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Single cell digestion of tumor tissue

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We use this protocol and it's working

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Abstract

The purpose of this protocol is to provide details on how to obtain a single-cell suspension from a patient-derived xenograft tumor.

Materials

	Materials	Source	Catalog Number
	DMEM	Gibco	11965092
	Fetal bovine serum (FBS)	Global Life Sciences Solutions	SH3039603
	Type I collagenase	Gibco	17100017
	DNase I	Thermo Scientific	89836
	ROCK Inhibitor (Y-27632)	Sigma Aldrich	SCM075
	Gentamicin	Gibco	15750060
	HEPES-buffered saline (HBS)	Gibco	15630080
	Red Cell Lysis Buffer	Roche	11814389001
	Cell Freezing Medium-DMSO 1×	Sigma Aldrich	C6164
	Scissors	Kent Scientific	INS600393
	Scalpels with blade no.10	Fisher Scientific	12-460-451
	Petri dishes	Millipore Sigma	P5481
	15 mL conical tubes	Corning	430055
	50 mL conical tubes	Corning	430829
	70 µm cell strainers	Corning	431751
	40 µm cell strainers	Corning	431750

	Materials	Source	Catalog Number
	Pipettes tips (1000 ul)	Millipore Sigma	AXYT1000B
	Trypan Blue dye	Sigma Aldrich	93595
	Corning CoolCell	Corning	432000
	Cryogenic vial	Nalgene	V5007

Preparation of Medium

1 Prepare digestion medium
DMEM supplemented with
a. [M] 10 % volume FBS,
b. [M] 200 units/ml of type I collagenase,
c. [M] 1 units/ml DNase I,
d. [M] 2 micromolar (µM) Y-27632 and
e. [M] 100 ug/ml Gentamicin

2 Prepare storage medium
DMEM supplemented with
a. [M] 10 % volume FBS, and
b. [M] 100 ug/ml Gentamicin

Harvest Tumor Tissue

3 Prepare an ice bucket with ice and 50 ml falcon tubes with 35-45 mL HBS.

4 Euthanize the tumor-bearing mouse and extract the tumor.

5 Blot the tumor on a Kim-wipe to get rid of excess blood.

6 Weigh the tumor and note the weight.

7 Place the fresh tumor tissue in a falcon tube containing 35-45 mL HBS.

8 Transfer the falcon tube into a BSL2 hood.

Tissue digestion process (performed in BSL2 hood) 6h 3m

9 Prepare a petri dish containing fresh HBS and place it on ice.

10 Transfer the tissue from the falcon tube into the petri dish containing HBS.

11 Wash tumor multiple times with 10 mL HBS and remove excess HBS. Repeat this process two more times.

12 Mince the tumor with scissors or scalpels in the petri dish.

Note

For larger tissue (larger than 1 cm^3), use multiple rounds of mincing

13 Add digestion medium to the petri dish containing tumor tissue and add enough volume to thoroughly cover the entire tumor.

14 Place the petri dish in an incubator at 37 °C 95% air/5% CO2 over 02:00:00 - 04:00:00 hrs. 6h

Note

Vigorously mix the tissue every 10-15 minutes using a pipette to aid dissociation.

15 Periodically assess the digestion under the microscope and stop once small clusters or clumps of cells are released from the tissue and before complete digestion to single cells occurs.

16 Transfer the above solution into a 50 ml falcon tube and centrifuge the above solution at 300 x g, 4°C for 5 mins.

17 Discard the supernatant and incubate the digested tissue with 5 mL Red Cell Lysis Buffer for 00:03:00 mins. 3m

18 Centrifuge the above solution at 300 x g, 4°C for 5 mins.

19 Discard the supernatant and resuspend the cell pellet in 50 mL fresh DMEM prepared in step 2.

20 Pass the above solution containing tumor cells through a 70 um filter and collect the solution containing tumor cells that passed through the filter in a fresh 50 ml falcon tube.

21 Pass the above solution containing tumor cells through a 40 um filter and collect the cells that passed through the filter in a fresh 50 ml falcon tube.

22 Centrifuge the solution containing tumor cells at 300 x g, 4°C for 5 mins.

23 Discard the supernatant and resuspend the cell pellet in fresh DMEM prepared in step 2.

24 Perform Trypan Blue dye exclusion test and note the live and total cell count.

25 Prepare cryogenic vials with 900 µL of 1 M live cells and 100 µL of DMSO per cryovial.

26 Place the prepared cryovials in a freezing container and place the container in -80 °C freezer for 24 hours.

27 After 24 hours, transfer the cryovials into liquid nitrogen storage for long-term storage.