

Sep 05, 2019

# Single Cell Dissociation of Small Tumor Biopsies

 Cell

DOI

[dx.doi.org/10.17504/protocols.io.65rhg56](https://dx.doi.org/10.17504/protocols.io.65rhg56)

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DOI: <https://dx.doi.org/10.17504/protocols.io.65rhg56>

External link: <https://doi.org/10.1016/j.cell.2020.07.017>

**Protocol Citation:** Ashley Maynard 2019. Single Cell Dissociation of Small Tumor Biopsies. **protocols.io**  
<https://dx.doi.org/10.17504/protocols.io.65rhg56>



### Manuscript citation:

Maynard A, McCoach CE, Rotow JK, Harris L, Haderk F, Kerr DL, Yu EA, Schenk EL, Tan W, Zee A, Tan M, Gui P, Lea T, Wu W, Urisman A, Jones K, Sit R, Kolli PK, Seeley E, Gesthalter Y, Le DD, Yamauchi KA, Naeger DM, Bandyopadhyay S, Shah K, Cech L, Thomas NJ, Gupta A, Gonzalez M, Do H, Tan L, Bacaltos B, Gomez-Sjoberg R, Gubens M, Jahan T, Kratz JR, Jablons D, Neff N, Doebele RC, Weissman J, Blakely CM, Darmanis S, Bivona TG, Therapy-Induced Evolution of Human Lung Cancer Revealed by Single-Cell RNA Sequencing. Cell 182(5). doi: [10.1016/j.cell.2020.07.017](https://doi.org/10.1016/j.cell.2020.07.017)

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**Protocol status:** In development

**We are still developing and optimizing this protocol**

**Created:** September 05, 2019

**Last Modified:** September 05, 2019

**Protocol Integer ID:** 27537

**Keywords:** dissociation, single-cell, single cell dissociation of small tumor biopsy, single cell dissociation, small core biopsies into single cell, single cell, small tumor biopsy, small core biopsy, cell

## Abstract

Dissociate small core biopsies into single cells.

## Materials

### <sup>1</sup>Base Buffer

DMEM F12

6% FBS

\*good for 1 month at  4 °C C

### <sup>2</sup>Collagenase Buffer

Add 2mg/ml of Collagenase II to Base Buffer.

### <sup>3</sup>FACS Buffer


9.5 mL Miltenyi PBS + EDTA Buffer

0.5 mL Miltenyi BSA

## Troubleshooting









- 1 Make and/or warm Base Buffer<sup>1</sup> to  37 °C .
- 2 Pour the sample(s) into a 10cm dish. Take a picture of the samples in a 10cm dish (in the media it comes in).

Example:



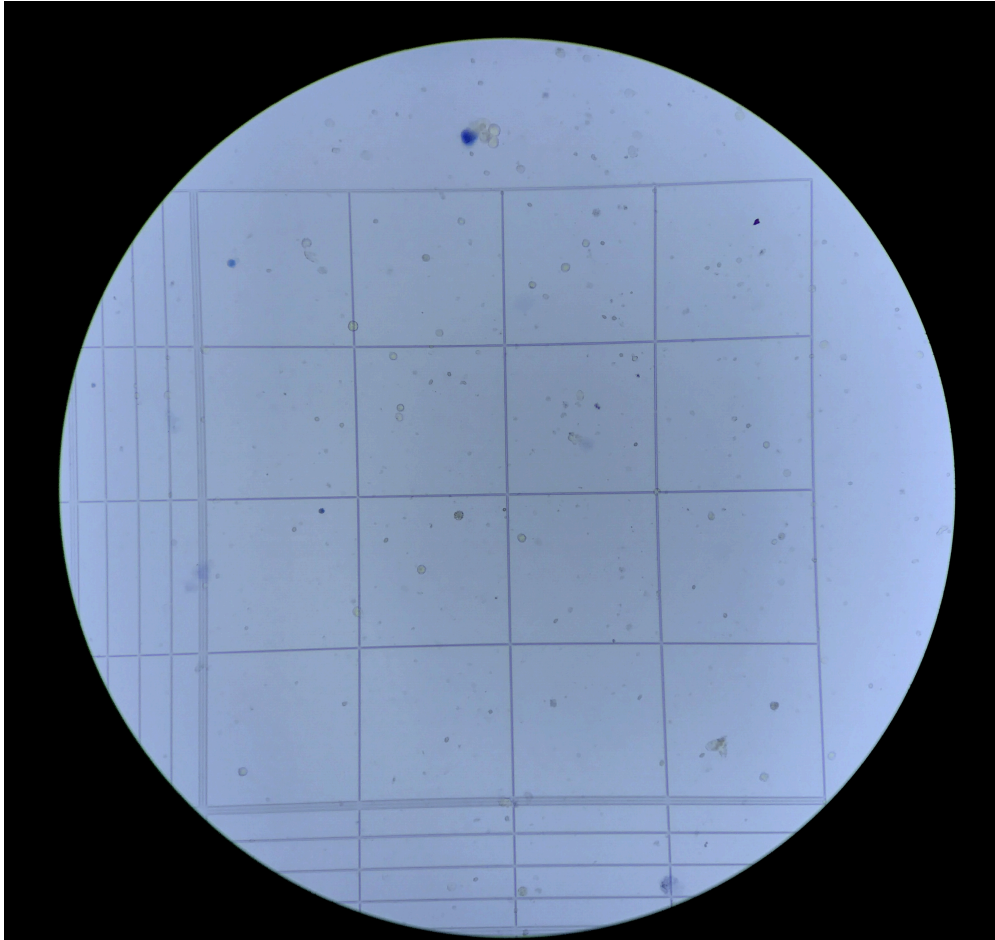
Core biopsy sample in 10cm viewed in 10cm dish.

