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Step by Step from Environmental Samples to Preservation Vials

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Protocol status: Working

We use this protocol and it's working

Created: April 05, 2023



Last Modified: September 13, 2023

Protocol Integer ID: 80039

Keywords: microbes isolation from environmental sample, microbes isolation, environmental samples to preservation vial, environmental sample, preservation vial, lab collection, isolation, sample, lab manager

Abstract

This protocol shows how to generate a MICROBES ISOLATION FROM ENVIRONMENTAL SAMPLES into LAB COLLECTION. Please follow the instructions and do not modify the protocols by yourself. If any changes are needed please inform the lab manager or admin.

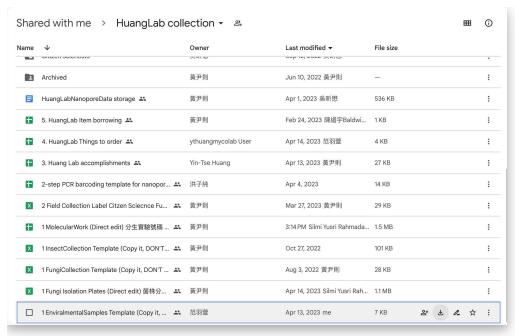
Troubleshooting



Isolate Primary Culture from Environmental Samples

1 Once you received your environmental samples.

Please catalog them in your Environmental Samples spreadsheet.

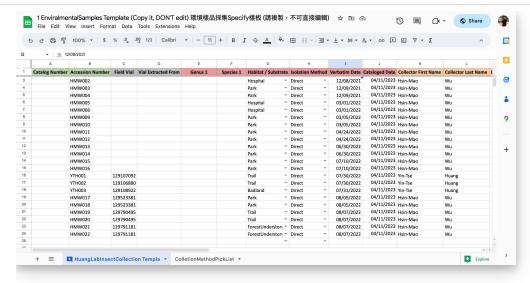


Make a copy of the template for yourself. The template "1 EnvironmentalSamples Template" marked with blue box is available in HuangLab Collection.

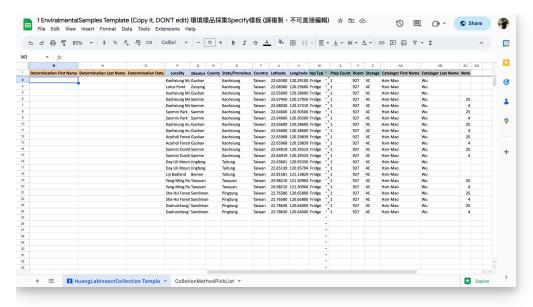
Fill in the columns of the spreadsheet step by step (As shown in the following steps 2.1-2.15)

- *In the template, some template data are there for your reference, please **delete them in your copy** and fill in only your data.
- * Remember to scroll to the right and fill in as much as information you can.





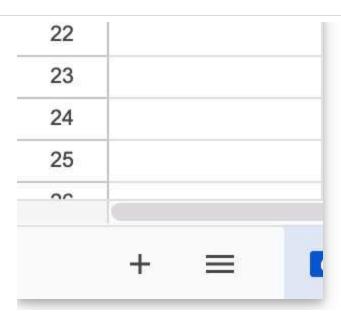
Column in the left



Scrolling to the right and fill them in

2.1 Leave the Catalog Number (Column A) blank. The Catalog Number will be generated automatically from Specify. Copy and paste it directly after the upload.

A
Catalog Number



2.2 Accession Number (Column B) is for your own reference, you can name your sample as you want, there is no specific format for this.



В **Accession Number HMW002 HMW003 HMW004 HMW005 HMW006 HMW009 HMW010 HMW011 HMW012 HMW013 HMW014 HMW015 HMW016** YTH001 YTH002 YTH003 **HMW017 HMW018 HMW019**

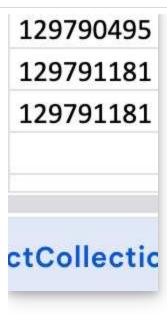




2.3 Field Vial (Column C) is the iNaturalist code. You only need to fill this column when you got the samples from citizen scientists.

С
Field Vial
129107092
129106880
129108922
129523381
129523381
129790495

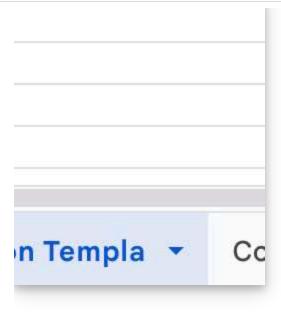




2.4 Leave the Vial Extracted From (Column D) blank.

D
Vial Extracted From





2.5 For Column E, F, G, G: If your samples are **SOIL SAMPLES**, please follow this example.



E	F	G	Н
Genus 1	Species 1	Habitat / Substrate	Isolation Method
		Hospital	Direct
		Park -	Direct
		Park -	Direct
		Hospital	Direct
		Hospital	Direct
		Park -	Direct
		Park ~	Direct
		Trail	Direct
		Trail	Direct
		Badland	Direct
		Park ~	Direct
		Park	Direct
		Trail	Direct
		Trail	Direct
		ForestUnderstory *	Direct
		ForestUnderstory *	Direct
		_	,

This is the example for Soil Samples.

Genus 1 & Species 1: Leave it blank.
Habitat/Substrate: What is the environment you collected your samples.

Isolation: How you isolate your samples (the default is Direct).

If your samples are **ENVIRONMENTAL SAMPLES**, please follow this example.



