

Mar 08, 2024

Version 1

Sequencing-based Neutralization Assay for Influenza A Virus V.1

DOI

<https://dx.doi.org/10.17504/protocols.io.kqdg3xdmpg25/v1>

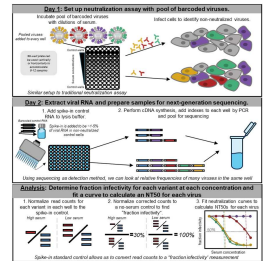
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Protocol Citation: Andrea Loes, Rosario Araceli Tarabi, Jesse D. Bloom 2024. Sequencing-based Neutralization Assay for Influenza A Virus. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.kqdg3xdmpg25/v1>

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Protocol status: Working

We use this protocol and it's working

Created: January 20, 2024

Last Modified: May 08, 2025

Protocol Integer ID: 93844

Keywords: influenza, virus, neutralization, titer, influenza virus test, genomic segment of the influenza virus, neutralization of all of the virus, neutralization assay, influenza virus, approach for neutralization assay, based neutralization assay, assays with serum sample, sequencing, single viral strain, single serum sample, serum sample, single serum sample in each measurement, nucleotide barcode into the hemagglutinin, assay, traditional neutralization measurement, titers of serum sample, serum, nucleotide barcode, many virus, barcoded virus, traditional neutralization, hemagglutinin, same serum volume, genomic segment

Funders Acknowledgements:

National Institutes of Health

Grant ID: R01AI165821

Howard Hughes Medical Institute

Grant ID: Investigator

National Institutes of Health

Grant ID: 75N93021C00015

Abstract

Traditional neutralization assays for influenza virus test a single viral strain against a single serum sample in each measurement. Here we describe a sequencing-based approach for neutralization assays that can measure the titers of serum samples against many viruses at once using the same serum volume and workflow of a traditional neutralization measurement. This method relies on incorporating a nucleotide barcode into the hemagglutinin genomic segment of the influenza virus, pooling many barcoded viruses together, and then using Illumina sequencing to read out the neutralization of all of the viruses in the pool at once. Here we provide the step-by-step protocol for running these assays with serum samples.



Materials

(BEFORE DAY 1):

- ☒ Phosphate Buffered Saline **Thermo Fisher Scientific Catalog #28374** (1X)
- ☒ Receptor destroying enzyme (RDE) **Denka Seiken Co., Ltd**
- Barcoded influenza virus library containing HAs from strains of interest
- RNA spike-in control
- Serum samples

(DAY 1):

- D10 Media (recipe below)
- Influenza Growth Media (recipe below)
- Tissue-culture-treated 96-well plates
- Barcoded influenza virus library
- RDE-treated sera from before day 1
- Confluent 100 mm plate of MDCK-SIAT1 cells
- Trypsin-EDTA solution for dissociating cells from plate

D10 Media Recipe:

🧴 450 mL DMEM (High glucose)

🧴 50 mL Heat-inactivated Fetal Bovine Serum (FBS)

🧴 5 mL Penicillin Streptomycin Solution (10000 U/mL)

🧴 5 mL L-Glutamine (200 mM)

→sterile filter into a 500mL bottle

→store 🌡 4 °C

Influenza Growth Media Recipe:

🧴 20 mL BSA Fraction V Solution, 7.5%

🧴 5 mL Penicillin Streptomycin Solution (10000 U/mL)

🧴 500 µL Filtered Calcium Chloride (100 mg/mL)

🧴 50 µL Heat-Inactivated Fetal Bovine Serum (FBS)

🧴 475 mL Opti-MEM I

→ sterile filter into a 500mL bottle

→ store 🌡 4 °C

(DAY 2)

- ☒ iScript™ cDNA synthesis kit **Bio-Rad Laboratories Catalog #170-8841**

-  iScript™ RT-qPCR Sample Preparation Reagent **Bio-Rad Laboratories Catalog #1708898**
- RNA spike-in control
- Gene specific cDNA primer (5' CTCCCTACAATGTCGGATTTGTATTTAATAG-3')
- Phosphate buffered Saline
-  KOD Hot Start DNA Polymerase **Merck MilliporeSigma (Sigma-Aldrich) Catalog #71086-3**
- Forward Primer (5'- GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTCTCCCTACAATGTCGGATTTGTATTTAATAG-3')
- Reverse Primer (5'-AGTAGAAACAAGGGTGTTCCTTATATTTCTGAAATCC-3')
- Indexing Primer mixes (5 uM stocks of mixed Fwd and Rev Primers)
- 80% Ethanol (EtOH)
-  Agencourt AmPure XP beads **Catalog #A63880**
- Elution buffer

Safety warnings

-  Current and recent strains of human influenza virus are biosafety-level 2, but some historical human strains and animal strains are biosafety level 3 or higher. If you aren't sure of the biosafety level of the strains you are working with, check with your biosafety committee.

