

Feb 27, 2019

Sanger Sequencing

 In 1 collection

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Celeste M M. Karch¹, Rita Martinez¹, Jacob Marsh¹

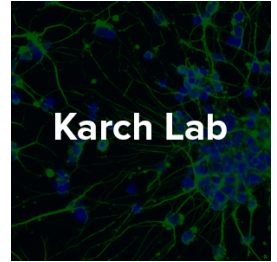
¹Washington University in St Louis

Neurodegeneration Method Development Community
Tech. support email: ndcn-help@chanzuckerberg.com



Celeste M M. Karch

Washington University in St Louis



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Protocol status: Working

We use this protocol and it's working

Created: February 18, 2019

Last Modified: February 27, 2019

Protocol Integer ID: 20467



Attachments



Comprehensive

Genomi...

31KB

Guidelines

This protocols is part of the Screening Edited iPSC Clones collection.

Safety warnings

⚠ Please refer to the SDS (Safety Data Sheet) for information about hazards, and to obtain advice on safety precautions.

Before start

After identifying iPSC clones with the appropriate banding pattern, gDNA from iPSC clones of interest should be sequenced in order to confirm genotype. To do this, set up a sequencing reaction as outlined below.

Perform PCR on gDNA using the following TouchDown Protocol (50µL reaction). The PCR amplicon size should be between 300-400 bp. The primers used for the RFLP screening can also be used for Sanger Sequencing.





PCR

- 1 Set up PCR on ice, add reagents in desired order (however it is best to add the polymerase and gDNA last).

	Volume	x# rxns
5x Green GoTaq Flexi Buffer	10 µl	
25mM MgCl ₂	6 µl	
25mM dNTPs	0.8 µl	
Forward Primer (10µM)	2 µl	
Reverse Primer (10µM)	2 µl	
GoTaq DNA Polymerase (5U/µL)	0.25 µl	
Milli-Q H ₂ O	26.95 µl	
QuickExtract gDNA	2 µl	
Total	50 µl	

Segment	Cycles	Temperature	Time
1	1	94°C	5 minutes
2	10	94°C	30 seconds
		65°C - 1°C/ cycle	30 seconds
		72°C	1 minute
3	35	94°C	30 seconds
		55°C	30 seconds
		72°C	1 minute
4	1	72°C	10 minutes
5	1	4°C	Forever

After product has been run in the thermocycler

- 2 Run  15 µL of each sample on a 2% gel to determine presence of PCR product.
- 3 Run product on gel at 150 V for  01:30:00 .



- 4 Image gel.
- 5 Save image.
- 6 After confirming the presence of PCR product, move on to sequencing reaction protocol below:

	Volume	x# rxns
PRC product	1 μ l	
Primer (10 μ M)	1 μ l	
Milli-Q H ₂ O	10 μ l	
Total	12 μ l	

Send out a Forward and a Reverse for each sample.