- 3 Cut sample into small pieces and place into a 1.5mL tube (or multiple tubes if necessary, don't overload the tube with tissue). Add 1.5 mL of collagenase buffer<sup>2</sup> and digest for  00:30:00 ,  37 °C , shaking in a thermomixer @ 800-1000 rpm (the tissue should not be sitting at the bottom of the tube when it's mixing). If chunks of sample remain, pipet the sample up and down 15 times. Return the sample to the thermomixer for  00:30:00 at  37 °C shaking.






- 4 Remove the sample from the thermomixer. Pipet the sample up and down 15 times.
- 5 Filter the sample over a 100-micron filter into a new 1.5 mL tube.
- 6 Wash the sample filter with 1-2mL of collagenase buffer.
- 7 Put the cells that passed through the filter on ice.
- 8 Check cells with Trypan Blue and Hemocytometer (10ul Trypan blue mixed with 10ul of sample), take picture on microscope and label with date and sample ID and record # of live/dead cells.

Example:



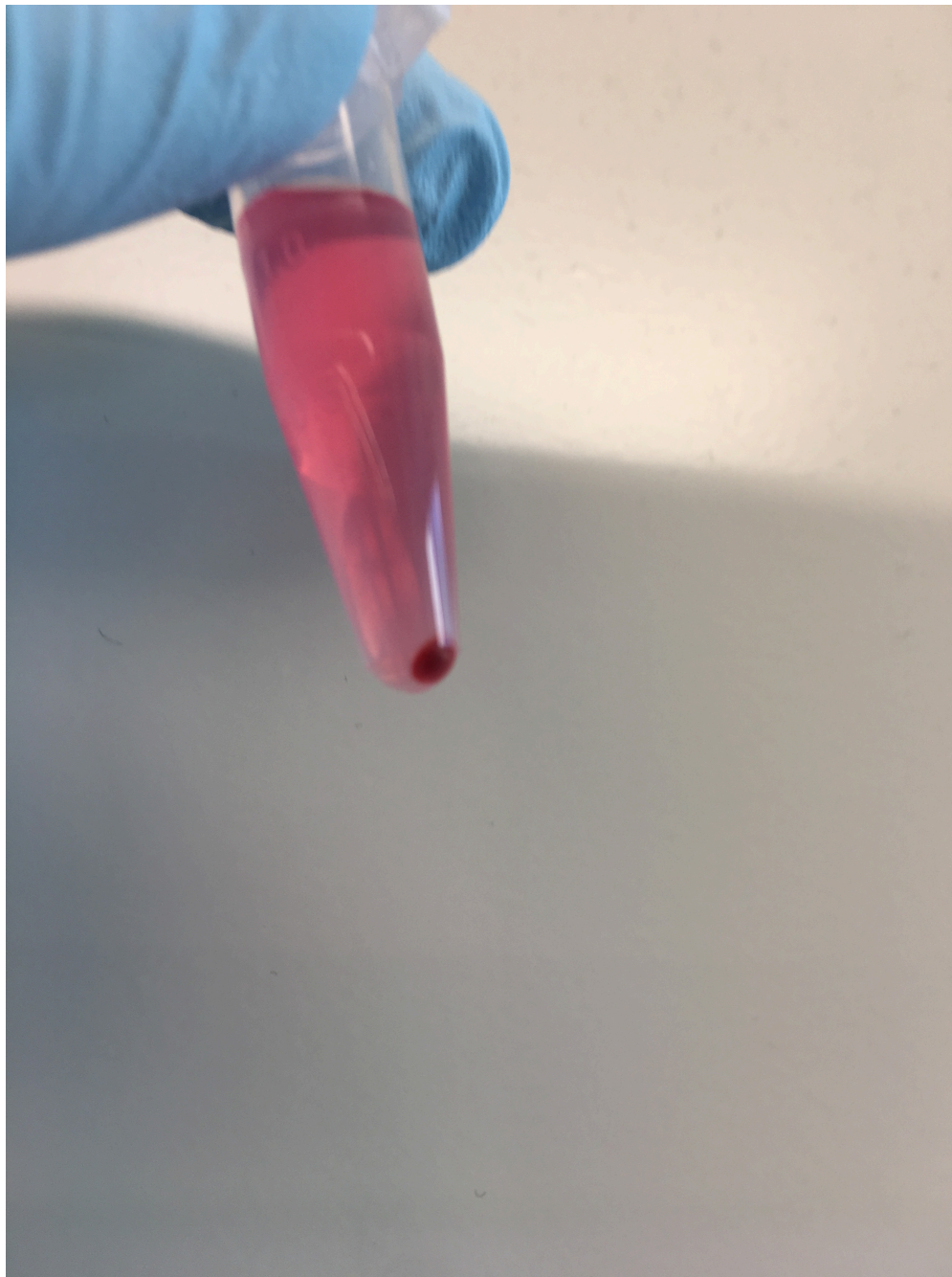
Cells viewed at 10x stained with trypan blue.

- 9    OPTIONAL: If there are persistent chunks of tissue that did not pass through the filter, gather them into a new 1.5 mL tube and add 0.5 mL 0.05% warmed trypsin. Digest the tissue for  00:10:00 at  37 °C , shaking in a thermomixer @ 800-1000 rpm. Quench the reaction with 1 mL Base Buffer. Filter the sample over a 100-micron filter into a new 1.5 mL tube. Wash filter with 