Several steps within the protocol are performed in a biosafety cabinet as they pose aerosol hazards. Prior to heat inactivation of the sera, all serum samples should be processed within a biosafety cabinet. In addition, as this protocol uses influenza A virus, setup for neutralization assays and preparation of cDNA reactions should be carried out in a biosafety cabinet. To reduce the hazard of working with aerosols, prepare a beaker of 1:10 dilution of bleach before work within the biosafety cabinet, immediately after pipetting any potential aerosol hazards, rinse pipette tips in the bleach solution by pipetting up and down. Seclude bleached tips to be disposed with other virus-contaminated materials as biohazardous waste.



Barcoded influenza HA library

1

Note

For this protocol, a pooled library of influenza virus strains which each contain a unique barcode sequence within the HA segment is needed. The reagents and protocols used to generate a barcoded influenza library are described in the associated manuscript. Prior to the start of this protocol, a barcoded influenza library must be obtained as well as a custom RNA spike-in control, which is used in the normalization of barcode count data in order to calculate neutralization titers from sequencing data.

(BEFORE DAY 1) RDE-treat & heat-inactivate sera

3h 40m

2

Note

It is important to treat serum with receptor destroying enzyme (RDE) prior to use in influenza neutralization assays. This process removes residual sialic acids from serum components that can otherwise inhibit HA-mediated infection. This protocol was modified slightly from Zost et al, 2017 (<https://doi.org/10.1073/pnas.1712377114>).

Keep in mind that the processes in these steps effectively dilute the final RDE-treated sera 1:4, and that dilution factor should be taken into account when calculating neutralization titers.

3 Thaw sera  On ice

30m

4 Prepare RDE solution, by adding  20 mL 1X phosphate buffered saline (PBS) into one receptor destroying enzyme (RDE) bottle (VWR, Cat. No. 370013). Replace stopper and gently invert to mix. Sterile filter the solution through a 0.22 um filter prior to use. Aliquots of this RDE solution can be stored at -20 C for future use, if needed.

10m

5 Pipette RDE solution into labeled, externally threaded tubes, such that you can dilute the sera 1:4. For our sera samples, we added  300 µL RDE solution . Then, added  100 µL serum .

10m



- 6 To remove residual sialic acids in serum incubate serum and RDE at 37 °C 02:30:00 2h 30m
- 7 Then, to heat-inactivate the RDE and serum, incubate at 56 °C 00:30:00 30m
- 8 Move samples to ice, and then store heat-inactivated and RDE-treated serum samples at -80 °C until you are ready to begin the neutralization assays.

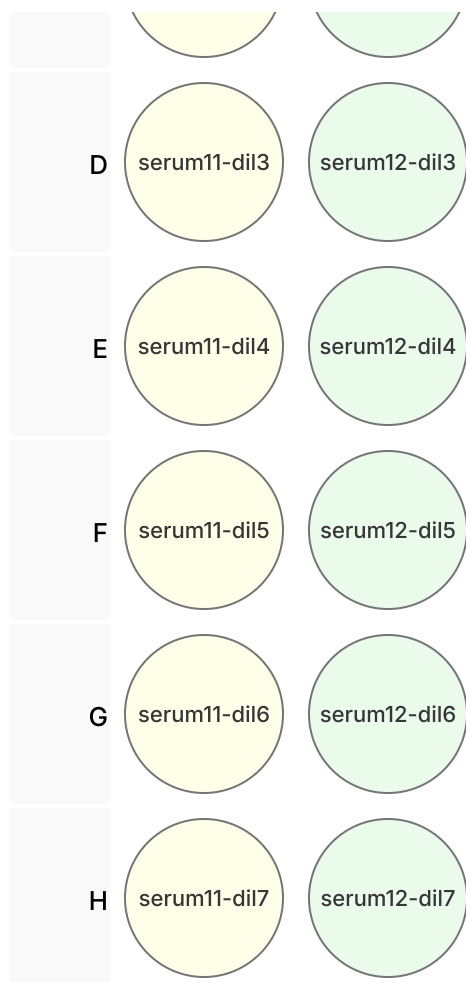
(DAY 1) Determine your plate setup

- 9 Different plate setups may be used depending on how many samples of sera you are testing, and how many dilutions you wish to run for each serum. For example, you could perform serial dilutions of serum down the plate vertically, running up to 12 serum samples with 7 different dilutions (plus a no-serum control) per serum sample. Or you could perform dilutions across the plate horizontally, running 8 serum samples with 11 different dilutions per serum-sample (plus a no-serum control). Additional plate set up designs are also feasible to meet the needs of your particular project.

