

Jul 11, 2019

SPARC Serotonin 2B Receptor (5-HT2BR) Immunohistochemistry Protocol in Rat Tissues Labeled with Cholera Toxin B-fragment

 In 1 collection

DOI

dx.doi.org/10.17504/protocols.io.2kigcue

Elisa Gonzalez-Rothi¹, Yasin Seven¹, Latoya Allen¹, Marissa Ciesla¹, Gordon Mitchell¹

¹University of Florida

SPARC

Tech. support email: info@neuinfo.org



Elisa Gonzalez-Rothi

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.2kigcue>

Protocol Citation: Elisa Gonzalez-Rothi, Yasin Seven, Latoya Allen, Marissa Ciesla, Gordon Mitchell 2019. SPARC Serotonin 2B Receptor (5-HT2BR) Immunohistochemistry Protocol in Rat Tissues Labeled with Cholera Toxin B-fragment. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.2kigcue>



License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: May 03, 2019

Last Modified: July 11, 2019

Protocol Integer ID: 22890

Keywords: serotonin 2B receptor, 5HT2BR, 5-HT2BR, immunofluorescence, phrenic, immunohistochemistry, CT-B, cholera toxin B fragment, sparc serotonin 2b receptor, serotonin 2b receptor expression, serotonin 2b receptor expression in ctb, sparc serotonin, serotonin, labelled phrenic motor neuron, phrenic motor neuron, surrounding phrenic motor neuron, receptor expression, receptor, immunohistochemistry protocol in rat tissue, immunohistochemistry protocol, immunofluorescent labeling technique, rat tissue

Abstract

This protocol describes the immunofluorescent labeling technique used to identify serotonin 2B receptor expression in CtB-labelled phrenic motor neurons and within a defined region of interest surrounding phrenic motor neurons.

Troubleshooting



- 1 Day 1: primary antibodies required:
5-HT2BR: Mouse anti-5-HT2BR (SantaCruz Biotechnology #C-6; SC-376878)
Cholera toxin B-fragment: Goat anti-CT-B (Millipore #227040)
- 2 Place 40um transverse spinal cord sections into 1xPBS in 12 well cell culture plates
- 3 5x washes in 1xPBS for 5 minutes each at room temperature
- 4 Blocking: place tissues in 5% Normal Horse Serum (NHS) in 1xPBS-Triton (0.1%) +0.5% bovine serum albumin (BSA) for 60 minutes at room temperature
- 5 Primary Antibody Incubation: Incubate tissues in: 5%NHS in 1xPBS-Triton (0.1%), Mouse anti-5-HT2BR (1:1000), and Goat anti-Ct-B (1:2500) overnight at 4 degrees C
- 6 Day 2: secondary antibodies required:
AlexaFluor 488: donkey anti-goat (Invitrogen, Ref#A11055)
AlexaFluor 594: donkey anti-mouse (Invitrogen, Ref#A21203)
- 7 5x washes in 1xPBS for 5 minutes each at room temperature
- 8 Secondary Antibody Incubation: Incubate tissues in: 5%NHS in 1xPBS-Triton (0.1%), donkey anti-goat (1:1000), and donkey anti-mouse (1:1000) for 2 hours at room temperature
- 9 5x washes in 1xPBS for 5 minutes each at room temperature
- 10 Mount tissues on Superfrost Plus microscope slides
- 11 Allow slides to dry overnight
- 12 Coverslip with VectaShield Antifade Hard Set Mounting Medium (Cat#:H-1400)