E	F	G	Н
Genus 1	Species 1	Habitat / Substrate	Isolation Method
Anisandrus	ursulus	Ficus fistulosa	Direct
Euwallacea	interjectus	Ficus fistulosa	Direct -
Planiculus	bicolor	Ficus fistulosa	Direct -
Euwallacea	fornicatus	Mucuna macrocarpa	Direct -
Corticoid		Dried fruiting bodies	Direct -
Phellinus		Dried fruiting bodies	Direct
Oligoporus		Dried fruiting bodies	Direct -
Phellinus		Dried fruiting bodies	Direct
Agaricales		Dried fruiting bodies	Direct
Phellinus		Dried fruiting bodies	Direct
Polyporus	squamosus	Dried fruiting bodies	Direct
Phellinus		Dried fruiting bodies	Direct
Oligoporus		Dried fruiting bodies	Direct -
Agaricales		Dried fruiting bodies	Direct
Polyporus		Dried fruiting bodies	Direct -
Oligoporus		Dried fruiting bodies	Direct •
Irpex	laceratus	Dried fruiting bodies	Direct -
Hyphodontia		Dried fruiting bodies	Direct
Cerrena	zonata	Dried fruiting bodies	Direct -
Trametes		Dried fruiting bodies	Direct
Trichaptum		Dried fruiting bodies	Direct
Pleurotus		Dried fruiting bodies	Direct
Polyporoid		Dried fruiting bodies	Direct
Polyporales		Dried fruiting bodies	Direct
Laetiporus		Dried fruiting bodies	Direct
Oligoporus		Dried fruiting bodies	Direct

This is the example for Environmental Samples (e.g., ambrosia & bark beetles, fruiting bodies, etc).

Genus 1 & Species 1: Fill in the scientific name of your samples (unable to identify just leave it blank).

Habitat/Substrate: What is the environment you collected your samples (e.g., the host plant for the ambrosia & bark beetles).

Isolation: How you isolate your samples (the default is Direct).

2.6 Fill in the Verbatim Date (Column I) with the date you COLLECTED the samples. Fill in the Cataloged Date (Column J) with the date you UPLOADED the data to Specify.

1	J
Verbatim Date	Cataloged Date
12/08/2021	04/11/2023
12/09/2021	04/11/2023
12/09/2021	04/11/2023
03/01/2022	04/11/2023
03/01/2022	04/11/2023
03/05/2022	04/11/2023
03/05/2022	04/11/2023
04/24/2022	04/11/2023
04/24/2022	04/11/2023
06/30/2022	04/11/2023
06/30/2022	04/11/2023
07/10/2022	04/11/2023
07/10/2022	04/11/2023
07/30/2022	04/11/2023
07/30/2022	04/11/2023
07/31/2022	04/11/2023
08/05/2022	04/11/2023
08/05/2022	04/11/2023
08/07/2022	04/11/2023

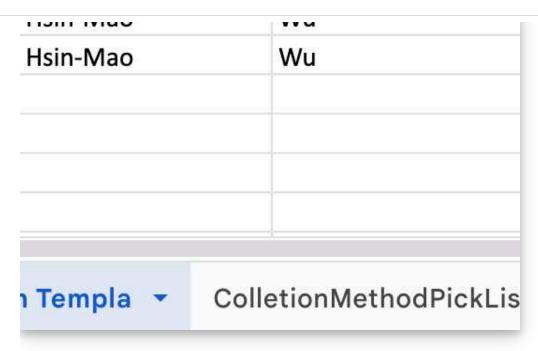
08/07/2022	04/11/2023
08/07/2022	04/11/2023
08/07/2022	04/11/2023

2.7 Collector First Name (Column K) and Collector Last Name (Column L) Fill in the name of the collector.



K	L
Collector First Name	Collector Last Name
Hsin-Mao	Wu
Yin-Tse	Huang
Yin-Tse	Huang
Yin-Tse	Huang
Hsin-Mao	Wu
Hsin-Man	Wii





2.8 Fill in these columns when the sample is identified with a scientific name, remember to fill Column E and F (Genus and Species names) as well. If not, leave it blank. Fill in the date you did the determination (identification).



M	N	0
Determination First Name	Determination Last Name	Determination Date
▼		

2.9 Fill in these columns with the geographical information of your samples. Leave it blank, when you don't have the information or the materials is outsourced lacking this info.



P	Q	R	S	Т	U	V
Locality	District	County	State/Pronvince	Country	Latitude	Longitude
Kaohsiung Mı	Gushan		Kaohsiung	Taiwan	22.65500	120.29100
Lotus Pond	Zuoying		Kaohsiung	Taiwan	22.68300	120.29600
Kaohsiung Mu	Gushan		Kaohsiung	Taiwan	22.65600	120.28600
Kaohsiung Me	Sanmin		Kaohsiung	Taiwan	22.67450	120.27950
Kaohsiung Me	Sanmin		Kaohsiung	Taiwan	22.68030	120.27310
Sanmin Park	Sanmin		Kaohsiung	Taiwan	22.64600	120.30500
Sanmin Park	Sanmin		Kaohsiung	Taiwan	22.64600	120.30500
Kaohsiung mu	Gushan		Kaohsiung	Taiwan	22.65600	120.28600
Kaohsiung mu	Gushan		Kaohsiung	Taiwan	22.65600	120.28600
Aozihdi Fores	Gushan		Kaohsiung	Taiwan	22.65968	120.29839
Aozihdi Fores	Gushan		Kaohsiung	Taiwan	22.65968	120.29839
Sanmin DunSi	Sanmin		Kaohsiung	Taiwan	22.64919	120.29523
Sanmin DunSi	Sanmin		Kaohsiung	Taiwan	22.64919	120.29523
Day Lili Moun	Jingfeng		Taitung	Taiwan	22.63601	120.95336
Day Lili Moun	Jingfeng		Taitung	Taiwan	22.65139	120.95794
Liji Badland	Beinan		Taitung	Taiwan	22.81581	121.13829
Yang-Ming Pa	Taoyuan		Taoyuan	Taiwan	24.98210	121.30904
Yang-Ming Pa	Taoyuan		Taoyuan	Taiwan	24.98210	121.30904
Sha-Hsi Fores	Sandima	n	Pingtung	Taiwan	22.76580	120.65800
Sha-Hsi Fores	Sandima	n	Pingtung	Taiwan	22.76580	120.65800
Dashuichong	Sandima	n	Pingtung	Taiwan	22.78630	120.64000
Dashuichong	Sandima	n	Pingtung	Taiwan	22.78630	120.64000
			MethodPickList			

2.10 The values of these columns were set using a formula. You only need to choose the **Prep Type** (Column W), which is the way you preserve your samples. *Inform admin when other Prep Type is needed.

