6

	Question	Possible Solutions
	Low Selectivity	Suboptimal antibody concentration ▪ To achieve maximum sensitivity and assay window, the AMP^2/GMP^2 Antibody concentration must be optimized for each starting NAD^+ concentration.
	No change in FP observed	Low antibody/tracer activity or Δ mP signal. ▪ The tracer and antibody are stable for up to 6 freeze-thaw cycles. For frequent use aliquot the antibody and tracer and store the aliquots at -20°C. Use a minimum of 20 µL aliquots.
	Is a standard curve required?	No, it is not required to run a standard curve. We recommend running the AMP standard curve if you want to convert raw mP values to product formed. While designing a standard curve, make sure that most of the points are within the area of interest (initial velocity conditions). We do not recommend using a standard curve from previous experiments, rather generate a new curve with each experiment to achieve the most accurate result.
	Can this assay be used with cell lysates?	The assay will only work with purified recombinant protein. The presence of nucleases in the lysates prohibits the use of Transcreener assays with lysates.
	High background signal or change in signal after incubation with detection mixture.	Interference from impurities ▪ Nuclease contamination in the buffer can cause the assay window to collapse, causing a change in FP. We recommend using nuclease-free water and freshly prepared buffer for each assay. ▪ Some compounds may interfere with the detection mixture, causing a change in signal. ▪ Bovine serum albumin (BSA) at concentrations >1% interferes with the detection reagents. Detergents, such as Brij-35, can be substituted for BSA in the enzyme reaction to prevent nonspecific binding of enzymes and substrates to the plate.

Appendix

7

Standard Curve

7.1 Use of a standard curve for conversion of values to amount of AMP formed allows quantitative measurement of the enzyme activity and accurate IC_{50} determinations; it is not typically done for screening at single concentrations. The standard curve mimics the Enzyme Reaction; the NAD^+ concentration remains constant to account for the background generated due to its degradation and reaction with the CE. The AMP standard curve allows calculation of the concentration of AMP produced in the Enzyme Reaction and, therefore, the percent AMP conversion.

In this example, a 12-point standard curve was prepared using the concentrations of AMP shown in the following Table. Commonly, 8- to 12-point standard curves are used.

	Data Point	AMP (µM)
	1	100.00
	2	50.000
	3	25.000
	4	12.500
	5	6.250
	6	3.125
	7	1.563
	8	0.781
	9	0.391
	10	0.195
	11	0.098
	12	0.000

Table 1. AMP Standard Curve. Concentrations of AMP to prepare a 12-point standard curve. Concentrations of NAD^+ are kept constant at a concentration equivalent to the highest concentration of AMP used (100 µM in this case).

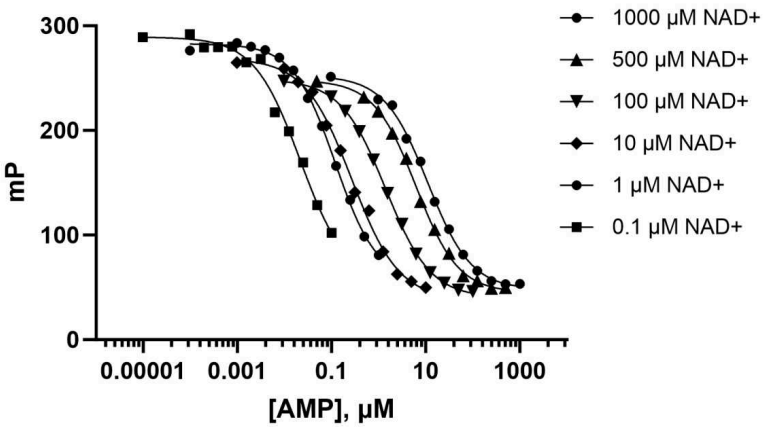


Figure 4. AMP Standard Curve. Sample data for 0.1 µM, 1 µM, 10 µM, 100 µM, and 1,000 µM AMP standard curves. The nucleotide concentration reflects the amount in the enzyme reaction, prior to the addition of the 1X AMP Detection Mix. Curves are obtained from a 20 µL Complete Assay consisting of of 4 nM AMP^2/GMP^2 Alexa Fluor® 633 Tracer, 0.5X Stop and Detect Buffer B, 0.5X reaction buffer (50 mM Tris-HCl pH 7.5, 10 mM $MgCl_2$, and 0.1% Triton 100-X), 0.5X Substrate/CE Mix (1X CE, and NAD^+), AMP standards, and AMP^2/GMP^2 Antibody at varying concentrations based on the initial amounts of NAD^+, as described in **Section 4.2**.

