

Nov 30, 2022

Version 2

## Synthesis of Silver Nanoparticles V.2

 In 1 collection

DOI

[dx.doi.org/10.17504/protocols.io.q26g7yywkgwz/v2](https://dx.doi.org/10.17504/protocols.io.q26g7yywkgwz/v2)

Raphael D. Ayivi<sup>1</sup>, Bukola Adesanmi<sup>1</sup>, Eric S McLamore<sup>2,3,4</sup>, Sherine O. Obare<sup>1,5</sup>

<sup>1</sup>Department of Nanoscience, Joint School of Nanoscience and Nanoengineering, University of North Carolina, Greensboro, NC 27412, USA;

<sup>2</sup>Agricultural Sciences Department, Clemson University, USA;

<sup>3</sup>Department of Environmental Engineering and Earth Sciences, Clemson University, USA;

<sup>4</sup>Global Alliance for Rapid Diagnostics, Michigan State University, USA;

<sup>5</sup>Department of Nanoengineering, Joint School of Nanoscience and Nanoengineering, North Carolina A & T State University, Greensboro, NC 27411, USA \* corresponding author

SNAPS research group



**Eric S McLamore**

Clemson University, North Carolina State University

### Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



**DOI:** <https://dx.doi.org/10.17504/protocols.io.q26g7yywkgwz/v2>

**Protocol Citation:** Raphael D. Ayivi, Bukola Adesanmi, Eric S McLamore, Sherine O. Obare 2022. Synthesis of Silver Nanoparticles. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.q26g7yywkgwz/v2> Version created by **Eric S McLamore**

**Manuscript citation:**

**1.** Brown, A. N. et al. Nanoparticles functionalized with ampicillin destroy multiple-antibiotic-resistant isolates of *Pseudomonas aeruginosa* and *Enterobacter aerogenes* and methicillin-resistant *Staphylococcus aureus*. *Appl. Environ. Microbiol.* 78, 2768–2774 (2012); **2.** Perry, J. & Baum, J. Assessing the Laboratory Environment. in *Accessibility in the Laboratory* vol. 1272 3–25 (American Chemical Society, 2018).

**License:** This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

**Protocol status:** Working

**We use this protocol and it's working**

**Created:** November 29, 2022

**Last Modified:** November 30, 2022

**Protocol Integer ID:** 73333

**Keywords:** nanoparticle, synthesis, silver, AgNP, synthesis of silver nanoparticle, silver nanoparticle, colorimetric sensing of environmental sample, colorimetric sensing, other biochemical application

**Funders Acknowledgements:**

**National Science Foundation**

Grant ID: ECCS-2025462

**National Science Foundation**

Grant ID: EIR-1832134

**National Science Foundation**

Grant ID: CBET-2019435

## Disclaimer

Any opinions, findings, and conclusions or recommendations expressed in this piece are those of the author(s) and do not necessarily reflect the views of the National Science Foundation.

## Abstract

This protocol describes the synthesis of silver nanoparticles for colorimetric sensing of environmental samples and for other biochemical applications. The protocol requires 110 minutes to complete (excluding working solution prep the protocol requires 65 min).

