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Version 2

# Standard DAB Staining for Free-floating Fixed NHP Brain Tissue V.2

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We use this protocol and it's working

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### Abstract

This protocol details the procedure for immunohistochemical 3,3'-Diaminobenzidine (DAB) staining of freefloating fixed brain tissue sections using the avidin/biotin ABC complex.

This protocol has been tested with free-floating non-human primate (NHP) and rodent (mouse, rat) brain tissue that has been fixed (10% formalin or 4% paraformaldehyde), cryoprotected (sucrose or glycerol gradients), and cryo-sectioned → ← 20 µm - → ← 50 µm .

#### Guidelines

When using 6 well tissue culture plates [Falcon, 353046] to react individual sections, you will need 2+ mL solutions for *each* well plate.

When using circular staining nets [e.g., Brain Research Laboratories #4115] to react multiple series of sections, you will need **50 mL** solutions for *each*.



#### **Materials**

#### Tissue:

Brain tissue sections (20 - 50 μm).

#### **Materials/Equipment:**

- Tissue culture plates or circular staining nets
- Orbital shaker
- Fume hood
- Nitrile Gloves
- Glass slides (charged or subbed)

#### Reagents:

- Phosphate-buffered saline (PBS)
- Hydrogen Peroxide: H<sub>2</sub>O<sub>2</sub> (3% or 30%)
- Distilled water: dH<sub>2</sub>O
- Primary Antibody
- Secondary Antibody (to match the host of the primary antibody)
- Normal Serum Blocking Solution (e.g., Normal Horse Serum, S-2000-20; Normal Goat Serum, S-1000-20)
- Vectastain Elite ABC Peroxidase Kit (Standard) (PK-6100) (Vector Laboratories)
- ABC-HRP Kit

#### Examples:

Vectastain ABC-HRP Kit, Peroxidase (Mouse IgG) (PK-4002, Vector Laboratories)

Vectastain ABC-HRP Kit, Peroxidase (Rabbit IgG) (PK-4001, Vector Laboratories)

DAB Substrate Kit

### Examples:

Peroxidase (HRP) with Nickel (3,3'-diaminobenzidine) (SK-4100) (Vector Laboratories) ImmPACT DAB (SK-4105)

# **Troubleshooting**



# Safety warnings

Use appropriate care when using hydrogen peroxide (reactive, can cause skin/eye damage) and DAB (suspected carcinogen). Collect DAB solution for chemical waste disposal.



### Part I (Day 1)

3h

Bring tissue to Room temperature in buffer (e.g., Phosphate buffered saline, PBS) on an orbital shaker for 30 minutes. 00:30:00.

30m

Prepare **Peroxide Solution (0.3 - 3 % H\_2O\_2)** in  $dH_2O$ .

5m

E.g., for  $\perp$  10 mL 0.3% H<sub>2</sub>O<sub>2</sub> use:

- 4 100 µL 30% H<sub>2</sub>O<sub>2</sub>
- 4 9900 μL dH<sub>2</sub>O
- Prepare **Blocking Serum Solution** (e.g. Normal Horse, Normal Goat Serum) using a serum that matches the **host of the secondary antibody** (e.g. Normal Horse Serum for a Horse anti-Mouse secondary, Normal Goat Serum for a Goat anti-Rabbit secondary).

  E.g., in und buffer (PBS) add:

5m

Δ 150 μL normal serum (or 3 drops of normal serum if using an ABC kit, e.g.
 Vectastain ABC-HRP Kit, Peroxidase Mouse IgG PK-4002, Rabbit IgG PK-4001)

5m

4 Prepare **Primary Antibody Solution** at the appropriate dilution in buffer (e.g., 1:1000 in PBS).

15m

Rinse in buffer (e.g. PBS) on a shaker at Room temperature: 3 × 3-5 minutes.

Quench endogenous peroxide in **Peroxide Solution (0.3 - 3 % H<sub>2</sub>O<sub>2</sub>)** on a shaker at Room temperature: **30 - 60 minutes.** © 00:30:00 - © 01:00:00

1h

7 Rinse in buffer (e.g. PBS) on a shaker at Room temperature : 3 × 3-5 minutes.

Incubate in **Blocking Serum Solution** on a shaker at RT: **1 hour**.

15m

**(5)** 00:03:00 **- (6)** 00:05:00

8

1h

DO NOT RINSE after blocking serum.



9 Incubate in **Primary Antibody Solution** on a shaker at 4 °C Overnight, or 20h longer (20 - 72 hours depending on the antibody).

## Part II (Day 2)

4h

10 Bring tissue (in the **Primary Antibody Solution**) to Room temperature on a shaker (**30 - 60 minutes**). ( ) 00:30:00 | - ( ) 01:00:00

30m

11 Prepare ABC Solution in buffer (e.g. PBS) (at least 30 minutes before use).

5m

**(?)** 00:30:00

5m

- 12 Prepare **Secondary Antibody Solution** (1:200) in buffer (e.g. PBS).
  - In 4 10 mL buffer add:
  - Δ 150 μL (= 3 drops of normal serum from a Vector Labs kit) of normal serum (matched to the host of your secondary antibody)
  - $\perp$  50  $\mu$ L (= 1 drop secondary antibody from a Vector Labs kit) of biotinylated secondary antibody (matched to the host of your primary Antibody )
- 13 **Rinse** in buffer (e.g. PBS) on a shaker at **§** Room temperature : **3 × 3-5 minutes.** (a) 00:03:00 - (b) 00:05:00 .

15m

- 14 Incubate in **Secondary Antibody Solution** on a shaker at 8 Room temperature : **30** minutes. (5) 00:30:00 .

**Rinse** in buffer (e.g. PBS) on a shaker at Room temperature: 3 × 3-5 minutes.

30m

15

15m

**♦** 00:03:00 **- ♦** 00:05:00 .

16 Incubate in **ABC Solution** on a shaker at **§** Room temperature : **60 minutes.** 

1h

**(:)** 01:00:00 .



17 **Rinse** in buffer (e.g. PBS) on a shaker at **&** Room temperature : **3 × 3-5 minutes.** 15m **♦** 00:03:00 **- ♦** 00:05:00 . 18 Prepare **Peroxide Substrate Solution** in dH<sub>2</sub>O. 5m To use the Vector Labs DAB Peroxidase Substrate Kit (SK-4100): In  $\perp$  5 mL dH<sub>2</sub>O: 2 drops Reagent 1 4 drops Reagent 2 2 drops Reagent 3 • [optional] 2 drops of Reagent 4 (Nickel) if a black reaction product is desired Note: Mix well before use. Use immediately. 19 Incubate in **Peroxide Substrate Solution on** a shaker at Room temperature: 6m (\*) 00:03:00 **-** (\*) 00:06:00 . **Note:** Watch the tissue closely to avoid high background staining. 20 **Rinse** in buffer (e.g. PBS) on a shaker at **\$\\$\\$** Room temperature : **3 × 3-5 minutes.** 15m **♦** 00:03:00 **- ♦** 00:05:00 . 21 Mount tissue on glass slides (subbed or charged) in 1:8 buffer in dH<sub>2</sub>O and let air dry. 22 Rinse slides with dH<sub>2</sub>O and let air dry (preferably in a hood). 23 Coverslip clean and dry slides with Cytoseal 60 (Thermo Fisher #830-16).

### Protocol references

https://vectorlabs.com/productattachments/protocol/VL\_SK-4100\_UserGuide\_LBL02267.pdf https://vectorlabs.com/products/vectastain-elite-abc-hrp-kit-standard