Use the following equations to calculate the Z′ factor:

$$Z' = 1 - \frac{[(3 \times SD_{\text{No Enzyme}}) + (3 \times SD_{\text{Complete Reaction}})]}{|(\text{Mean mP}_{\text{No Enzyme}}) - (\text{Mean mP}_{\text{Complete Reaction}})|}$$

Optimizing the AMP^2/GMP^2 Antibody Concentration

7.2 Using an antibody concentration calculated in **Section 4.2** will produce excellent results for most users. If it does not produce the results you require, we recommend that you perform an AMP^2/GMP^2 Antibody titration in the buffer system ideal for your enzyme target. This titration will determine the optimal antibody concentration for your assay conditions. The substrate concentration in the enzyme reaction generally determines the appropriate concentration of AMP^2/GMP^2 Antibody. We recommend using a concentration of antibody equivalent to the EC_{70} to EC_{85} acquired from the titration.

7.2.1. Titrate the AMP^2/GMP^2 Antibody

1. Prepare the reaction buffer. Example: 50 mM Tris-HCl pH 7.5, 10 mM $MgCl_2$ and 0.1% Triton 100-X.
2. Add 5 µL of the reaction buffer to wells 2–12 (including replicates). Do not add it to well 1.
3. Dilute the AMP^2/GMP^2 Antibody to 800 µg/mL in reaction buffer. Add 10 uL to well 1 of each replicate.
4. Transfer 5 µL from well 1 to well 2 and mix by pipetting, then transfer 5 µL from well 2 to well 3 and mix by pipetting; repeat this serial dilution process until well 12 has

received AMP^2/GMP^2 Antibody.

IMPORTANT: After mixing the last well in the dilution series, remove 5 µL from that well only and discard, so that all the wells contain 5 µL final volume.

5. Prepare Substrate/CE Mix in reaction buffer and add 5 µL into each well. Gently mix the plate for few seconds using a plate shaker.

Note: Prepare the Substrate/CE Mix right before use to avoid substrate degradation. CE concentration is 2X in the Substrate/CE Mix.

6. Add 10 µL of 8 nM AMP^2/GMP^2 Alexa Fluor 633 Tracer in 1X Stop & Detect Buffer B to each well

7. Gently mix on a plate shaker for 40 to 60 seconds and then allow it to incubate at room temperature for 120 minutes before reading.

7.2.2 Calculate the Optimal AMP^2/GMP^2 Antibody Concentration

To determine the AMP^2/GMP^2 antibody concentration for EC_X, input the EC_{50} and hillslope values from a sigmoidal dose-response curve fit into the equation below.

$$EC_X = (X \div (100 - X))^{(1 \div |\text{hillslope}|)} \times EC_{50}$$

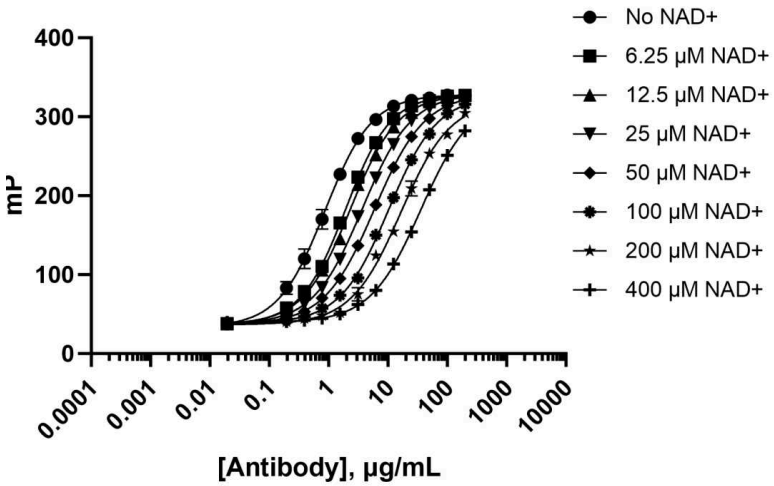


Figure 5. AMP^2/GMP^2 Antibody Titration at various NAD$^+$ concentrations. The 20 µL Complete Assay consisted of 4 nM AMP^2/GMP^2 Alexa Fluor® 633 Tracer, 0.5X Stop and Detect Buffer B, 0.5X reaction buffer (50 mM Tris-HCl pH 7.5, 10 mM $MgCl_2$, and 0.1% Triton 100-X), 0.5X Substrate/CE Mix (1X CE, and NAD$^+$) and AMP^2/GMP^2 Antibody. Though shown for 6.25–400 µM NAD$^+$, the relationship is valid from 0.1 µM to 1000 µM NAD$^+$.

Using the Assay with Different Volumes and Plate Formats

7.3

	Component	Total Volume	Enzyme Reaction Volume	1X AMP Detection Mix Volume
	96 Well Low Volume Plate	50 µL	25 µL	25 µL

	Component	Total Volume	Enzyme Reaction Volume	1X AMP Detection Mix Volume
	384 Well Low Volume Plate	20 µL	10 µL	10 µL
	1536 Well Low Volume Plate	8 µL	4 µL	4 µL

Please check the working plate volumes from the manufacturer to ensure they are within the suggest volumes ranges of your plate.

Links to Applicable Application Notes

7.4
- A Guide to Navigating Hit Prioritization After Screening Using Biochemical Assays
- A Guide to Measuring Drug-Target Residence Times with Biochemical Assays
- List of Commonly Used Plate Readers and Settings

Contact Information

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