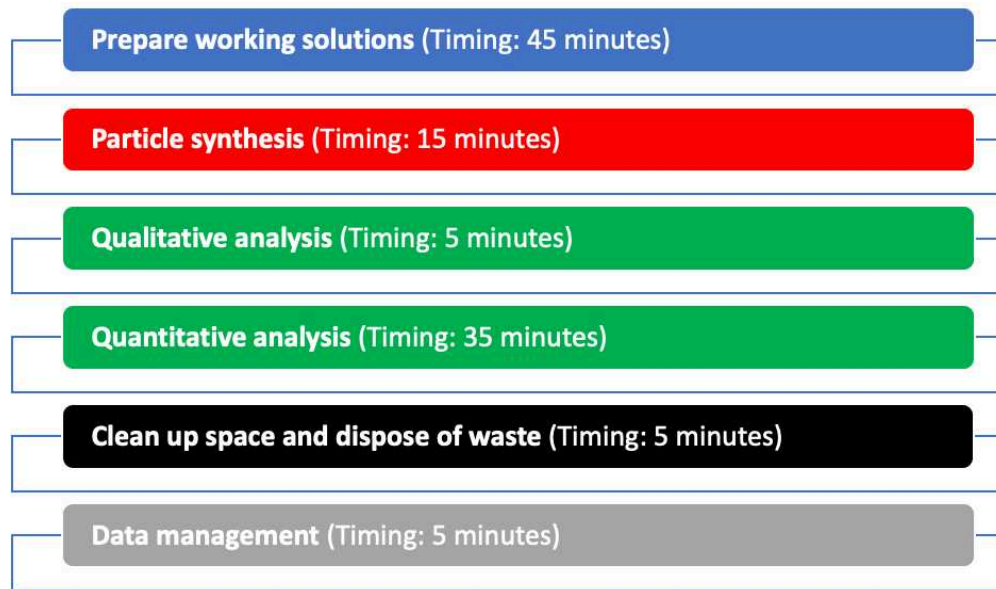
## Image Attribution

bWWoIVW4RnsOA58BTTcfNA==

## Guidelines

This protocol describes the synthesis of silver nanoparticles for colorimetric sensing of environmental samples and for other biochemical applications. The steps for the process are based on Brown et al (REF 1).;

The steps for the process are based on Brown et al <sup>1</sup>; summarized in **Figure 1**



Process flow for synthesis of silver nanoparticles. The protocol is organized by sections: preparation (blue), particle synthesis (red), particle analysis (green), cleanup (black), and data management (grey).

## Materials

### MATERIALS

- Sodium citrate ( $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$ ) safety data sheet ([link here](#))
- Silver nitrate ( $\text{AgNO}_3$ ) safety data sheet ([link here](#))
- Sodium borohydride ( $\text{NaBH}_4$ ) safety data sheet ([link here](#))
- Aluminum foil
- Deionized Water

### HARDWARE / CHARACTERIZATION INSTRUMENTS

- Stir plate (Corning PC-420D; [link here](#))
- UV-Vis Spectroscopy (Agilent-Cary 60; link to manual is [here](#))
  - Fluorescence spectroscopy (Horiba Fluoro-Max 4 Fluorometer; link to manual is [here](#))
  - Inductively coupled plasma mass spectrometry (ICP-ES) (Agilent Varian 710-ES ICP Spectrophotometer; link to manual is [here](#))
- For all instruments and equipment in lab, ADA-compliant guidance<sup>2</sup> is followed (see safety and accessibility section for details)

### SOFTWARE

- Optional: Color Name AR app ([link here](#)), colorimeter or ([link here](#)), or spectroradiometer ([link here](#)) for qualitative sample analysis

## Troubleshooting

## Safety warnings

### SAFETY

#### *General*

- Lab coat, gloves, and closed-toed shoes are mandatory
- Nitrile gloves (powder-free type as applicable)

#### *Chemical safety hazard*

- When using scale weight to measure chemicals, ensure the residues are cleaned and discarded in the solid chemical waste container as described by the solid waste disposal procedure and in reference to the SDS for all chemicals.
- All chemicals, particularly reducing agent (sodium borohydride) must be contained within a chemical hood at all times. Sodium borohydride is soluble in water, but reacts with water. When dissolved in water, hydrogen gas (extremely flammable) is formed as a reaction byproduct.
- If chemicals are spilled, follow procedures in SDS (links provided with materials section).

#### *Eye protection*

- Goggles or eye protection is required when handling acidic solutions outside of a chemical hood (for transport or analysis only)

#### *Skin*

If any solutions are spilled and contact skin, immediately rinse under water and wash with soap for at least 5 min.

#### *Fumes/aerosols*

- Any open containers should be processed/handled under a certified chemical hood/safety cabinet.
- All acidic solutions should be covered with parafilm when outside chemical hood
- Aerosolization of particles (nano or colloidal) and subsequent health effects are unknown, and samples should be treated as hazardous and capped

#### *Disposal*

- Vials of nanoparticles/colloids should be discarded in the waste container and not disposed in the sink.

### ACCESSIBILITY

The following guidance is summarized from Perry and Baum <sup>1</sup> where relevant to this protocol.

#### 1) General building codes for laboratory

- Minimum 2 Exits for labs  $\geq 500$  ft<sup>2</sup> net area.
- Minimum 2 Exits for labs using chemical fume hoods or glove box



- Minimum 2 Exits for labs using flammable and combustible: liquids, gases, cryogenics, dusts and solids.
- Minimum 2 Exits for labs using oxidizers, unstable reactives, water reactives, organic peroxides, highly toxics, corrosives.