1-2 mL of collagenase buffer. Check cells with Trypan Blue and Hemocytometer, take picture on microscope and label with date and sample ID and record # of live/dead cells. Combine the filtered cells from step 7 and 9 into one tube.
- 10    Spin at 1500 rpm for  00:10:00 .



- 11 OPTIONAL (Only if pellet is visibly red): Add 0.5 mLs RBC lysis buffer to sample tubes. Leave at RT for 2-5min. Add 1.0 mL DMEM F12 + 6% FBS to each tube and spin again.




Example:



Example of red pellet of cells that needs RBC lysis treatment.

- 12 Aspirate off supernatant. Resuspend in 120  $\mu$ L of FACS Buffer<sup>3</sup>.



- 13 Count cells with Trypan Blue and hemocytometer (10ul Trypan blue mixed with 10ul of sample). Record the number of live and dead cells (stained blue). Take picture on microscope and label with date + sample ID + "final"
- 14 Remove 10ul of sample into a new 1.5mL tube, add 350uL of FACS Buffer and set aside for unstained control.
- 15 Stain remaining cells with 10ul CD45-FITC (if cell count is under  $1 \times 10^7$ ) and 1ul of Hoechst stain (diluted 1:10). Incubate on ice in dark for  00:20:00 .
- 16 Add 1mL of FACS Buffer to stained cells. Spin at 1500 rpm for  00:10:00 and aspirate off supernatant. Resuspend with 0.5 mLs of FACS Buffer.
- 17  00:05:00 before sorting add, 0.5 uL of PI and mix well. Transfer unstained and stained samples to FACS tubes and leave on ice in the dark.