W		X	Υ	Z
Prep Type	-	Prep Count	Room	Storage
Fridge	~	1	927	4C
Fridge	-	1	927	4C
Fridge	~	1	927	4C
Fridge	•	1	927	4C
Fridge	-	1	927	4C
Fridge	~	1	927	4C
Fridge	~	1	927	4C
Fridge	•	1	927	4C
Fridge	~	1	927	4C
Fridge	-	1	927	4C
Fridge	~	1	927	4C
Fridge	•	1	927	4C
Fridge	_	1	927	4C
Fridge	-	1	927	4C
Fridge	~	1	927	4C
Fridge	V	1	927	4C
Fridge	~	1	927	4C
Fridge	-	1	927	4C
Fridge	-	1	927	4C
Fridge	*	1	927	4C

•			
*			
•			
Fridge	1	927	4C
Fridge 🔻	1	927	4C
Fridge 🔻	1	927	4C

2.11 Fill in the name (Column AA and AB) of who uploaded the data to Specify.



AA	AB				
Cataloger First Name	Cataloger Last Name				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Mao	Wu				
Hsin-Man	Wii				



HUIH IVIGO	***	
Hsin-Mao	Wu	
Hsin-Mao	Wu	

2.12 You can leave any informative notes of your samples. If not, leave it blank.



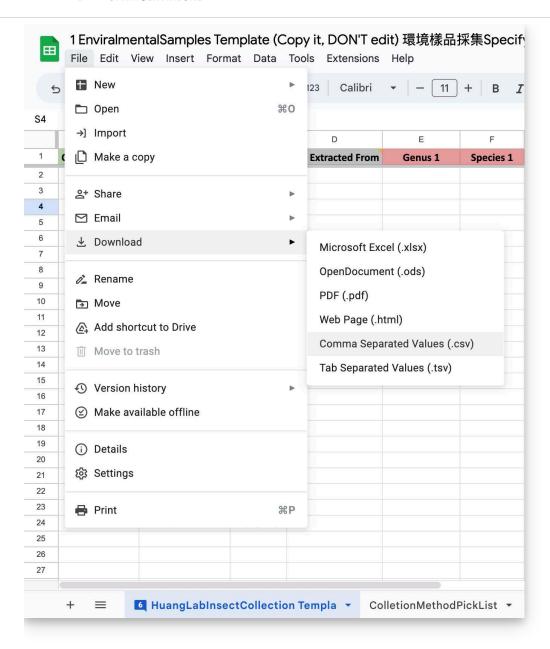
AC

Note

25
4
25
4

2.13 Done!! Download the spreadsheet as .csv format.





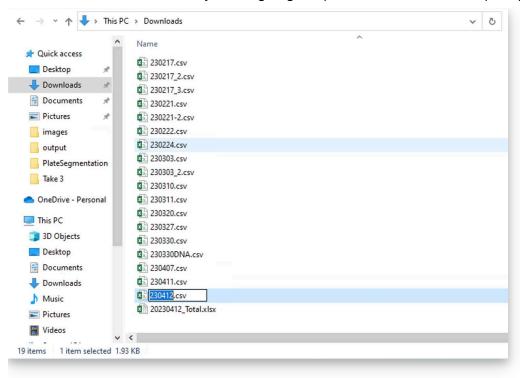
2.14 Delete the row with existed Catalog Number if any.

**Leave the not-uploaded rows only.

	A	В	С	D	Ε	F	G	н	1	J	K	L
1	Catalog Number	Accession Nu	Field Vial	Vial Extracte	Genus 1	Species 1	Habitat / Si	ub Isolation Me	Verbatim Da	Cataloged Da	Collector Fire	Collector Las
2	2023-000133	HMW002					Hospital	Direct	12/08/2021	04/12/2023	Hsin-Mao	Wu
3		HMW003					Park	Direct	12/09/2021	04/12/2023	Hsin-Mao	Wu
4		HMW004					Park	Direct	12/09/2021	04/12/2023	Hsin-Mao	Wu
5		HMW005					Hospital	Direct	03/01/2022	04/12/2023	Hsin-Mao	Wu
6		HMW006					Hospital	Direct	03/01/2022	04/12/2023	Hsin-Mao	Wu
7		HMW009					Park	Direct	03/05/2022	04/12/2023	Hsin-Mao	Wu



2.15 Rename the file with the date you are going to upload the datasheet to Specify.

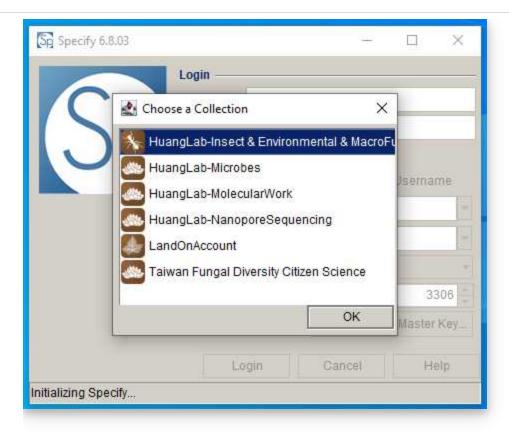


Upload your datasheet to Specify database

3 **The account setup is skipped**

> Login to the Specify database. Choose the first collection (HuangLab-Insect & **Environmental & MacroFungi).**





