

May 25, 2020

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) real-time RT-PCR ORF1ab 2020

Forked from Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) real-time RT-PCR ORF1ab 2020 (Wuhan-ORF1ab; 2019-nCoV-related test)

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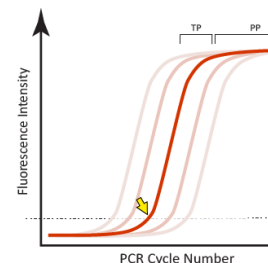
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Protocol status: Working

We use this protocol and it's working

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Abstract

- A real-time RT-PCR to specifically detect SARS-CoV-2 betacoronavirus also called nCoV-2019 or Wuhan seafood market pneumonia virus. Based on sequence [MN908947](#) made available by Professor Yong-Zhen Zhang, Fudan University, Shanghai, China.
- The target region is within the ORF1ab sequence.

Notes

1. Assay is fully optimised (as of 24Jan2020).
2. This test has identified a clinical positive cases of coronavirus disease (COVID-19)

Guidelines

- If using a different brand or model of real-time thermocycler, check the concentration of ROX is adequate.
- Method assumes the user is familiar with the thermocycler and software used to run the protocol.

Materials

STEP MATERIALS

 SensiFAST™ Probe Lo-ROX One-Step Kit **Bioline Catalog #BIO-78001**

Protocol materials

 SensiFAST™ Probe Lo-ROX One-Step Kit **Bioline Catalog #BIO-78001**



Mix

1 Oligonucleotides

	Oligo Name	Sequence 5'-3'	Location based on NC_045512*
	WuhanORF1ab-F	AATCCACCTGCTCTACAAGATG	545 5- 547 6
	WuhanORF1ab-R	CATCACCTAACTCACCTACTGTC	556 6- 554 4
	WuhanORF1ab-P	6FAM- AGCTTCACCAGCCCTTGCTCT-BHQ1	550 5- 548 5

*GenBank accession NC_045512 Wuhan seafood market pneumonia virus isolate Wuhan-Hu-1

2 Reagents

 SensiFAST™ Probe Lo-ROX One-Step Kit **Bioline Catalog #BIO-78001**

3 Synthetic controls

Synthetic controls are produced using the binary synthetic template oligonucleotide positive control for in-house diagnostic real-time RT-PCR method.

The oligonucleotide sequences required to make controls for this assay are:

Probe control:

AAAATAATACGACTCACTATAGGGTGAAGAGAATCCACAAGGAATTGAAAGCTTCACCAGCC
CTTGCTCTACAGTGTTTCAGCAGGTCCTGTTGAAAA



Primer control:

AAAATAATACGACTCACTATAGGGAATCCACCTGCTCTACAAGATGATGATCTGGCACGGGA
CCCTCCAAGACAGTAGGTGAGTTAGGTGATGAAAA

4 Reaction Set-up

- Assay has been designed to be used on both a Rotor-Gene 6000 / Rotor-Gene Q 5-plex using 100-place rotor discs and a ABI 7500 Fast real-time machine.
- Total reaction volume is 20 μ L.
- Prepare sufficient for number of reaction plus a 'dead volume' usually 2 extra. Adjust as necessary if using a robotic dispenser.

Reagent	Volume (ul) X1	Final reaction concentration
Nuclease free water	4.21	
WuhanORF1ab-F (200uM)	0.07	700nM
WuhanORF1ab-R (200uM)	0.09	900nM
WuhanORF1ab-P (100uM)	0.03	150nM
2 X SensiFast Probe Lo-Rox One-Step mix*	10	1X
RiboSafe RNase Inhibitor*	0.4	
Reverse transcriptase*	0.2	
TOTAL VOLUME	15	

*SensiFAST™ Probe Lo-ROX One-Step Kit (BIO-78005)

Dispense 15 μ L to each reaction well.

Add 5 μ L of template, extracted RNA, controls or NTC (nuclease-free water).

Total reaction volume is 20 μ L.

Amplification

5 PCR amplification

1 cycle	50 cycles
50°C 5min	95°C 3 seconds
95°C 2min	60°C 30 seconds*

*Fluorescence acquisition step

Result Analysis

- 6 The definition used for a satisfactory positive result from a real-time fluorogenic PCR should include each of the following:
1. A sigmoidal curve – the trace travels horizontally, curves upward, continues in an exponential rise and followed by a curve towards a horizontal plateau phase
 2. A suitable level of fluorescence intensity as measured in comparison to a positive control (y-axis)
 3. A defined threshold (C_T) value which the fluorescent curve has clearly exceeded (Fig.1 arrow) and which sits early in the log-linear phase and is <40 cycles
 4. A flat or non-sigmoidal curve or a curve that crosses the threshold with a C_T value >40 cycles is considered a negative result
 5. NTCs should not produce a curve

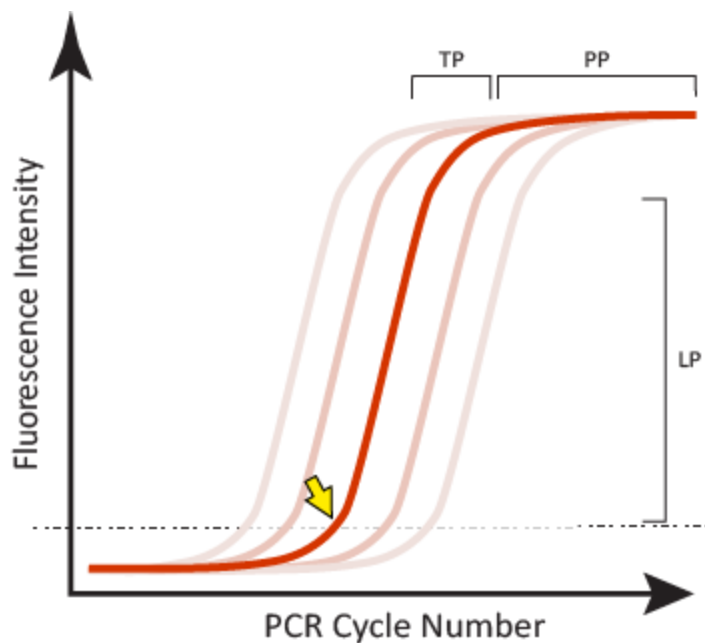


Figure 1. Examples of satisfactory sigmoidal amplification curve shape when considering an assay's fluorescent signal output. The crossing point or threshold cycle (C_T) is indicated (yellow arrow); it is the value at which fluorescence levels surpass a predefined (usually set during validation, or arbitrary) threshold level as shown in this normalized linear scale depiction. LP-log-linear phase of signal generated during the exponential part of the PCR amplification; TP-a slowing of the amplification and accompanying fluorescence signal marks the transition phase; PP-the plateau phase is reached when there is little or no increase in fluorescent signal despite continued cycling.