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Version 4

SARS-CoV-2 Mpro fluorescence dose response V.4

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Haim Barr^{1,2}, Noa Lahav^{1,2}

¹Weizmann Institute of Science; ²ASAP Drug Discovery Consortium

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ASAP Discovery



Noa Lahav

Weizmann Institute

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Protocol status: Working

We use this protocol and it's working

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Abstract

This is a **functional**, **biochemical assay** used to identify treatments for viral infectious disease that target SARS-COV-2 Main Protease (MPro).

Utilizing a direct enzyme activity measurement method, the experiment was performed in a 384-well plate reading the fluorescence intensity. This assay tested the mode of action of inhibition.

It was developed at the Weizmann Institute of Science, as a part of the ASAP Drug Discovery Consortium.

Experiment Assay Concentrations

А	В	С
Reagent	Final Assay Concentrati on	Units
SARS Mpro	5	nM
SARS Substrate	375	nM
HEPES (pH 7.3)	20	mM
NaCl	50	mM
Glycerol	10	% by volume
TWEEN 20	0.01	% by volume
TCEP	1	mM

For more information, please check out the "Materials" Section

Guidelines

Plate Information:

Total Assay Volume: 20 μL

Compounds Top Assay Concentration: 100 µM

Dilution Factor: 2

Dose Response Points: 12 Number of Replicates: 2 Backfill with DMSO: Yes



Materials

Assay Buffer Reagents (Concentration listed are Stock Solution Concentrations)

- 1. [M] 40 millimolar (mM) SHEPES 1M Solution pH 7.3 Fisher Scientific Catalog #AAJ16924K2 (or similar)
- 2. [M] 100 millimolar (mM) Sodium chloride Merck MilliporeSigma (Sigma-Aldrich) Catalog #S9888-25G (or similar)
- 3. [M] 50 % volume Slycerol Merck MilliporeSigma (Sigma-Aldrich) Catalog #G5516 (or similar)
- 5. [M] 1000 millimolar (mM)
 - Tris(2-carboxyethyl)phosphine hydrochloride Merck MilliporeSigma (Sigma-Aldrich) Catalog #75259 (TCEP) (or similar)
- *Note: all components are added fresh to the assay buffer before each experiment

Additional Reagents:

[M] 710 micromolar (μM) SARS MPro Enzyme

- The Enzyme original stock was originally [M] 750 micromolar (μM) and was diluted to create aliquots of [M] 20000 nanomolar (nM) using a **storage buffer** (50 mM Tris pH 7.5, 1 mM DTT, 50 mM NaCl, 1 mM EDTA, 50% Glycerol).
- Before an experiment, the 20000 nM aliquots were **diluted with Assay Buffer** to create a IMI 10 nanomolar (nM) solution to be loaded into the Combi.

[M] 20000 micromolar (µM) SARS MPro Substrate

- SARS MPro Substrate Stock ([5-FAM]-AVLQSGFR-[Lys(Dabcyl)-K-amide) was purchased and dissolved in
 DMSO and yielded a concentration of [M] 20000 micromolar (μΜ)
- Before an experiment, the SARS MPro Substrate Stock had an *intermediate dilution step* with **DMSO** to yield a [M] 100 micromolar (μM)
 SARS MPro Substrate Solution. Then, before adding the SARS MPro Substrate to the Combi, it was diluted again with **Assay Buffer** to yield a concentration of [M] 750 nanomolar (nM)
 The final concentration of **SARS MPro Substrate** for the assay was [M] 375 nanomolar (nM)

Troubleshooting

Safety warnings

Please be sure to wear proper Personal Protective Equipment (PPE) while performing this experiment.



Before start

Note: Inhibitor compounds stock concentration is 20 mM. Compounds are pre-dispensed into 384 plates and stored at -200°C until use.



Prepare 384 Well Plate

- PRIME Multi-Drop Combi Tube Dispensing Cassette with **Assay Buffer** by selecting the **PRIME** button on the Combi Dispenser until the tubes are filled completely.
- 1.1 **DISPENSE** 4 10 µL Assay Buffer to Columns 1 and 23 of assay plate
 - Note: These will represent the *inhibitor control columns* (Contain: Substrate, Assay Buffer, DMSO; no experimental compounds)
- 1.2 **EMPTY** Multi-Drop Combi Tube Dispensing Cassette (by selecting the **EMPTY** button on the Combi Dispenser until the tubes of the cassette are emptied). Discard the Assay Buffer discharged from the cassette.

Prepare Reagents

- PRIME Multi-Drop Combi Tube Dispensing Cassette with

 [M] 10 nanomolar (nM) SARS MPro | by selecting the PRIME button on the Combi

 Dispenser until the tubes were filled completely.
 - **Note:** Be sure to cycle dispensing several times on a a clean plate lid (This confirms there are no bubbles in the Dispensing Cassette).
- 2.1 **DISPENSE** Δ 10 μL [M] 10 nanomolar (nM) SARS MPro to Columns 2 through 22 and also Column 24.

Note:

- [M] 10 nanomolar (nM) SARS Mpro is two times the final concentration for the assay.

 It is diluted to be a final concentration of [M] 5 nanomolar (nM) SARS MPro .
- Column 2 and Column 24 are *neutral control columns* (Contain: Enzyme, Substrate, DMSO; no experimental compounds)
- 2.2 **EMPTY** Multi-Drop Combi Tube Dispensing Cassette (by selecting the **EMPTY** button on the Combi Dispenser until the tubes of the cassette are emptied). Discard the [M] 10 nanomolar (nM) SARS MPro discharged from the cassette.
- 3 **CENTRIFUGE** § 15000 rpm, Room temperature, 00:01:00 plate to remove bubbles

4 **INCUBATE** plate for (5) 00:15:00 at 8 Room temperature

1m

15m



- PRIME Multi-Drop Combi Tube Dispensing Cassette with Assay Buffer by selecting the PRIME button on the Combi Dispenser until the tubes are filled completely. Then, EMPTY the Multi-Drop Combi Tube Dispensing Cassette (by selecting the EMPTY button on the Combi Dispenser until the tubes of the cassette are emptied). Discard the Assay Buffer discharged from the cassette.
- PRIME Multi-Drop Combi Tube Dispensing Cassette with

 [M] 750 nanomolar (nM) SARS Substrate by selecting the PRIME button on the Combi Dispenser until the tubes were filled completely.
 - **Note:** Be sure to cycle dispensing several times on a a clean plate lid (This confirms there are no bubbles in the Dispensing Cassette).
- 6.1 **DISPENSE** Δ 10 μL [M] 750 nanomolar (nM) SARS Substrate into Columns **1 through 24** (the full plate)

Note:

- IMJ 750 nanomolar (nM) SARS Substrate is two times the final concentration for the assay. It is diluted to be a final concentration of

 IMJ 375 nanomolar (nM) SARS Substrate
- 7 **CENTRIFUGE** plate at 15000 rpm, Room temperature, 00:01:00 in plate centrifuge to remove bubbles
- 8 INCUBATE plate at Room temperature for 00:30:00

 A Make sure the plate is protected from light!

Recommended: Clean/Empty the Multi-Drop Combi Reagent Dispenser and Dispensing Cassette during this incubation step

Read Plate Fluorescence

9 **READ** and **RECORD** the plate Relative fluorescence units (RFU) via the **"SARS Endpoint protocol"** on the **PHERAstar FS Control Software**.

Expected result

Gain 300 should yield ~10,000 RFU in full reaction and ~6,000 RFU in Buffer Control

1m