3.1 Go to the Workbench tab.





3.2 Click Import Data.

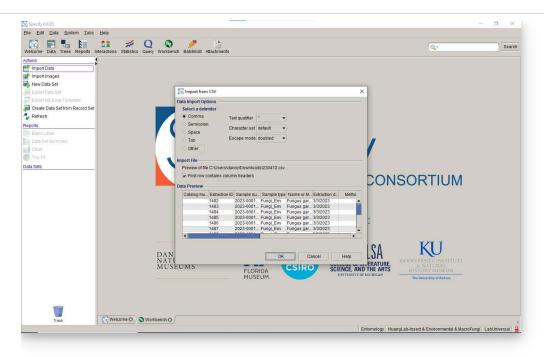


3.3 Choose the file you wanted to import (upload), and click **Open**.

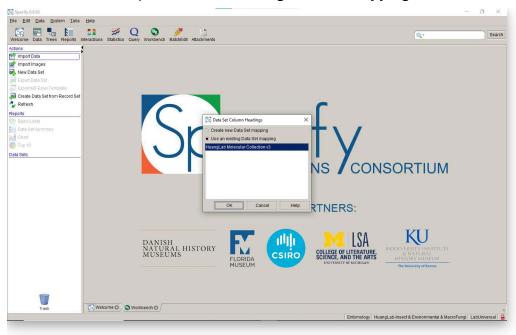


3.4 Let everything as default. Then, click **OK**.





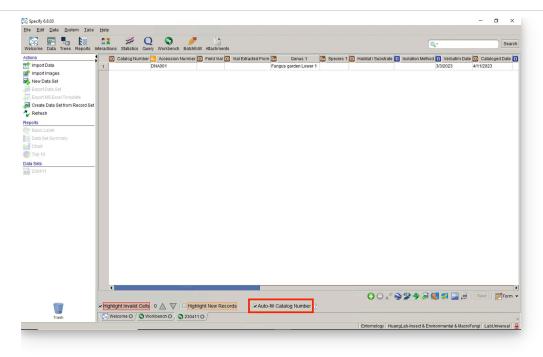
3.5 Select the second option: **Use an existing Data Set mapping**.



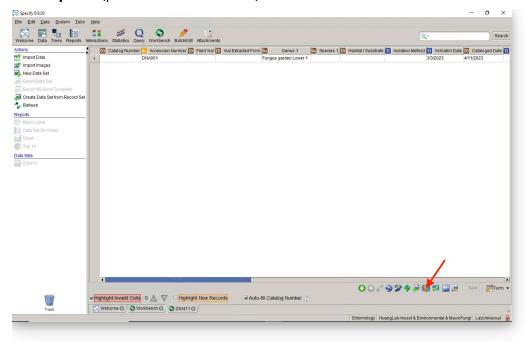
3.6 Check for any invalid cells (red highlighted cells).

Remember to tick Auto-fill Catalog Number (marked with red box).



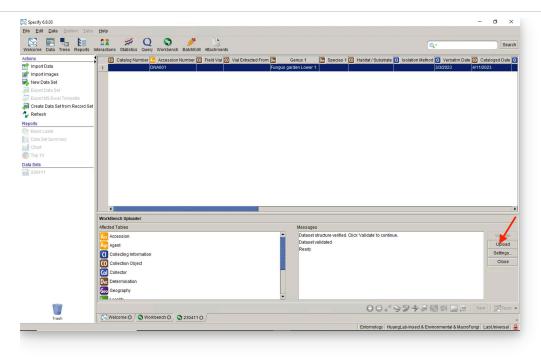


3.7 Click **Upload** (pointed with red arrow).

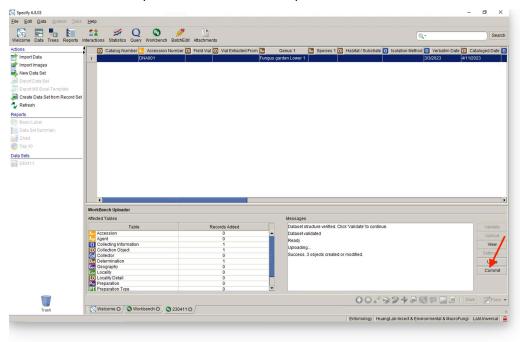


3.8 Click **Upload** again (pointed with red arrow).



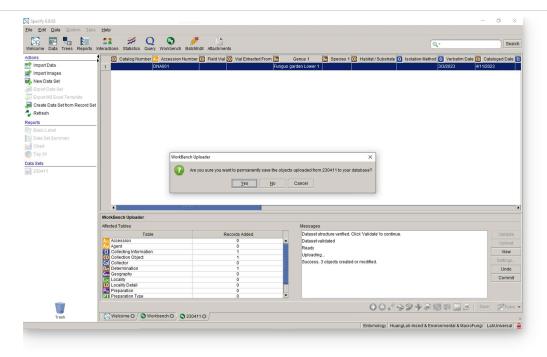


3.9 Then Click Commit (marked with red arrow).



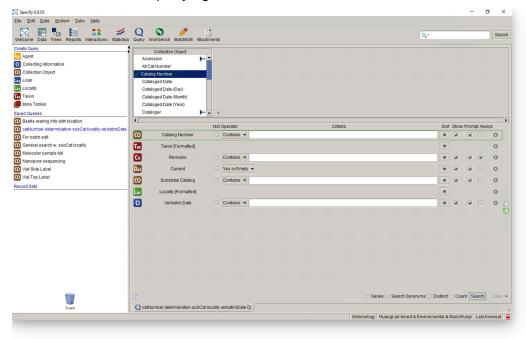
3.10 Click Yes to finish the upload progress.





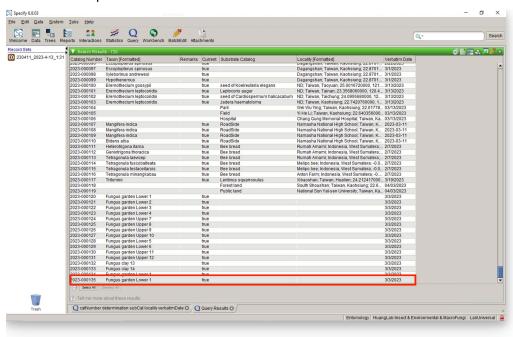
3.11 Now you can go to **Query** → **catNumber determination...**(under the Saved Queries in left panel) to check if the upload succeeded.

