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Stable Cell Line Generation – RAW 264.7

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Protocol status: Working

We use this protocol and it's working

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Abstract

This protocol details steps taken to create stably expressing cell lines in RAW 264.7 cells.

Materials

DMEM (Thermo Fisher Scientific, 11965-092)
FBS (Thermo Fisher Scientific, 16140-071)
Penicillin/Streptomycin (Thermo Fisher Scientific, 15140122)
CellStripper (Corning, 356230)
Lipofectamine 2000 (Invitrogen, 11668019)
FuGene HD (Promega, E2311)
Opti-Mem (Thermo Fisher Scientific, 31985062)
Puromycin (Gibco A11138-03)
Blasticidin (Invivogen ant-bl)



Day 0

- 1 Plate 2.5×10^5 RAW 264.7 cells per well in a 6-well dish.

Day 1

30m

- 2 Transfect cells using Lipofectamine 2000 or FuGene HD. Ensure to use a 1:2 ratio of Piggybac transposon plasmid:gene-of-interest plasmid for a total of 1 uG DNA.

2.1 For Lipofectamine 2000 (HALO-LRRK2):

30m

In Tube 1 combine  150 μ L Opti-MEM and  8 μ L Lipofectamine 2000 then incubate for  00:05:00 . In Tube 2 combine  150 μ L Opti-MEM and 1 uG total DNA (see above) then incubate for  00:05:00 . Combine tube 1 and tube 2. Incubate for  00:20:00 . Add drop-wise to 1 well containing  1.7 mL of media.

Day 3

- 3 Change the media.

Day 4

- 4 Add media containing selection-specific antibiotics.
- 4.1 Use Puromycin (3.5 uG/mL) or Blasticidin (10 uG/mL).

Day 6-7

- 5 Change the media.

Day 8+



- 6 After cells have recovered from antibiotic selection, plate single cells into 96-well dishes to obtain clonal cell lines. After subculturing, confirm cell lines via immunoblotting.