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Stereotaxic Surgery for Viral Injection and Mild 6-OHDA Lesion Model in Mice V.1

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Marta Graziano^{1,2}, Ioannis Mantas^{1,2}, Konstantinos Meletis^{1,2}

¹Karolinska Institutet;

²Aligning Science Across Parkinson's (ASAP) Collaborative Research Network, Chevy Chase, MD 20815



Cláudia C. Mendes

University of Oxford

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We use this protocol and it's working

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Abstract

This protocol outlines the procedure for performing stereotaxic surgery for viral or drug injections and the establishment of a mild 6-OHDA lesion model in mice. The protocol ensures accurate targeting and minimal discomfort, while also detailing postoperative care and monitoring for effective recovery.

Materials

Anesthetics and Analgesics:

- Isoflurane (4% induction, 1.5-2.5% maintenance)
- Buprenorphine (0.1 mg/kg, subcutaneous)
- Xylocaine/Lidocaine (4 mg/kg, local)
- Carprofen (5 mg/kg, subcutaneous)

Pharmacological Agents for 6-OHDA Lesion:

- Desipramine (25 mg/kg, i.p.; Cat# D3900, Sigma-Aldrich)
- Pargyline (5 mg/kg, i.p.; Cat# P8013, Sigma-Aldrich)
- 6-OHDA (0.6 µg/µL in 0.02% ascorbate; Cat# 162957, Sigma-Aldrich)

Surgical Equipment:

- Stereotaxic frame (Harvard Apparatus, Holliston, MA)
- Quintessential Stereotaxic Injector (Stoelting, Wood Dale, IL)
- Capillary needle for viral injection and 6-OHDA injection
- Heating pad
- Eye ointment
- Surgical scissors and forceps
- Sterile gauze and cotton swabs

Sterile Supplies:

- Surgical gloves
- Face mask
- Sterile drapes

Troubleshooting

Animal Preparation

- 1 Anesthesia and Analgesia:
 - 1.1 Induce anesthesia with 4% isoflurane in an induction chamber.
 - 1.2 Maintain anesthesia at 1.5-2.5% isoflurane throughout the procedure.
 - 1.3 Administer Buprenorphine (0.1 mg/kg, subcutaneous) for preoperative analgesia.
 - 1.4 Apply Xylocaine/Lidocaine (4 mg/kg) locally to the surgical site.
- 2 Pre-treatment for 6-OHDA Lesion Model:
 - 2.1 Administer Desipramine (25 mg/kg, i.p.) to protect noradrenergic neurons.
 - 2.2 Administer Pargyline (5 mg/kg, i.p.) to inhibit monoamine oxidase and prolong 6-OHDA action.
 - 2.3 Wait for at least 30 minutes after pretreatment before proceeding with surgery.
- 3 Positioning and Stabilization:
 - 3.1 Place the anesthetized mouse in the stereotaxic frame.
 - 3.2 Secure the head using ear bars to ensure stability.
 - 3.3 Maintain body temperature using a heating pad set to approximately 37°C.

3.4 Apply eye ointment to protect the eyes from drying.

Surgical Procedure

4 Incision and Exposure:

4.1 Make a midline scalp incision to expose the skull.

4.2 Clean the exposed area using sterile saline or ethanol.

5 Coordinate Identification and Drilling:

5.1 Identify the target injection site using stereotaxic coordinates.

5.2 Drill a small hole at the specified coordinates on the skull.

6 Viral Injection (Optional):

6.1 Attach a capillary needle to the Quintessential Stereotaxic Injector.

6.2 Slowly lower the needle to the target brain region.

6.3 Inject the viral vector at a controlled rate.

6.4 Leave the needle in place for 5 minutes post-injection to prevent backflow.

7 6-OHDA Injection:

- 7.1 Prepare a 6-OHDA solution (0.6 $\mu\text{g}/\mu\text{L}$ in 0.02% ascorbate).
- 7.2 Use the following stereotaxic coordinates for injection into the medial forebrain bundle of the right hemisphere: AP: -1.2 mm , ML: $+1.25\text{ mm}$, DV: -4.75 mm (relative to bregma and dural surface).
- 7.3 Inject 1 μL of 6-OHDA solution at a controlled rate.
- 7.4 Leave the needle in place for 5 minutes post-injection to minimize backflow.
- 7.5 Slowly retract the needle from the brain.

8 Needle Retraction and Closure:

- 8.1 Close the incision with sutures or surgical glue.
- 8.2 Clean the surgical area with sterile saline.

Postoperative Care

9 Immediate Postoperative Care:

- 9.1 Administer Carprofen (5 mg/kg, subcutaneous) as a postoperative analgesic.
- 9.2 Place the mouse in a warm recovery area and monitor until fully awake.

10 Continued Analgesia and Monitoring:



- 10.1 Provide a second dose of Carprofen 18-24 hours after surgery.
- 10.2 Monitor the animal daily for signs of pain, distress, or infection.
- 10.3 Record body weight and behavior to assess recovery.
- 11 Special Postoperative Care for 6-OHDA Lesion Model:
 - 11.1 Provide sweetened milk (1:3 dilution in tap water) in the home cage to support recovery and maintain body weight.
 - 11.2 Monitor the animal daily for one week for any signs of distress or motor deficits.
 - 11.3 Histological or behavioral assessments should be conducted at least two weeks post-injection to evaluate the efficacy of the lesion.