Click Search to start querying.





3.12 Ta-Da!! The upload was succesful.



**Remember to delete the Data Sets & Record Sets before closing the Specify.



3.13 Paste the **Catalog Number** of uploaded data back to your spreadsheet.



Isolate the microbes from your environmental samples

4 Prepare the equipment and materials first for sample isolation. Put them on another bench first.



- 1. Alcohol lamps
- 2. 95% ethanol cooling bottle
- 3. Beakers (small & large)
- 4. Tweezers
- 5. Scalpel with blade
- 6. Separator
- 7. Grafting tube
- 8. Lighter
- 9. Marker pen
- 10. Media
- 4.1 Sterilize the microbes isolation area (air and bench) using a 75% ethanol sprayer.
 - **Make sure the mist is emitted.**





Spray it a few times up to the air.



Spray it a few times onto the bench.



Wipe the bench unidirectionally using paper towels.

4.2 Now, move the equipment and materials for microbes isolation back to the sterilized area.



4.3 Before isolation, key the information about your isolation (how many plates you are going to isolate, what kind of media you will use, etc) into the 1 Fungi Isolation Plates (Direct edit) 菌株分離培養基編號 (直接編輯) spreadsheet first

Α	В		С		D		E	F		G	Н	1	
Plate numk 🔻	Project	Ŧ	Media	Ŧ	Isolation part	Ŧ	Insect_Env_Cataloge(=	Diluti	o =	Cataloge =	Cataloger =	Cataloged D; =	Notes
P701	Trichosporon project		DG18	*	Bird feces	-	2023-000007	0.25		Yu-Hsuan	Fan	1/5/2023	
P702	Trichosporon project		DG18		Bird feces	*	2023-000007	0.25	*	Yu-Hsuan	Fan	1/5/2023	
P703	Trichosporon project	*	DG18		Bird feces	*	2023-000007	0.25	*	Yu-Hsuan	Fan	1/5/2023	
P704	Trichosporon project	*	DG18		Bird feces	*	2023-000008	0.25	*	Yu-Hsuan	Fan	1/5/2023	
P705	Trichosporon project	*	DG18		Bird feces	~	2023-000008	0.25	-	Yu-Hsuan	Fan	1/5/2023	
P706	Trichosporon project	*	DG18		Bird feces	w	2023-000008	0.25	*	Yu-Hsuan	Fan	1/5/2023	
P707	General Isolation	*	PDA		Fungus	*	2023-000009	Direct	-	Yu-Hsuan	Fan	1/6/2023	
P708	General Isolation		PDA		Fungus	*	2023-000009	Direct	*	Yu-Hsuan	Fan	1/6/2023	
P709	General Isolation	*	PDA	*	Fungus	~	2023-000010	Direct	-	Yu-Hsuan	Fan	1/6/2023	
P710	General Isolation	*	PDA		Fungus	w	2023-000010	Direct		Yu-Hsuan	Fan	1/6/2023	
P711	General Isolation	*	PDA		Fungus	*	2023-000011	Direct	*	Yu-Hsuan	Fan	1/6/2023	
P712	General Isolation	*	PDA		Fungus	*	2023-000011	Direct	*	Yu-Hsuan	Fan	1/6/2023	
P713	General Isolation		PDA		Fungus	*	2023-000012	Direct	*	Yu-Hsuan	Fan	1/6/2023	
P714	General Isolation	*	PDA		Fungus	*	2023-000012	Direct	*	Yu-Hsuan	Fan	1/6/2023	
P715	General Isolation	*	PDA		Fungus	*	2023-000013	Direct	*	Yu-Hsuan	Fan	1/6/2023	
P716	General Isolation		PDA		Fungus	~	2023-000013	Direct	~	Yu-Hsuan	Fan	1/6/2023	
P717	General Isolation	*	PDA		Fungus	*	2023-000014	Direct	-	Yu-Hsuan	Fan	1/6/2023	
P718	General Isolation	w	PDA		Fungus	*	2023-000014	Direct	*	Yu-Hsuan	Fan	1/6/2023	
P719	General Isolation	*	PDA		Fungus	*	2023-000015	Direct	*	Yu-Hsuan	Fan	1/6/2023	
P720	General Isolation	*	PDA		Fungus	-	2023-000015	Direct	-	Yu-Hsuan	Fan	1/6/2023	
P721	General Isolation	*	PDA		Fungus	*	2023-000016	Direct	*	Yu-Hsuan	Fan	1/6/2023	

Plate number: The number is **auto-generated**.

Project: Choose an existing one or create one for yourself for latter identified.

Media: The media to isolate your microbes.

Isolation part: The part of the environment samples you isolated your microbes. Insect_Env_Cataloged #: The catalog number of the environmental samples. Dilution: The dilution factor of this particular plate for isolation (e.g., 0.001, 0.01, or Direct).

Cataloger First Name & Cataloger Last Name: The name of who uploads the data. Cataloged Date: The date on which the data is uploaded.

Notes: Any information you wanted to remark on the certain plate. Leave it when you didn't have any notes.

4.4 Write the plate information on the downside of the plate following the example photo shown below.



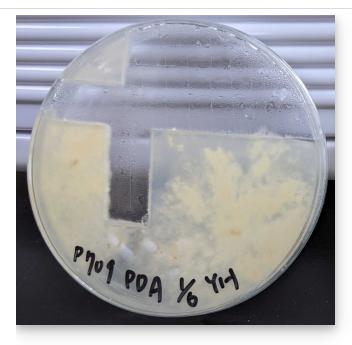
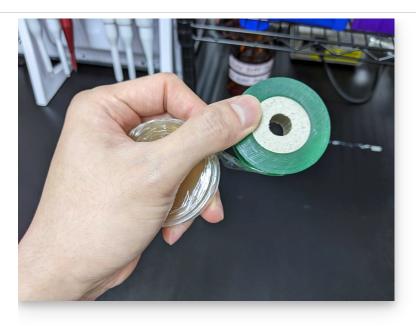


Plate number, Media, Date, Your name

- 4.5 Make sure that you sterilize every tool before using them for each isolation:
 - 1. Dip the scalpel, tweezer, or spreader into 95% ethanol cooling bottle, and flame it immediately.
 - 2. Repeat the step(dip and flame) **THREE times** for sterilization of equipment.
- 4.6 Now, you can start to isolate microbes from your environmental samples. We called them primary cultures.
 - **Isolation method varied depend on the type of sample, check published papers or consult with seniors if needed**
- 4.7 After finishing all the isolation, seal the plates with grafting tape for 2 rounds.





