

Aug 17, 2023

sample_prep_urine.nan

Forked from a private protocol

 In 1 collection



DOI

<https://dx.doi.org/10.17504/protocols.io.j8nlkowkv5r/v1>

NAN KB¹, Mario Uchimiya², John Glushka², Leandro Ponce², Saraa Al Jawad², Laura Morris², Arthur Edison²

¹Network for Advanced NMR (NAN); ²University of Georgia

Leandro Ponce: Protocol review;

Saraa Al Jawad: Protocol review



NAN-KB UGA

Network for Advanced NMR

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.j8nlkowkv5r/v1>

Protocol Citation: NAN KB, Mario Uchimiya, John Glushka, Leandro Ponce, Saraa Al Jawad, Laura Morris, Arthur Edison 2023. sample_prep_urine.nan. protocols.io <https://dx.doi.org/10.17504/protocols.io.j8nlkowkv5r/v1>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: August 03, 2023

Last Modified: August 17, 2023

Protocol Integer ID: 85925

Keywords: protocol for nmr metabolomic, nmr metabolomic, urine sample, urine, sample

Funders Acknowledgements:

National Science Foundation

Grant ID: 194670

Disclaimer

This protocol is developed and maintained by Network for Advanced NMR (NAN). The protocol content here is for informational purposes only and does not constitute legal, medical, clinical, or safety advice, or otherwise; content added to this protocol is not peer reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol should not substitute for independent professional judgment, advice, diagnosis, or treatment. Any action you take or refrain from taking using or relying upon the information presented here is strictly at your own risk. You agree that neither the Company nor any of the authors, contributors, administrators, or anyone else associated with this protocol, can be held responsible for your use of the information contained in or linked to this protocol or any of our Sites/Apps and Services.



Abstract

This is a modified protocol for NMR metabolomics for urine samples. This method was originally proposed by:

Citation

Dona AC, Jiménez B, Schäfer H, Humpfer E, Spraul M, Lewis MR, Pearce JT, Holmes E, Lindon JC, Nicholson JK (2014)
. Precision high-throughput proton NMR spectroscopy of human urine, serum, and plasma for large-scale metabolic phenotyping.
Analytical chemistry.

<https://doi.org/10.1021/ac5025039>

LINK

See also:

Citation

Emwas AH, Luchinat C, Turano P, Tenori L, Roy R, Salek RM, Ryan D, Merzaban JS, Kaddurah-Daouk R, Zeri AC, Nagana Gowda GA, Raftery D, Wang Y, Brennan L, Wishart DS (2015)
. Standardizing the experimental conditions for using urine in NMR-based metabolomic studies with a particular focus on diagnostic studies: a review.
Metabolomics : Official journal of the Metabolomic Society.

<https://link.springer.com/article/10.1007/s11306-014-0746-7>

LINK



Materials

1. Chemicals and reagents

- Phosphate buffer in D₂O (1.5 M, pH 7.4, 1.11 mM DSS-d₆)

2. Equipment

- Calibrated micropipettes (100 μ L, 200 μ L, and 1000 μ L)
- Pippette tips
- 1.5-mL Eppendorf tubes
- 5-mm SampleJet NMR tubes from Bruker
- Centrifuge
- Vortex mixer

3. Study samples

- Urine samples

Before start

This protocol assumes that the original urine samples have >600 μ L.



7m 10s

1 Day-1/1

1.1 Thaw samples On ice or at 4 °C

1.2 Centrifuge the samples at 4 °C at 12000 x g for 00:05:00 min

5m



Note

- This step is to remove particles in the samples
- Sample tube and volume specifications vary depending on the study

1.3 Add 60 µL of the phosphate buffer to 540 µL of the supernatant for each sample



- Use 1.5-mL Eppendorf tubes

1.4 Vortex the samples at 4 °C for 00:02:00

2m

1.5 Centrifuge the samples at 4 °C for 00:00:10

10s



Note

This step is to remove samples attached to tube caps, and no centrifugation speed specified

1.6 Transfer 590 µL of the supernatant to an NMR tube for each sample

**Note**

No stickers/labels on the caps and tubes allowed

Citations

Dona AC, Jiménez B, Schäfer H, Humpfer E, Spraul M, Lewis MR, Pearce JT, Holmes E, Lindon JC, Nicholson JK. Precision high-throughput proton NMR spectroscopy of human urine, serum, and plasma for large-scale metabolic phenotyping.

<https://doi.org/10.1021/ac5025039>

Emwas AH, Luchinat C, Turano P, Tenori L, Roy R, Salek RM, Ryan D, Merzaban JS, Kaddurah-Daouk R, Zeri AC, Nagana Gowda GA, Raftery D, Wang Y, Brennan L, Wishart DS. Standardizing the experimental conditions for using urine in NMR-based metabolomic studies with a particular focus on diagnostic studies: a review.

<https://link.springer.com/article/10.1007/s11306-014-0746-7>