Pictured below is an example plate setup for 12 serum samples with 7 concentrations of serum

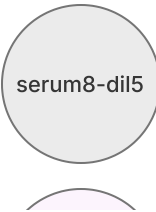
	1	2	3	4	5
A	no serum	no serum	no serum	no serum	no serum
B	serum1-dil1	serum2-dil1	serum3-dil1	serum4-dil1	serum5-dil1
C	serum1-dil2	serum2-dil2	serum3-dil2	serum4-dil2	serum5-dil2
D	serum1-dil3	serum2-dil3	serum3-dil3	serum4-dil3	serum5-dil3
E	serum1-dil4	serum2-dil4	serum3-dil4	serum4-dil4	serum5-dil4
F	serum1-dil5	serum2-dil5	serum3-dil5	serum4-dil5	serum5-dil5
G	serum1-dil6	serum2-dil6	serum3-dil6	serum4-dil6	serum5-dil6
H	serum1-dil7	serum2-dil7	serum3-dil7	serum4-dil7	serum5-dil7
	6	7	8	9	10
A	no serum	no serum	no serum	no serum	no serum





Pictured below is another example plate setup (8 serum samples with 11 concentrations of serum).



	1	2	3	4	5
A	No serum	serum8-dil1	serum8-dil2	serum8-dil3	serum8-dil4
B	No serum	serum7-dil1	serum7-dil2	serum7-dil3	serum7-dil4
C	No serum	serum6-dil1	serum6-dil2	serum6-dil3	serum6-dil4
D	No serum	serum5-dil1	serum5-dil2	serum5-dil3	serum5-dil4
E	No serum	serum4-dil1	serum4-dil2	serum4-dil3	serum4-dil4
F	No serum	serum3-dil1	serum3-dil2	serum3-dil3	serum3-dil4
G	No serum	serum2-dil1	serum2-dil2	serum2-dil3	serum2-dil4
H	No serum	serum1-dil1	serum1-dil2	serum1-dil3	serum1-dil4
	6	7	8	9	10
A	serum8-dil5	serum8-dil6	serum8-dil7	serum8-dil8	serum8-dil9
					



**(DAY 1) Set up incubations of serum and virus**

1h 20m

10

Safety information

The following steps should be carried out in a biosafety cabinet because they involve influenza virus. Current and recent strains of human influenza virus are biosafety level 2, but some historical human strains and animal strains are biosafety level 3 or higher. If you aren't sure of the biosafety level of the strains you are working with, check with your biosafety committee.

Prepare a beaker of 10% bleach immediately before performing any of the steps for rinsing pipette tips. Be sure to rinse tips here after interaction with virus samples, and seclude tips to be disposed with the rest of your lab's virus waste.

11

Thaw virus and RDE-treated sera Room temperature 00:30:00

30m

12

Pick an initial serum dilution factor for your assay: this will be the highest serum concentration tested, and other wells will contain serial dilutions of this concentration. You also need to choose what dilution factor to use as you dilute the serum from one well to the next. For example, you might start with an initial dilution of 1:20 and perform 3-fold dilutions down the plate. Recall that the RDE treated sera has already been diluted 1:4 in the steps above.

Note

The best choice of the initial dilution and dilution-factor used will depend on the serum that is being tested with this method, as well as whether you want to always have the dilution range capture the neutralization titer 50% (NT50), or if just determining a bound on the NT50 is sufficient. If a larger range in NT50s is expected, and downstream analysis which requires that all NT50s measured are within range (such as assessing fold-change), a larger dilution factor might be preferred. If you are trying to measure small changes in neutralization titers within a given range, then a smaller dilution factor may be preferred, even though this may result in some NT50s being out of range of dilutions tested.

