

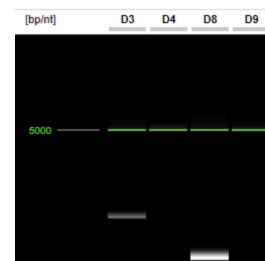
Mar 07, 2020

Version 2

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) RdRp nested RT-PCR V.2

DOI

<https://dx.doi.org/10.17504/protocols.io.bdcpi2vn>



Judy Northill¹, Ian Mackay¹

¹Public Health Virology, Forensic and Scientific Services

Public Health Virology, F...

Coronavirus Method De...



Judy A Northill

Public Health Virology, Forensic and Scientific Services

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.bdcpi2vn>

Protocol Citation: Judy Northill, Ian Mackay 2020. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) RdRp nested RT-PCR. protocols.io <https://dx.doi.org/10.17504/protocols.io.bdcpi2vn>

Manuscript citation:

Hu D, Zhu C, Wang Y, Ai L, Yang L, Ye F, et al. Virome analysis for identification of novel mammalian viruses in bats from Southeast China. Scientific Reports. 2017;7(1):10917 . <https://www.nature.com/articles/s41598-017-11384-w>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: March 06, 2020

Last Modified: March 07, 2020

Protocol Integer ID: 33903

Keywords: COVID-19, SARS, coronavirus, Rd-Rp, RT-PCR, PCR, SARS-CoV-2, severe, severe acute respiratory syndrome coronavirus, coronavirus rt, specific to sar, sar, pcr, nested rt, rdrp region, rdrp, sequencing,

Abstract

A nested RT-PCR targeting the RdRp region of the sub-genus *Sarbecovirus*. The primers are modified from the pan-coronavirus RT-PCR published by Hu *et al.* 2017 to be more specific to SARS-CoV-2.

Assay may be used in resource poor settings where real-time cyclers are not available.

Sanger sequencing can be used to confirm SARS-CoV-2 where WGS is not available or where WGS fails due to poor quality sample.

Materials

STEP MATERIALS

 SuperScript III One-Step RT-PCR System with Platinum Taq **Invitrogen - Thermo Fisher Catalog #12574026**

 MyFi Mix **Bioline Catalog #BIO-25049**

Protocol materials

 SuperScript III One-Step RT-PCR System with Platinum Taq **Invitrogen - Thermo Fisher Catalog #12574026**

 MyFi Mix **Bioline Catalog #BIO-25049**

Oligonucleotides

1

	Name	Purpose	5'-3'
	Wuhan15283-F	Round 1 forward primer	ATG GGT TGG GAT TAT CCT AAA TG
	Wuhan15887-R	Round 1 reverse primer	TGT TGA GAG CAA AAT TCA TG
	Wuhan15286-F	Round 2 forward primer	GGT TGG GAT TAT CCT AAA TGT GA
	Wuhan15725-R	Round 2 reverse primer	GCA TCG TCA GAG AGT ATC ATC AT

Round 1 amplified products will produce a band of 605bp.

Round 2 amplified products will produce a band of 440bp.

Mix



2



SuperScript III One-Step RT-PCR System with Platinum Taq Invitrogen - Thermo
Fisher Catalog #12574026



MyFi Mix Bioline Catalog #BIO-25049

Round 1 mix

	Rea gent	Volu me x1 (μl)	Final Con cent ratio n
	Nucl ease -free water	3.2	
	2 x Reac tion mix*	10	1 X
	Wuh an15 283- F (20p mol/ μ l)	0.5	500 nM
	Wuh an15 887- R (20p mol/ μ l)	0.5	500 nM
	Sup ersc ript/ Plati num Taq mix*	0.8	
	TOT AL	15	

*SuperScript III One-Step RT-PCR System with Platinum Taq #12574026

Dispense mix in 15 μ l amounts in 0.2ml PCR tubes suitable for thermocycler.

Mix maybe stored frozen if necessary.

Round 2 mix

	Rea gent	Volu me x1 (μl)	Final Con cent ratio n
	Nucl ease -free wate r	4	
	MyFi 2X mix*	10	1X
	Wuh an15 286- F (20p mol/ μ l)	0.5	500 nM
	Wuh an15 725- R (20p mol/ μ l)	0.5	500 nM
	TOT AL	15	

*MyFi™ Mix Bioline #BIO-25049

Dispense mix in 15 μ l amounts in 0.2ml PCR tubes suitable for thermocycler.
Mix maybe stored frozen if necessary.

AMPLIFICATION ROUND 1

3

- Thaw required number of tubes with round 1 mix.
- Add 5 μ l of extract, control, or NTC (nuclease-free water) to round 1 mix above for a final reaction volume of 20 μ l.
- Label the tubes and record details.

The assay has been used with Eppendorf thermocyclers.

**PCR cycling times**

	1 cycl e	40 cycl es	1 cycl e	Hold
	50° C 30 minu tes	94° C 30 seco nds	68° C 5 minu te	15°C
	94° C 2 minu tes	55° C 30 seco nds		
		68° C 1 minu te		

AMPLIFICATION ROUND 2

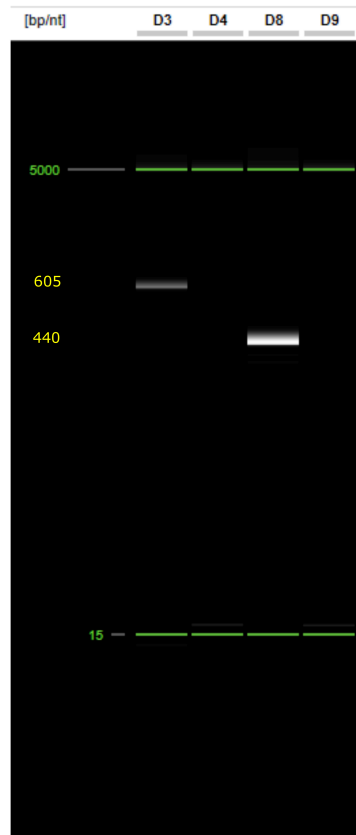
- 4
- Dilute each of the round 1 amplicons 1/100 in nuclease-free water (2ul + 198ul is ideal)
 - Thaw required number of tubes with round 2 mix.
 - Add 5µl of the diluted round 1 amplicon.
 - Total reaction volume is 20µl.

PCR cycling times

	1 cycl e	40 cycl es	1 cycl e	Hold
	95° C 3 minu tes	95° C 15 seco nds	72°C 1 minu te	15°C
		55° C 15 seco nds		
		72°C 15 seco nds		

GEL ELECTROPHORESIS

- 5 Amplified products are analysed by gel electrophoresis or equipment such as a QIAxcel or equivalent.
Round 1 amplified products will produce a band of 605bp.
Round 2 amplified products will produce a band of 440bp.
No template controls (NTC) should be not detected.



Example of result from a QIAxcel.

D3-Round 1 band, D8-round 2 band, D4 and D9 are NTC in round 1 and 2 respectively.