

Mar 20, 2023

🌐 Synaptic immunohistochemistry - wholemount via acetone permeabilization

DOI

<https://dx.doi.org/10.17504/protocols.io.261ge38q7l47/v1>

Anya Suppermpool¹, FishFloorUCL¹

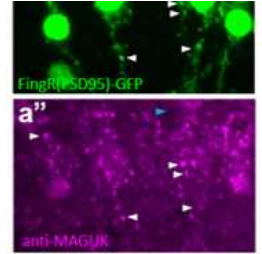
¹University College London

FishFloorUCL



Anya Suppermpool

UCL



Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.261ge38q7l47/v1>

Protocol Citation: Anya Suppermpool, FishFloorUCL 2023. Synaptic immunohistochemistry - wholemount via acetone permeabilization. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.261ge38q7l47/v1>



License: This is an open access protocol distributed under the terms of the **[Creative Commons Attribution License](#)**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

Created: March 08, 2023

Last Modified: March 22, 2023

Protocol Integer ID: 78330

Keywords: synaptic immunohistochemistry, mount immunohistochemistry, immunohistochemistry, wholemount via acetone permeabilization whole, acetone permeabilization, acetone permeabilization whole, maguk

Abstract

Whole-mount Immunohistochemistry – acetone permeabilization

Works with anti-MAGUK ab.

Modified from M Westerfield protocol.

Lavinia Sheets: T. Nicolson Lab September, 2010

Revised August 2011

Anya Suppermpool Synaptic (MAGUK) Version 2018

Anya's Benchling Protocol Archive at:

<https://benchling.com/s/prt-nLJhuyARGTPcTi887wL1?m=slm-FGIOfoYTo8bWHpVnUZ0v>



Materials

PO₄ buffer

8 parts 0.1M NaH₂PO₄

2 parts 0.1M Na₂HPO₄

BT buffer (make fresh)

1.0g sucrose

18.75 μL 0.2M CaCl₂

PO₄ buffer to 15mL

BT fix (make fresh)

1.2mL PO₄ buffer

4.8mL BT buffer

2mL 16% PFA (used bought Thermofisher fix)

PBS/BSA/DMSO

50mL PBS

0.5g BSA

500 μL DMSO



Reagents

- 1 PO₄buffer
8 parts 0.1M NaH₂PO₄
2 parts 0.1M Na₂HPO₄
- 2 BT buffer (make fresh)
1.0g sucrose
18.75μL 0.2M CaCl₂
PO₄buffer to 15mL
- 3 BT fix (make fresh)
1.2mL PO₄buffer
4.8mL BT buffer
2mL 16% PFA (used bought Thermofisher fix)
- 4 PBS/BSA/DMSO
50mL PBS
0.5g BSA
500μL DMSO

Day 1

- 5 Tricaine, Dechorionated beforehand (2dpf)
- 6 Fix 1.5-2h in BT fix (*use bought fix) at 4C
- 6.1 (less fix time higher SNR but mushy) 5h fixation works for 5dpf – 9dpf larvae, time may need to be adjusted for different ages
- 7 Replace fixative with PO₄ buffer.
(Optional) Store at 4C overnight (shaking not necessary)
- 8 Wash for 5' RT with shaking if continuing with permeabilization and blocking in same day
- 9 Chill a small volume (~50mL glass bottle) acetone at -20C at least 20' prior to perm steps.



(optional) if rushed/forgotten, put acetone in -80C for a few minutes, but this could lead to over-permeabilization

- 10 Transfer larvae to glass vials using glass Pasteur pipette.
Larvae will be sticky so be careful while transferring between tubes!

- 11 Wash with dH₂O 5' RT with rocking.
Timing is critical for permeabilization steps—process tubes in same order for each step and adjust volume of solutions to allow for quick handling

- 12 Wash with cold acetone 5' at -20C (put in freezer)

- 13 Wash with dH₂O 5' RT with rocking.
This step can go slightly longer (~10') if necessary

- 14 Wash with PO₄, 5' at RT (Lavinia's original protocol extra step)

- 15 Block >2h RT in PBS/BSA/DMSO + 2% goat serum.
(optional) can go overnight at 4C

- 16 Incubate in primary antibodies in PBS/BSA/DMSO 4C overnight.
Use ~1mL antibody mix per tube

Day 2

- 17 Remove primary antibody mix.
Add 1:1000 20% NaN₃ to mix if saving for future use

- 18 Wash 5X+ with PBS/BSA/DMSO >20' RT with rocking.
(optional) any washes after antibody incubation can go overnight at 4C if necessary

- 19 Incubate with secondary antibodies in PBS/BSA/DMSO 2-3h at RT or overnight at 4C in the dark

Day 3

- 20 Wash 3X with PBS/BSA/DMSO >20' RT with rocking
(Wash with PBS/BSA/DMSO + DAPI (1:2000 dilution) >20' RT with rocking)



- 21 Wash with PBS > 20' RT with rocking like 5x
- 22 Move to glycerol – 20%, 40%, 60%, 80%
- 23 Mount and image using glycerol lens (did 20x before)

24

	Reagents	B	C
	Anti-MAGUK	1:500	MABN72 Anti-pan-MAGUK, clone K28/86
	Fix	#28906	ThermoFisher (from Ana Faro)
	tRFP	1:500	Anti-tRFP Rabbit Polyclonal (AB233 - evrogen)