13

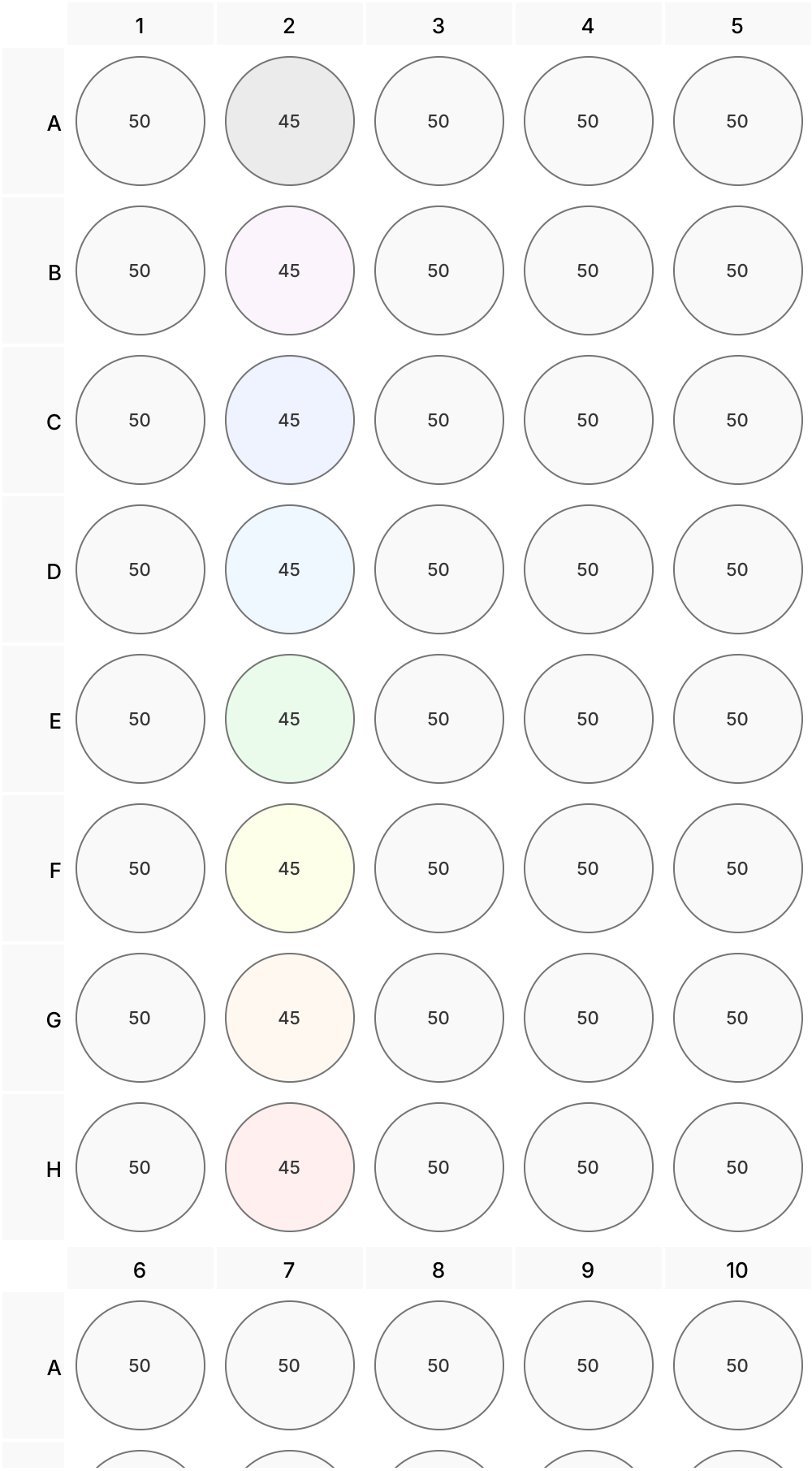
Add influenza growth media (Opti-MEM supplemented with 0.1% heat-inactivated FBS, 0.3% bovine serum albumin, 100 µg per mL of calcium chloride, 100 U per mL penicillin, and 100 µg per mL streptomycin) to your plate such that all rows contain either 50 uL or the volume needed for your initial dilution of sera.

5m



An example is shown below, using the setup described above where 8 serum samples are run horizontally across the plate. In this example, we use an initial dilution of serum of 1:20. As the serum has been diluted 1:4 during RDE treatment and will be diluted 1:2 with virus in a later step, you'd need to dilute the RDE-treated sera 2:5 for the initial dilution in the plate to obtain a final dilution of 1:20.

For this example, you would add  45 µL influenza growth media to the wells in your initial dilution column and  50 µL influenza growth media in every other well:



B	50	50	50	50	50
C	50	50	50	50	50
D	50	50	50	50	50
E	50	50	50	50	50
F	50	50	50	50	50
G	50	50	50	50	50
H	50	50	50	50	50
	11	12			
A	50	50			
B	50	50			
C	50	50			



- 14 Add 30 μL RDE-treated serum to each well in column 2. This leaves you with a 1:10 sera dilution of the original serum (a 2:5 dilution of RDE-treated serum that was already diluted 1:4 during RDE treatment) in 75uL total volume in the wells in column 2.

5m

- 15 Perform serial 3-fold dilutions, pipetting 25 μL serum dilution from column 2 into column 3, and mixing by pipetting up and down several times. Repeat this step with column 3 into column 4 and so forth until you get to row 12, after which you will pipette the residual 25 uL from this column directly into bleach. After this step, you will have completed the serial 3-fold dilutions across the plate. You should still have no serum in column 1, and all 96 wells should now contain 50uL total volume.