2) Egress for wheelchair 360° Turn is 1.5 m (5 ft) clearance. Wheelchair clearance must be provided for:

- Both sides of Exit and Entry doors to
- Emergency Eyewash & Safety Shower
- In front of wall benches, sinks, equipment
- In front of chemical fume hoods
- At chalk/marker board
- Between benches
- Aisles that lead to Primary Exits, back to front
- Aisles that allow passage side to side in lab

3) Standard accommodations for use of chemical hood or other exhaust air containment systems

- knee space obstructions
- adjustable work surface height
- accessible receptacles and alarm control

#### *Common equipment*

- Where visual inspection is utilized, alternative technologies should be listed as optional (colorimeters, spectro-radiometers, etc.)

#### **References**

1. Perry, J. & Baum, J. Assessing the Laboratory Environment. in *Accessibility in the Laboratory* vol. 1272 3–25 (American Chemical Society, 2018).

## **Before start**

- Be sure to wear appropriate safety PPE throughout (lab coat, gloves, eyewear).
- Electronic or physical lab notebook may be used throughout
- See experimental plan guide for tips on planning your work



## SECTION 1) Preparation

45m

### 1 **Prepare stabilizer solution** (0.01 M Sodium citrate; $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$ )

10m

- Prepare a glass beaker with 50 ml of deionized water and label
- Weigh 0.1290 g of trisodium citrate on scale
- Dissolve sodium citrate powder in prepared glass beaker with deionized water
- Place magnetic stir bar in beaker and place on a stir plate
- Stir for 15 minutes at 150-200 rpm
- After stirring, inspect the beaker to ensure dissolution of the solute
- The trisodium citrate serves as the stabilizer and is ready to be used for particle synthesis

### 2 **Prepare precursor solution** (0.01 M Silver nitrate; $\text{AgNO}_3$ )

10m

- Prepare a glass beaker with 50 ml of deionized water and label
- Weigh 0.085 g of silver nitrate on scale
- Dissolve powder in prepared glass beaker with deionized water
- Place magnetic stir bar in beaker and place on a stir plate
- Stir for 15 minutes at 150-200 rpm
- After stirring, inspect the beaker to ensure dissolution of the solute
- The silver nitrate is the precursor and is ready to be used for particle synthesis

### 3 **Prepare reducing agent solution :** (0.01 M Sodium borohydride ( $\text{NaBH}_4$ ))

25m

- Prepare a sealable container with 50 ml of 0.1N NaOH and label. Place container in the chemical hood

#### Note

**Note:** *If preferred, smaller volumes of sodium borohydride (1ml is sufficient for this protocol) may be prepared fresh prepared and kept in ice-cold environment rather than preparation of a large 50ml solution as described.*

- Cool the NaOH solution in an ice bath under the chemical hood (cool to approximately 0 to 5°C)
- Working under a chemical hood, weigh 0.020 g of sodium borohydride on scale
- Immediately return sodium borohydride powder to proper storage system (away from moisture or water, stored under inert gas in a flammable containment unit)
- Dissolve sodium borohydride powder in cooled 0.1N NaOH and label
- Seal the container with the sodium borohydride
- Place magnetic stir bar in beaker and place on a stir plate



- Stir for 15 minutes at 150-200 rpm
- After stirring, inspect the beaker to ensure dissolution of the solute
- The sodium borohydride serves as the reducing agent and is ready to be used for particle synthesis
- Store the sealed container at 4°C until used (best practice is to place in a secondary spill container, particularly if sealed container is glass)

#### Note

**Safety note:** Sodium borohydride is a strong reducing agent. Solutions should always be kept in ice-cold environment.



GHS Classification for sodium borohydride :

Chemicals which, in contact with water, emit flammable gases (Category 1; H260)

Acute toxicity, Oral (Category 3; H301)

Skin corrosion (Category 1B; H314)

Serious eye damage (Category; H318)

Reproductive toxicity (Category 1B; H360)

## SECTION 2) Particle synthesis

15m

### 4 Combine stabilizer and precursor solutions

10m

- In an Erlenmeyer flask, add 18.5ml of deionized water
- Add a magnetic stir bar to the flask
- Pipette 0.5 ml of 0.01M sodium citrate to the flask
- Pipette 0.5 ml of 0.01M silver nitrate to the flask
- The resulting mixture in the flask should be gently stirred for 3 minutes at a temperature of 10°C. The stir rate should be the lowest stirring rate that maintains constant agitation

### 5 Add reducing agent solution to mixture

5m

#### Note

- **Critical step:** Stirring should be immediately suspended upon the first dropwise addition of  $\text{NaBH}_4$





- Slowly pipette 0.5 ml of 0.01M  $\text{NaBH}_4$  to the solution to the flask containing the reaction mixture with a stir rate no higher than 50 rpm.
- Slowly pipette 0.5 ml of 0.01M  $\text{NaBH}_4$  to the solution to the flask containing the reaction mixture with a stir rate no higher than 50 rpm.

