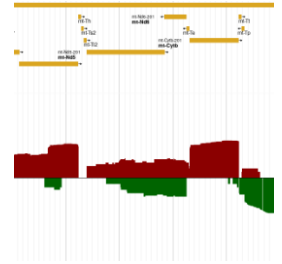


🌐 Stranded Mapping from Oriented Long Reads V.4

[dx.doi.org/10.17504/protocols.io.z9zf976](https://doi.org/10.17504/protocols.io.z9zf976)



¹Malaghan Institute of Medical Research (NZ)



Malaghan Institute of Medical Research (NZ)

OPEN  ACCESS



External link: <https://bioinformatics.stackexchange.com/a/3922/73>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

We are still developing and optimizing this protocol

Last Modified: April 28, 2019

Protocol Integer ID: 22553



Abstract

This protocol demonstrates how to map strand-oriented long reads to a genome, and visualise them in a genome browser.

The general idea is to use minimap2 to create stranded BAM files, which are split for forward/reverse orientation then converted into BigWig format for display in a genome browser.

Input(s):

- stranded fastq files (see protocol [Preparing Reads for Stranded Mapping](#))
- a FASTA file containing the genome / sequence of interest.

Output(s):

- Genome-mapped stranded BAM files
- Genome-mapped stranded BigWig files

Before start

You will need access to the following free and open-source software program(s):

- [minimap2](#)
- [samtools](#)

And the following additional data file(s):

- a FASTA file containing the genome / sequence of interest.



Orient Reads

- 1 Orient reads as per protocol **Preparing Reads for Stranded Mapping**.

If this has been done, then the following command should produce output without errors:

```
for bc in $(awk '{print $2}' barcode_counts.txt);  
do ls oriented/${bc}_reads_dirAdjusted.fastq.gz;  
done
```

Example output:

```
oriented/BC03_reads_dirAdjusted.fastq.gz  
oriented/BC04_reads_dirAdjusted.fastq.gz  
oriented/BC05_reads_dirAdjusted.fastq.gz  
oriented/BC06_reads_dirAdjusted.fastq.gz  
oriented/BC07_reads_dirAdjusted.fastq.gz  
oriented/BC08_reads_dirAdjusted.fastq.gz
```

Index Preparation

- 2 Prepare genome index for spliced alignment

Software

minimap2

NAME

Linux

OS

Heng Li

DEVELOPER

<https://github.com/lh3/minimap2>

SOURCE LINK



```
minimap2 -d mmus_ucsc_all-splice.idx -Q -t 10 -x splice  
mmus_ucsc_all.fa
```

Read Mapping

- 3 Map the long reads to the genome using minimap2, using samtools to convert to a sorted BAM format. This is where the reverse complementing done during demultiplexing gives a big saving of effort.

Software

SAMtools

NAME

Linux

OS

Wellcome Trust Sanger Institute

DEVELOPER

<https://github.com/samtools/samtools>

SOURCE LINK

```
mkdir -p mapped;  
for bc in $(awk '{print $2}' barcode_counts.txt);  
do echo ${bc};  
  minimap2 -t 10 -a -x splice mmus_ucsc_all-splice.idx  
oriented/${bc}_reads_dirAdjusted.fastq.gz | \  
  samtools view -b | samtools sort >  
mapped/mm2_called_${bc}_vs_MmusG.bam;  
done
```

Creating BigWig Coverage Files

4

 mpileupDC.pl



A bedGraph of coverage is created using samtools mpileup and [mpileupDC.pl](#), excluding any skipped intronic sequence. When 'mpileupDC.pl' is provided with a single file, it will output a bedGraph file with a header line starting with '##'; this header line is removed. The particular JBrowse plugin that I use for stranded display requires that the reverse strand have *negative* coverage values, so that file needs to be changed:

```
for bc in $(awk '{print $2}' barcode_counts.txt);
do echo ${bc};
samtools view -b -F 0x10 mapped/mm2_called_${bc}_vs_MmusG.bam | \
    samtools mpileup -A -B -Q 0 -q 0 -I -q 0 -Q 0 - | \
    mpileupDC.pl | tail -n +2 >
mapped/mm2_called_${bc}_vs_MmusG.bg.plus
samtools view -b -f 0x10 mapped/mm2_called_${bc}_vs_MmusG.bam | \
    samtools mpileup -A -B -Q 0 -q 0 -I -q 0 -Q 0 - | \
    mpileupDC.pl | tail -n +2 >
mapped/mm2_called_${bc}_vs_MmusG.bg.minus
perl -i -pe 's/([0-9]+)$/-${1}/'
mapped/mm2_called_${bc}_vs_MmusG.bg.minus
done;
```

- 5 Stranded bedgraph files are converted to bigwig. This requires BEDTools and a genome information file containing chromosome lengths (one for Mmus/mm10 is attached to this step).

Software

BEDTools

NAME

Quinlan laboratory, University of Utah

DEVELOPER

<https://github.com/arq5x/bedtools2/>

SOURCE LINK

```
for bc in $(awk '{print $2}' barcode_counts.txt);
do echo ${bc};
  basename="mapped/mm2_called_${bc}_vs_MmusG"
  bedGraphToBigWig ${basename}.bg.plus Mmus_genome.chrInfo.txt
  ${basename}.bw.plus
  bedGraphToBigWig ${basename}.bg.minus Mmus_genome.chrInfo.txt
  ${basename}.bw.minus
done
```



Mmus_genome.chrInfo.txt

JBrowse Configuration

- 6 Each track should have its own JBrowse configuration section using the *StrandedBigWig* class and *StrandedXYPlot* type. An example is shown here:

```
[tracks.BWCG004-4T1-BC04-both-track ]
storeClass      =
StrandedPlotPlugin/Store/SeqFeature/StrandedBigWig
urlTemplate     = bw/mm2_called_CG004_BC04_vs_MmusG.bw
category       = MinION - Coverage
type           =
StrandedPlotPlugin/View/Track/Wiggle/StrandedXYPlot
key            = MinION minimap2 coverage from CG004-4T1-WT
(combined strands)
scale          = log
scoreType      = maxScore
autoscale      = global
style.pos_color = darkred
style.neg_color = darkgreen
```

Sanity Check

- 7 If this has worked properly, then mapping human or mouse to the mitochondrial genome should show most expression appearing on the positive strand, with a small scattering of negative-strand expression, a bit like the *Expected Results* shown here.

If not, check for the following issues:

- Tracks not displaying at all in JBrowse -- make sure track IDs inside square brackets are of the form [*tracks.<unique-id-without-dots>-track*]
- JBrowse track is reflected in the X axis -- make sure that the reverse bedgraph file is orientated the correct way; it should be created with the '-f 0x10' flag (no capitalisation).
- JBrowse track only shows one direction -- make sure that the reverse bedgraph file has *negative* values, and re-generate the bigwig file

Expected result