4.8 Pack all the plates using a PP bag (no. 8, 10, or 12 depending on how many plates you have). Incubate the plates in your slot in the incubator for 7~21 days (depending on the growth rate of the microbes).

no. 8: 8 plates

no. 10: 24 or 32 plates

no. 12: 48 or 56 plates





Flip the PP bag over when keeping it in the incubator. The cover of the plates needed to be facing downwards to prevent the microbes from drowning by the water vapors.

4.9 Check the growth condition of the primary cultures every day. **Make sure they are not overgrown.**

Once they are matured, you can start to subculture them.

Record and upload the data of primary cultures

- 5 1. Examine the microbes colonies under a dissecting microscope.
 - 2. Choose your desired colonies, which are your targeted colonies.
 - 3. Mark the targeted colonies with their later corresponding secondary plate number.
 - 4. If there is more than one morphotype targeted colony in a certain plate, you need to record them into a distinct plate number, e.g., **TWO morphotypes will have TWO distinct secondary plate numbers** from the same primary plate number.





5.1 Record all the data of primary cultures to the second worksheet: **Secondary Isolate PDA** (in 1 Fungi Isolation Plates (Direct edit) 菌株分離培養基編號 (直接編輯)).
Fill in all the columns following the photos and instructions shown below.

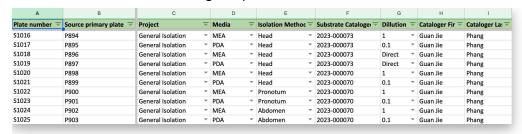
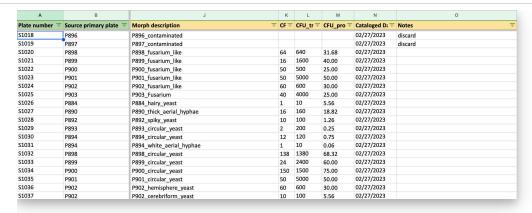


Plate number: It is auto-generated.

Column B to Column I: You can copy and paste directly from the Column A to Column H of the first worksheet **Primary Isolate MEA**; (Hint: **GREEN columns to GREEN columns**)





Morph description: This is generated automatically by the formula. Replace **YOUR DESCRIPTION** with what you do observe on your targeted colony under a dissecting microscope. Type **contaminated** if you got others than your target or all the colonies were overgrown by common molds.

CFU: The number of colonies of your target in a plate.

CFU_true: It is auto-filled based on the dilution factor of each plate.

CFU_proportion: The proportion of your targeted colony in a certain plate (i.e. what percentage of your target among all the observed colonies on the plate).

Cataloged Date: The date you do the subculture and upload the data.

Notes: Any information you think is important to record. If you have **contaminated** in Morph description, please type discard here.

5.2 Record or take a photo of the order of your plates that are ready for scanning. **6** plates as a group, so you can refer back during the photo editing.





5.3 Before scanning plate, you need to sterilize the environment and scanner glass using paper towels with 75% ethanol.





5.4 Wipe the scanner glass using paper towels with dishwashing detergent (this prevents water condensation on the glass).







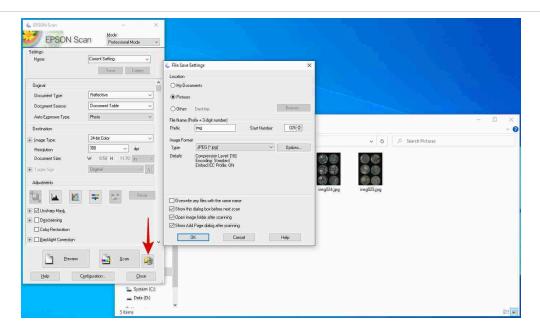
5.5 Keep the order of the plates, remove the plates cover, and put the plates upside down onto the scanner glass.

Do not change the order before and after scanning

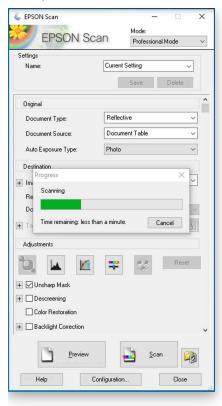


5.6 Search and open **EPSON Scan** software. Choose **Professional mode**. Go to the **File Save Settings** (red arrow), and make sure the image format is .jpg (follow all the settings as the photo below shown).