5m

- 16 The appropriate amount of virus library needed for each plate is experimentally determined as the amount of virus that can be added to the number of cells per well (50,000 MDCK-SIAT1 cells as described below) and still be in the linear range of viral transcriptional output changing with viral dilution. (See note for more detail). As 50 μL of virus will be added to each well, we first dilute the virus library that the appropriate amount of virus is in each 50 μL aliquot, and then add 50 μL virus dilution to each well (including the control, no serum row of wells).

5m

**Note**

This method requires that the number of sequencing counts for each HA viral barcode normalized by the spike-in RNA counts be directly proportional to the number of virions encoding that barcode that infect cells. The amount of virus library used per well for this assay should be experimentally determined to meet this condition. The range at which this is true depends on the number of transcriptionally active viral particles per cell (MOI). At low to moderate MOI, increasing the number of virions infecting cells will result in more barcoded viral RNA produced, but at high MOIs the capacity of infected cells to produce viral RNA becomes saturated. To identify the amount of virus library where there is a linear relationship between the amount of input library and the resulting spike-in normalized HA counts, one should perform a dilution series, infecting wells with different amounts of virus library, adding a known concentration of RNA spike-in control and sequencing these samples to identify the highest amount of virus library that can be added and still result in a two-fold decrease in the counts of viral barcodes relative to spike-in control barcodes for every 2-fold dilution of virus library.

A description of how to prepare the RNA spike in control (a single stranded RNA molecule, which resembles barcoded HA vRNA, but with the sequence of GFP in place of the HA ectodomain) is provided in the associated manuscript.

- 17 Incubate plate of virus and serum 🔥 37 °C in CO2 incubator ⌚ 01:00:00

1h

(DAY 1) Add cells to plate

30m

- 18 After the virus-serum mix has been incubated for 35 min of the 60 min incubation time, prepare cells to be added to the plate. First, aspirate media off of a confluent 100 mm plate of MDCK-SIAT1 cells. Wash the cells once with 🧴 2 mL PBS . Then, treat cells with 🧴 2 mL trypsin-EDTA , incubating at 🔥 37 °C in CO2 incubator until cells disassociate from the plate (this typically takes around 5-10 min, since MDCK-SIAT1 cells are highly adherent). Once the cells are mostly dissociated, pipetting gently can help dissociate the remainder.

12m

19

Note

We use MDCK-SIAT1 cells in this assay rather than MDCK-SIAT1-TMPRSS2 cells to limit secondary viral replication (TMPRSS2 cleaves HA in producing cells to activate it for infection). These cells adhere extremely well to tissue-culture plates, so make sure to allow for sufficient time to trypsin treat and wash such that these cells are ready after virus and serum have incubated for 1 hour.



- 20 Inactivate the trypsin by adding 4 mL D10 media , washing the cells off the plate and resuspending in this media. Transfer the resuspended cells to a conical tube. Centrifuge 300 x g, 00:03:00 , then aspirate the supernatant. Wash cells once with 5 mL influenza growth media , and then resuspend cells in 5 mL influenza growth media . Use a cell counter to determine cell concentration, and dilute cells to 1e6 cells/mL with more influenza growth media. You will need about 5.5 mL of cells at 1e6 cells/ml in influenza growth media for each 96-well plate that you wish to run. 8m
- 21 Once the plate with the serum and the virus library has been incubating for 1 hour total, add 50 μ L 1e6 cells/mL in influenza growth media to each well. 10m
- 22 Return the plate containing cells, serum, and virus library to the 5% CO₂ incubator. Incubate at 37 °C for 16:00:00 . This time allows non-neutralized viruses to infect cells and transcribe viral RNA. 16h

(DAY 2) cDNA synthesis

3h 7m

23

Note

In order to relate the counts of each barcoded vRNA in the sequencing data to the number of vRNA represented in the infected cells, we need to add a spike-in control at a known concentration to each well. We generate an RNA spike-in control using in vitro transcription. This construct is similar to the barcoded HA vRNA, but contains GFP in place of the HA ectodomain. A number of known barcodes are associated with the RNA spike-in control and these barcodes are used to normalize the counts for each barcoded HA in the eventual sequencing data. An detailed description of how to generate this construct is provided in the associated manuscript.

*We describe these next steps using the iScript Select Synthesis Kit and the iScript Sample Preparation Buffer from BioRad, which we have tested in this method for performing cDNA synthesis of viral RNA (vRNA) from infected cells without the need for RNA extraction. Other commercially available kits could be used for this method if preferred.

- 24 Approximately 15 hours into the incubation period of the cells and virus, start thawing reagents from the iScript Select Synthesis Kit (BioRad) needed to perform cDNA synthesis (5x reaction mix, nuclease-free water, GSP enhancer, gene-specific primer, iScript enzyme mix) and a 200 pM aliquot of the RNA spike-in control. Most of the 1h



reaction mix components can be thawed  Room temperature but the the enzyme mix and the RNA spike-in control should be thawed  On ice .