#### Note

**Critical step:** Reducing agent should be slowly added to ensure gentle nucleation and proper growth of nanoparticles.

- Seal Erlenmeyer flask after 0.5 ml has been transferred
- Under a chemical hood, carefully transfer solution to container with lid and label as AgNP solution
- Wrap container in aluminum foil
- Transfer AgNP solution from flask to container and seal

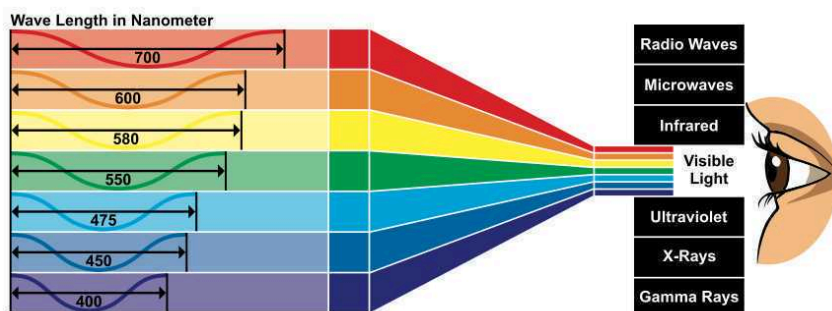
## SECTION 3) Particle analysis (post reaction)

40m

### 6 Qualitative analysis (color)

5m

- The first step of qualitative analysis is visual inspection of the solution color
- The color of the reaction mixture (gold or yellow) confirms the formation of AgNP and completion of the synthesis reaction.
- Analysis of polychromatic visible (VIS) color is an important quality control mechanism in particle synthesis, and may be carried out manually or using instrumentation (**Fig 1**)

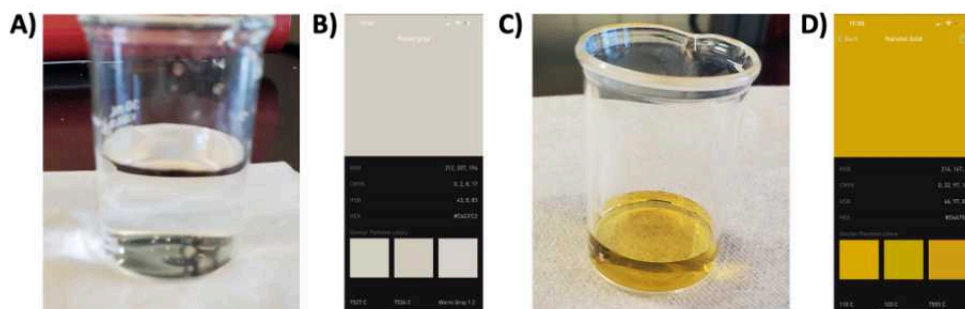


**Figure 1.** Visual inspection in the VIS spectra is an important quality control mechanism in particle synthesis. If visual inspection is not possible,, a colorimeter or other device may be used to visually inspect the samples (see note below for example of mobile phone app). Image courtesy of Shutterstock standard license (no. 514067857)

- A representative example of color analysis is shown in **Fig 2**
- Photographs before (**Fig 2A**) and after (**Fig 2C**) reaction are shown. Color analysis via a mobile phone app are also shown for before (**Fig 2B**) and after (**Fig 2D**) the reaction.