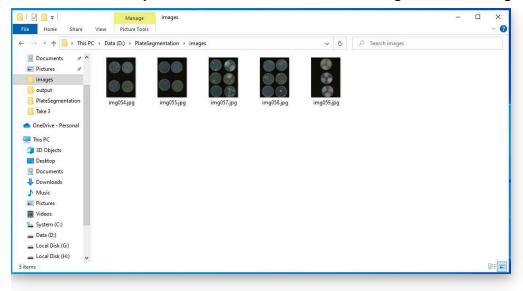


5.7 Then, click **OK**. (It will start scanning like the photo shown below).

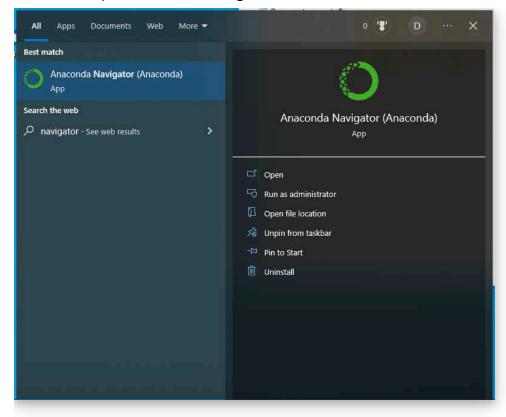




5.8 Move the files from your Pictures folder to the D:\PlateSegmentation\images

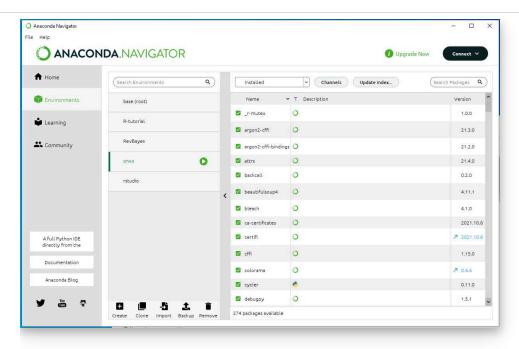


5.9 Search and open **Anaconda Navigator** software.

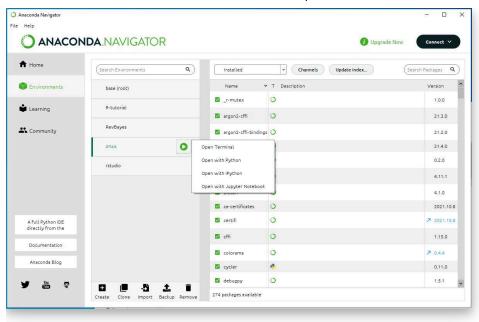


You will see the interface as the photo shown below.



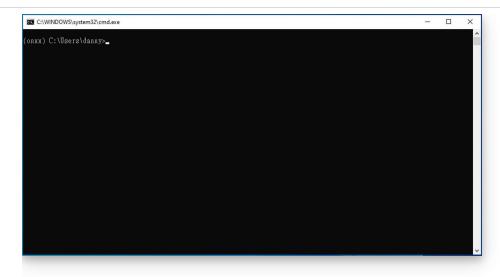


5.10 Select **onxx** under **Environments** tab (as the photo shown below).

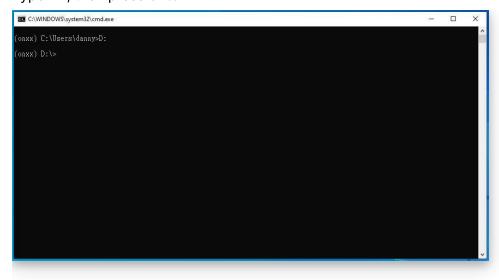


Left click the play button. Select Open Terminal.



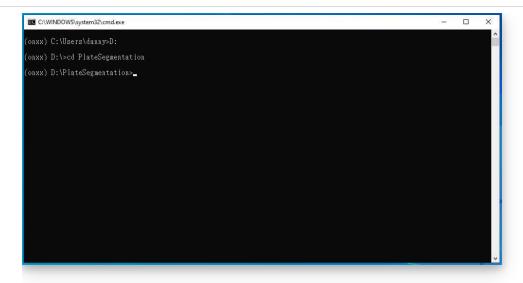


5.11 Follow the commands, and you will have the similar output as the photos shown below. Type **D:**, then press **enter**

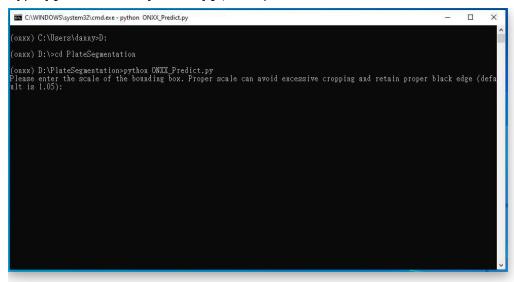


Type cd PlateSegmentation , then press enter

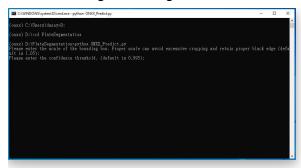




Type **python ONXX_predict.py** , then press **enter**



Press enter again and again. You will see a _ flashing.





```
(axx) C:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\text{0}:\Usera\danayo\
```

Then, a new command line comes out, which represent the program execution was complete.

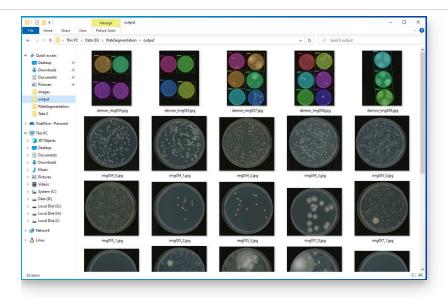
```
CAWINDOWS system 32 cmd.exe

(onxx) C: VUsers \danny>D:

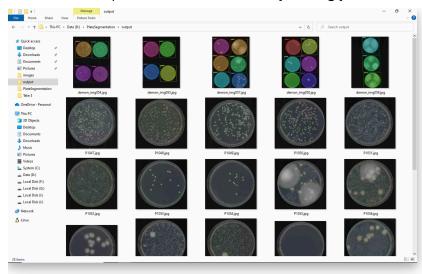
(onxx) D: \PlateSegmentation > python ONXX Predict.py
Please enter the scale of the bounding box. Proper scale can avoid excessive cropping and retain proper black edge (default is 1.05):
Please enter the confidence threshold. (default is 0.995):
Model loaded successfully
Inputs: ('input_image_meta']
Outputs: ('input_image_meta')
Outputs: ('input_image_', 'input_image_meta')
Outputs: ('incnat_rpn_box')
img054.jpg found, predicting...
4 instances detected in img054.jpg
img055.jpg found, predicting...
4 instances detected in img055.jpg
img057.jpg found, predicting...
6 instances detected in img057.jpg
img058.jpg found, predicting...
6 instances detected in img058.jpg
img059.jpg found, predicting...
6 instances detected in img058.jpg
img059.jpg found, predicting...
5 instances detected in img058.jpg
img059.jpg found, predicting...
6 instances detected in img057.jpg
img058.jpg found, predicting...
5 instances detected in img058.jpg
img059.jpg found, predicting...
5 instances detected in img059.jpg
(onxx) D:\PlateSegmentation>
```

5.12 The output files are in the **D:\PlateSegmentation\output**.





Rename the output files with their corresponding plate number.