25 When your RNA spike-in control is thawed, mix 15m

 55 μ L of 200 pM RNA spike-in control into

 5445 μ L iScript sample preparation reagent , thereby diluting the spike-in control 1:100 for a final concentration of 2 pM. You will end up with 5.5 mL total volume of lysis buffer using these specific amounts, which is sufficient for one plate. Adjust the above amounts according to how many plates of neutralization assays you set up.

26 When your cDNA synthesis reagents have thawed, prepare a cDNA synthesis mastermix. 15m

We use an 18 μ L final reaction volume for each well, typically preparing sufficient mastermix for 100 reactions per plate to allow excess prior to distributing the the mastermix to PCR tubes. For one plate, your mastermix can be prepared like so:

 400 μ L 5x reaction mix

 900 μ L nuclease-free water

 200 μ L GSP enhancer

 200 μ L gene-specific primer (2 uM stock)

 100 μ L iScript enzyme mix

Please adjust these amounts according to how many plates of neutralization assays you are running.

Aliquot  18 μ L cDNA synthesis mastermix into 8-strip PCR tubes or a 96-well PCR plate.

Note

The gene-specific cDNA synthesis primer used is:
cDNA_Fwd: 5' CTCCCTACAATGTCGGATTTGTATTTAATAG-3'

This primer binds immediately upstream of the barcode in the vRNA.



Safety information

The following section should be carried out in a biosafety cabinet because the steps involve influenza virus. Current and recent strains of human influenza virus are biosafety-level 2, but some historical human strains and animal strains are biosafety level 3 or higher. If you aren't sure of the biosafety level of the strains you are working with, check with your biosafety committee.

Filter tips are used for these steps to prevent issues with aerosol hazards and reduce RNase contamination.

Prepare a beaker of 10% bleach immediately before performing any of the steps for rinsing pipette tips. Be sure to rinse tips here after interaction with virus samples, and seclude tips to be disposed with the rest of your lab's virus waste.

- 28 Remove the supernatant from the plate using a multichannel pipette, taking care not to disturb the cells and pipette this directly into bleach solution. 5m
- 29 Wash the infected cells once with  150 μ L PBS per well. This wash step removes any residual virus from the supernatant and improves the cell lysis efficiency. Take care not to disturb the adhered cells when performing this wash step. When removing this liquid, pipette directly into bleach, and rinse tips with bleach solution. Examine the wells of the plate to ensure that residual liquid is completely removed from the wells of the plate prior to adding lysis buffer. This is essential as if residual liquid remains, this will dilute the spike-in control and result in noise in the measurements collected with this assay. 10m
- 30 Add  50 μ L lysis buffer (with the RNA spike-in control added) to each well. Wait 2-5 minutes for cells to lyse. Examine wells under the microscope to determine that cell lysis has occurred. This lysis buffer is gentle, expect that nuclei will remain adhered to the plate, though the cell borders will be less defined following lysis. 5m
- 31 After 2-5 minutes, transfer lysate to a new 96-well PCR plate so that the lysis of cells will not continue. 2m
- 32 Add  2 μ L cell lysate to the strip tubes (or 96-well PCR plate) containing  18 μ L iScript cDNA Select Synthesis mastermix 4m
- 33 Transfer the strip tubes (or 96-well PCR plate) containing cDNA synthesis reaction mixes to the thermocycler with the lid heated to  60 $^{\circ}$ C



Set the following protocol:

33.1 Incubate at 42 °C 01:00:00

1h

33.2 Inactivate enzymes at 85 °C 00:05:00

5m

33.3 Hold at 4 °C

*This is a potential stopping point, samples could be stored at -20 °C overnight, if needed before progressing to the next step of the assay.

(DAY 2) Round 1 PCR: adding overlap for indexing primers

35m

34 During cDNA synthesis, prepare Round 1 PCR mixes.

10m

Note

The following primers are used for Round 1 PCR:

Rnd1_Fwd: 5'-

GTGACTGGAGTTCAGACGTGTGCTCTTCCGATCTCTCCCTACAATGTCGGATTTGTATTTAATAG
-3'

Rnd1_Rev: 5'-AGTAGAAACAAGGGTGTTTTCTTATATTTCTGAAATCC-3'

For one sample, the mix is as follows:

1.5 µL Forward Primer

1.5 µL Reverse Primer

25 µL KOD Hot Start Master Mix

17 µL H₂O

5 µL template from cDNA synthesis

= 50 µL total volume

The following mix is enough to prepare for 96 samples:

150 µL Fwd Primer

150 µL Rev Primer



🧪 2500 µL KOD Hot Start Master Mix

🧪 1700 µL H₂O

= 🧪 4500 µL mastermix total volume

Aliquot 🧪 45 µL mastermix per well in a 96-well PCR plate.