## Note

**Note:** If preferred, a cell phone app may be used to detect the color of the sample. See **Fig 2**. For example, Color Name AR is a useful tool that is available for both iPhone and Android (as well as tablets) (<https://apps.apple.com/us/app/color-name-ar/id906955675>)



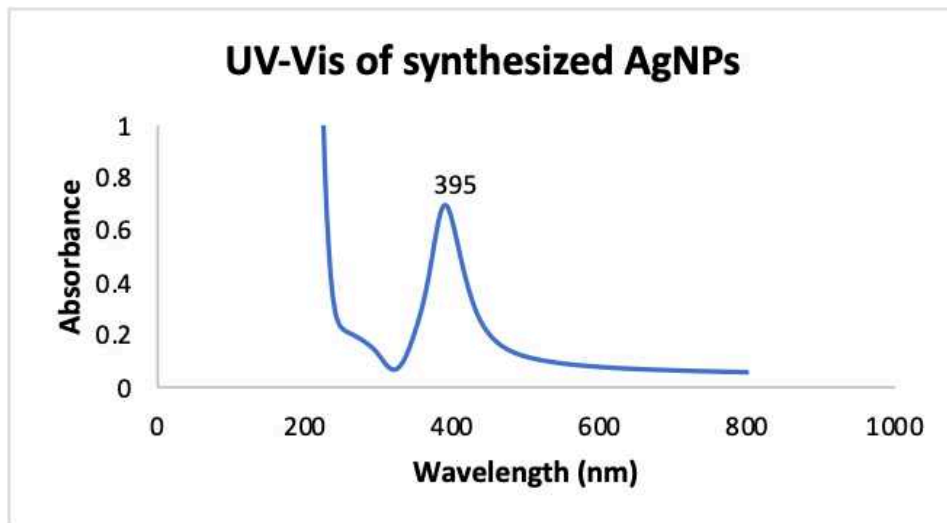
**Figure 2.** Qualitative analysis of solution color before and after particle synthesis. **A)** Photograph depicts colorless reactant solution before and at onset of reaction, and **B)** Color analysis; sample identified as Pasei grey; RGB 212, 207, 194). **C)** Photograph of AgNP particle solution after reaction, and **D)** Color analysis; Sample identified as Harvest gold; RGB 214, 167, 006). Color analysis was conducted using Color Name AR on iPhone 12.

## 7 Quantitative analysis (UV-VIS)

- If required, turn on the computer for the Agilent UV-VIS Cary-60 instrument and then turn on the instrument and wait until the LED stops blinking.
- Open the Agilent software and wait until the instrument fully initializes.
- To select experimental parameters, select Setup in the Agilent software.
- Select wavelength range if desired (default is 190-1100 nm)
- Click the baseline tab, select "Baseline correction" and select OK.
- Click on "Zero" for lamp intensity correction. When performing this procedure, the sample compartment has to be empty.
- Click on "Baseline".
- Fill an empty cuvette with DI water (blank) and another cuvette with the silver nanoparticle (AgNP) sample
- Wipe the blank cuvette with a Kimwipe and inspect the surface for any debris
- Place the blank cuvette in the sample compartment.
- Close the lid of the chamber
- Click OK to scan the baseline
- Remove the blank
- Wipe the sample cuvette with a Kimwipe
- Insert the sample cuvette into the holder
- Click Start to run the scan
- When finished with measurement, select Finish
- If processing multiple samples with the same parameters, select OK after each run.

20m

- If processing a large batch of samples (>20), scan a blank cuvette every 7 samples as a quality control (QC) procedure
- After the scan is complete, save the file under a unique name (see data management section)
- After finishing the experiments, close the software.
- Leave both the instrument and the computer on.
- Representative UV-VIS absorbance for the AgNP solution using an excitation wavelength of 190nm is shown in **Fig 3**
- A clear peak at 395nm is observed for the AgNP solution prepared using this protocol (no custom emission filters or signal processing was performed).



**Figure 3.** Quantitative analysis of as prepared particles using a UV-Vis Spectrophotometer (Agilent UV-VIS Cary-60). The observed wavelength shift is due to surface plasmon resonance and is typically between 390nm -700nm dependent on the size of the nanoparticles synthesized.

## 8 Quantitative analysis (fluorescence)

- If required, turn on the Horiba Fluoro-Max 4 Fluorometer instrument and wait for the software and hardware to initialize
- Open the Horiba software

15m

#### Note

**Note:** The instrument auto-calibrates on startup (initializes monochromator drives, assigns calibration wavelengths). However, it is a good practice to periodically check the instrument calibration. This check verifies the wavelength calibration of excitation monochromator. To perform check, close the lid and select "Experiment Menu" Button, Choose "Spectra", then "Excitation", and click "Run". The spectra should show a peak at 467 nm if working properly. If not working properly, see user manual to manually adjust (section 3-6; link to user's manual in Equipment section). A similar process may be periodically conducted for checking the emission photomultiplier tube calibration.

