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Splitting 96 Well Plates for gDNA Extraction and Freezing Down

 In 1 collection

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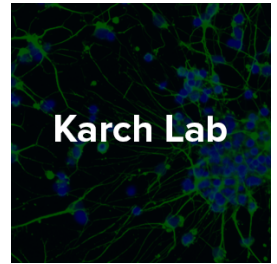
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Protocol status: Working

We use this protocol and it's working

Created: February 16, 2019

Last Modified: February 27, 2019

Protocol Integer ID: 20449



Attachments



Comprehensive

Genomi...

31KB

Guidelines

This protocols is part of the Screening Edited iPSC Clones collection.









Safety warnings

⚠ Please refer to the SDS (Safety Data Sheet) for information about hazards, and to obtain advice on safety precautions.

Before start

Split cells upon reaching maximum 80% confluence and minimum 40%.



- 1 Aspirate media from original plate.
- 2 Wash with  200 μL PBS and aspirate.
- 3 Add  25 μL of 0.05% Trypsin.
- 4 Incubate at  37 °C for  00:05:00
- 5 Tap to lift cells from plate.
- 6 Check under microscope to ensure that cells have detached from plate.
- 7 Add  45 μL mTesR1 to plate and tap to mix.
- 8 Transfer  15 μL to a 96 well PCR plate, while maintaining the location of each sample (this plate will be used for gDNA extraction).
- 9 Add  50 μL of 2X Freezing Media (20% DMSO in FBS) to cell plate and tap to mix.
- 10 Wrap plate in parafilm. Add tape over parafilm. Label the plate on the outside of the tape.
- 11 Place plate in Styrofoam box. Fill any open areas with diapers, paper towels, or Kimwipes. Cover box and make sure that lid is closed completely.
- 12 Store in  -80 °C for up to 4 months .