Add 🧪 5 µL template . Seal the 96-well PCR plate with a PCR plate seal that is appropriate for thermal cycling and cold storage, such as (**#MSF1001, BioRad**).

35 Set the PCR plate containing Round 1 PCR reactions in thermocycler with the lid heated to 🌡️ 100 °C
Set the following protocol:

35.1 Initial denaturation

🌡️ 95 °C ⌚ 00:02:00

2m

35.2 Denaturation

🌡️ 95 °C ⌚ 00:00:20

20s

35.3 Annealing

🌡️ 56 °C ⌚ 00:00:10

10s

35.4 Extension

🌡️ 70 °C ⌚ 00:00:20

20s

35.5 Repeat previous three PCR cycle steps for denaturation, annealing, and extension 19 more times

15m 50s

35.6 Final extension

🌡️ 70 °C ⌚ 00:02:00

2m

35.7 Hold 🌡️ 10 °C



*This is a potential stopping point, samples could be stored at  -20 °C overnight, if needed before progressing to the next step of the assay.

(DAY 2) Round 2 PCR: adding indices

40m

36

Note

The steps below describe how to add 10 bp unique dual indices to the samples, which is the method we use in our lab for demultiplexing these samples after an Illumina NextSeq run. Alternate methods for indexing could be used.

For this step, we ordered dual indices based on the index sequences designed by Twist Biosciences (<https://www.twistbioscience.com/resources/protocol/Unique%20Dual-Index-Sequences-reference-spreadsheets-and-sample-sheet-templates>) inside custom indexing primers ordered through IDT which have the following format:

UDI_i7: 5'-

CAAGCAGAAGACGGCATACGAGATnnnnnnnnnnGTGACTGGAGTTCAGACGTGTGCTCTTCCGATCT-3'

UDI_i5: 5'-

AATGATACGGCGACCACCGAGATCTACACnnnnnnnnnnACACTCTTTCCCTACACGACGCTCTTCCGATCT-3'

Indexing primer stock mixes are prepared at 5 uM final concentration, with i7 and i5 indexes pooled at equal concentration (i.e. 2.5 uM UDI_i7 and 2.5 uM UDI_i5).

*Note, we observed an error in sequencing of barcodes that start with 'GG' in some wells where indexes also contained a 'GG' sequence. We advise that when designing the virus library for this method, that barcodes starting with 'GG' be avoided.

37 Prepare mix for Round 2 PCR. For one sample, the reaction mix is as follows:

20m

 2.4 µL Indexing Primer Mix (5 uM stock, contains equal concentration mix of fwd and rev indexing primer pair)

 20 µL KOD Hot Start Mastermix

 16.6 µL H₂O

 1 µL Round 1 PCR product

=  40 µL total volume



We prepare a mastermix of KOD and H₂O and apply this first to the plate, then add the primers. The following mix is enough to prepare for 96 samples:

🧴 2000 μ L KOD

🧴 1660 μ L H₂O

= 🧴 3660 μ L mastermix total volume

Transfer 🧴 36.6 μ L mastermix into each well of a 96-well PCR plate.

Add 🧴 2.4 μ L Indexing Primer Mix (each mix of primers should match the number assigned to your sample).

Add 🧴 1 μ L Round 1 PCR result .

Seal the 96-well PCR plate with a PCR plate seal that is appropriate for thermal cycling and cold storage.

38 Set 96-well PCR plate containing the Round 2 reaction mixes in thermocycler with the lid heated to 🌡️ 100 °C

Set the following protocol:

38.1 Initial denaturation

🌡️ 95 °C ⌚ 00:02:00

2m

38.2 Denaturation

🌡️ 95 °C ⌚ 00:00:20

20s

38.3 Annealing

🌡️ 66 °C ⌚ 00:00:10

10s

38.4 Extension

🌡️ 70 °C ⌚ 00:00:20

20s

38.5 Repeat previous three PCR cycle steps for denaturation, annealing, and extension 19 more times

16m

38.6 Final extension

🌡️ 70 °C ⌚ 00:02:00

2m

38.7 Hold 🌡️ 10 °C



*This is a potential stopping point, samples could be stored at  -20 °C overnight, if needed before progressing to the next step of the assay.

(DAY 2) Pooling & Preparing for Sequencing

1h 22m

39

Note

Lastly, we will pool the samples and run on an agarose gel to remove any residual indexing primers, and remove any Round1 PCR product from the Round2 sample. Following this step, a magnetic bead cleanup step is performed to insure that the sample is in an appropriate buffer for next-generation sequencing. The final band for the indexed sample should be 181 bps in length.