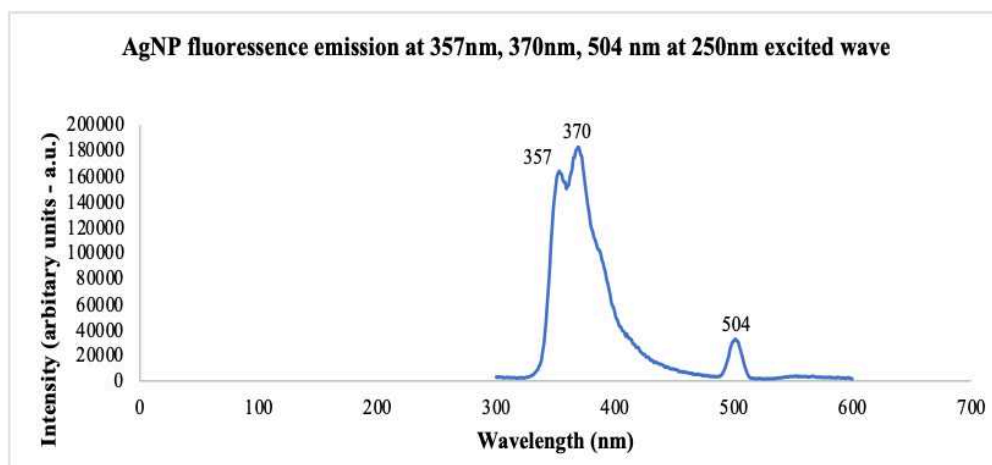
- Choose the "Experiment Menu" Button
- Choose "Spectra",
- then "Emission"
- Click the Experiment File field, and enter a new file name or select a previously saved file (see data management section)
- Verify that experimental parameters are correct
- Prepare a blank sample in a cuvette and the AgNP solution

#### Note

**Note:** Blank samples should be research-quality, triple-distilled or de-ionized water. Impure samples of water will cause elevated background levels as well as distorted spectra with (perhaps) some unwelcome peaks.

- Wipe the blank cuvette with a Kimwipe and inspect the surface for any debris
- Place the blank cuvette in the sample compartment.
- Close the lid of the chamber
- Click "Run" to scan the baseline
- Enter a name for the project, or browse for an existing project, then click OK
- Open the lid and remove the blank sample from the compartment
- Wipe the sample cuvette with a Kimwipe
- Insert the sample cuvette into the compartment
- Close the lid and then click "Run"
- When finished with measurement, select Finish
- If processing multiple samples with the same parameters, select Ok after each run.
- If processing a large batch of samples (>20), scan a blank cuvette every 7 samples as a quality control (QC) procedure
- After the scan is complete, save the file under a unique name (see data management section)
- After finishing the experiments, close the software.
- A representative fluorescence emission for the AgNP solution using an excitation wavelength of 250nm is shown in **Fig 4**.
- No custom emission filters or signal processing was performed

- A bimodal peak contains distinct peaks at 357nm and 370 nm, and an isolated smaller peak at 504nm are common.



**Figure 4.** Representative fluorescence under an excitation of 250nm from synthesized sample

## SECTION 5) Cleanup

5m

### 9 Clean up space and dispose of waste

5m

- Turn off magnetic stir plate and all equipment used
- Wrap storage vials in aluminum foil
- Store solution(s) in the dark at 4°C.
- Dispose of used chemicals according to the lab safety plan

#### Note

**Critical step:** There is a dedicated chemical disposal container for waste materials from this protocol in the satellite accumulation area.

- Wash all glassware with mild detergent and warm water.
- Clean up the chemical hood
- Rinse used pipette tips or other materials used for handling reducing agents (collecting rinse in waste jar)
- Dispose of rinsed tips per solid waste handling procedures (without this rinse step, used materials contain residue)



## SECTION 6) Data management

5m

- 10
- File naming: For saving all files from spectrometer, use the following file structure or similar structure as appropriate for each laboratory:

AgNP\_(instrument name)\_(date of experiment)\_(initials of researchers).txt

### Note

**Note:** *Text files may be referred to as ASCII files in some software*

- File storage: Store all spec data in the Desktop folder connected to the instrument using the naming procedure above.
- 
- Backup files: At least once per year, ensure that the folder is backed up on the lab external hard drive.