Upload them to the HuangLab Collection/Plate picture/20XX



Subculture the primary cultures



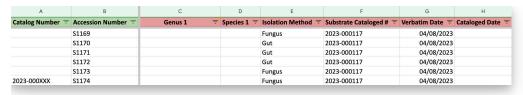
6 Follow 5 go to step #4.4 to 5 go to step #4.9 for the subculture of primary cultures.

Now, we called them secondary cultures.

You may need to keep the primary cultures for a while before the secondary plate are all secured.

Ready for preservation

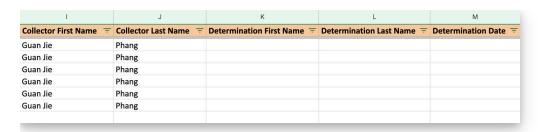
- After 7-21 days (depending on the growth rate of your targeted microbes), the secondary cultures are ready to be preserved. If the secondary cultures were contaminated, please redo the subculture from the primary cultures.
- 7.1 Before prepare preservation vials, the **For Specify cateloging COPY to MODIFY!!!** worksheet need to be filled.



Catalog Number: It is auto-generated from Specify, fill in after you upload the data. Column B, Column E to Column G: It is synchronized from the **Secondary Isolate PDA** worksheet.

Genus 1 & Species 1: Only fill in when you can identify them.

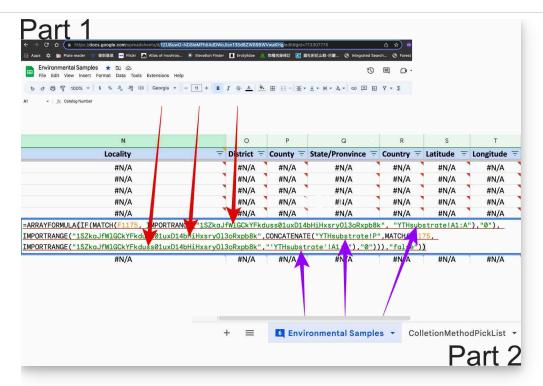
Cataloged Date: Same as previous spreadsheet.



Collector First Name & Collector Last Name: Same as previous spreadsheet. Column K to Column M: Same as previous spreadsheet.

There is 2 part for all the locality information, you need to replace the each information: spreadsheet code (red arrow pointed), worksheet name (purple arrow pointed) with your own spreadsheet information.

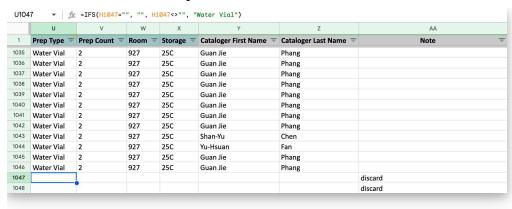




Part 1: Replace the red arrow pointed region of the written formula with your own spreadsheet code (highlighted region).

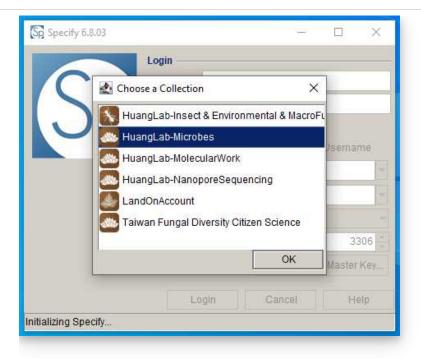
Part 2: Replace the purple arrow pointed region of the written formula with your own worksheet name (blue shaded region).

If the Cataloged Date is filled, the Column U to Column Z is auto-filled with the written formula. Do not change it unless it needed. The Column AA is synchronized with the Column O of the **Secondary Isolate PDA** worksheet.



7.2 Once the spreadsheet is filled, download the data with .csv format. Open **Specify**, go to **HuangLab-Microbes** collection.





- 7.3 Follow **5** go to step #3.1 to **5** go to step #3.13 for preservation vials data upload.
- 7.4 For each Catalog Number, you must prepare 2 water vials and 2 glycerol vials. You also need a 2 ml microtube with lysis mixture and beads (for details, check **DNA extraction** protocols).

Each tube needed a sticker with its corresponding Catalog Number on top of the cap. The label should be written with a black color marker pen or a technical pen alternative.

Please follow the example in the photo shown below.





For water vials: either tube must have a RED underline. For glycerol vials: both tubes must have a BLUE underline.

The lysis mixture microtubes that are ready for use will look like this.





7.5 Operate UV sterilization of the laminar flow for 15 minutes. After UV sterilization, you need to ethanol-sterilize the materials that are going into the laminar flow.

Light up the alcohol lamp and flame the material.

The materials you need are the same as go to step #4 , but you only need a scalpel instead of a tweezer and a spreader.

Culture preservation

- 8 Preserve the plates with one water and one glycerol vial for one colony. The rest is for DNA extraction. Make sure you flame-sterilize the scalpel every time before slicing the agar.
 - **The details of culture preservation are not provided here.**
- 8.1 Store the preservation vials in their corresponding freezing box.



For water vials, you can keep them directly.

For glycerol vials, you need to keep them in a freezing container and then freeze them inside a -20°C fridge for 1 day before keeping them in the -80°C fridge collection. **The freezing boxes of water vials are kept in the steel cabinet in front of Prof. Huang's office; the freezing boxes of glycerol vials are kept in the -80°C fridge outside N927**



The tubes are needed to be kept in the order shown above, but not the other way around.