40 Prepare a 1% agarose gel. This can be done during Round 2 PCR.

41 Pool Round 2 PCR products at equal volume (enough so you have at least ~200uL total volume).

*We pool using a multichannel, first pooling all columns of the plate into a single 8-strip (5 uL from each sample), then pooling all 8 of the pools for each row of the plate. This will result in total volume of 480 uL per plate. It is not necessary to prepare this total volume for sequencing, only approximately ~100 uL needs to be gel extracted and bead purified for sequencing, but we have found it useful to pipette a larger volume (5 uL as opposed to 2 uL) per sample when pooling to ensure that all samples are included in the final pool.

42 Run  100 µL Round 2 PCR results pool on your gel from step 33 at 85 V

40m

 00:40:00 . We often run  30-40 µL of the sample per well on the agarose gel (with loading buffer), this results in 3-4 lanes of sample to be cut out and processed for sequencing.

43

**Note**

Since you are submitting the results of this gel for sequencing, it is important to ensure it is exposed to as little contamination from prior runs and ultraviolet light as possible. With this in mind, we use a blue light and Sybr Safe Gel Stain (Invitrogen, S33102) for visualization of the gel. We also use a layer of plastic wrap between the gel and the light source that insures that the sample will not be contaminated by prior runs with the same indexing primers. If you are running multiple plates on the same day that were prepared with the same indexes, use separate gels for each plate.

Cut the band out of the gel, and extract the DNA using a Nucleospin Gel Extraction Kit (740609, Takara), or alternate gel extraction kit. Transfer the eluate to a strip tube for magnetic bead cleanup with AMPure XP SPRI Reagent (Beckman Coulter).

- 44 Perform a magnetic bead cleanup on your sample using AMPure XP SPRI Reagent to remove residual salts from the gel extraction:
- 44.1 Add 2X the volume of your sample of AMPure XP SPRI Reagent (i.e. if you eluted in 40 uL add 80 uL Ampure XP beads).
- 45 Incubate Room temperature 00:05:00 . 5m
- 45.1 Load strip of tubes onto magnetic block. Incubate 00:05:00 . 5m
- 45.2 Pull off liquid while being careful to avoid disrupting magnetic beads. Wash twice, using 150 µL 80% EtOH each time (again, taking care to not disrupt the magnetic beads). Extract all liquid from tube and wait for pellet to start drying. 5m
- 45.3 When the pellet is dry (but not so dry that it has started to crack, approximately 00:10:00) resuspend in 40 µL elution buffer . Remove strip of tubes from magnetic block and repeatedly pipette the liquid up and down to thoroughly mix the beads into the elution buffer. Incubate at Room temperature for 00:05:00 . 15m
- 45.4 Place strip tubes on magnetic block, incubate 00:05:00 . 5m
- 45.5 Once the beads are adhered to the magnet, remove the residual liquid from the beads, careful not to disturb the beads, and transfer this liquid to new 1.5 uL tube. 1m



- 46 Quantify total DNA concentration, for this we use a Qubit. We typically obtain  40 µL of sample at  0.005 µg/µL 3m
- 47 Label, appropriately dilute, and submit sample for sequencing as determined by the sequencing service you are using. We use Illumina NextSeq for these runs (a P1 run for 1 plate and a P2 should be used for 2-4 plates). We submit the 181 bp amplicon for sequencing with a 50 bp read length. We aim to obtain an average coverage of between 500,000-1,000,000 reads/well. This is likely higher coverage than necessary, however, by over-sequencing we allow for variability in loading of the different wells, which is helpful given that we are not performing any sample normalization prior to pooling. 3m

Data Analysis

- 48 Following demultiplexing of the sequencing run, sequencing data is analyzed using the modular analysis pipeline developed by the Bloom lab for processing high-throughput sequencing-based neutralization assays. This pipeline is available at <https://github.com/jbloomlab/seqneut-pipeline>

See associated manuscript for a detailed description of this analysis method.

Briefly, this pipeline takes in the FASTQ files from a sequencing run, calculates the counts for each barcoded viral variant and the RNA spike-in control in each well. The ratio of barcoded viral variant to spike-in control in each well containing serum is normalized to the ratio of barcoded viral variant to spike-in control in the no-serum control well to calculate a fraction infectivity. Then, neutralization titers are computed by fitting Hill-curve style neutralization curves to the fraction infectivity values using the **neutcurve** package; see the documentation for the details of these curves. The titers represent the reciprocal serum dilutions at which half the viral infectivity is neutralized. For more details, please see the description at <https://github.com/jbloomlab/seqneut-pipeline>