Jun 05, 2019 Version 2

RNA re-precipitation protocol V.2

DOI
dx.doi.org/10.17504/protocols.io.3signce

Cristian Riccio[1]

[1]University of Cambridge

Cristian Riccio
University of Cambridge

DOI: https://dx.doi.org/10.17504/protocols.io.3signce

Protocol Citation: Cristian Riccio 2019. RNA re-precipitation protocol. **protocols.io** **https://dx.doi.org/10.17504/protocols.io.3signce**

Protocol status: Working
We use this protocol and it's working

Created: June 05, 2019

Last Modified: June 05, 2019

Protocol Integer ID: 24106

Keywords: RNA reprecipitation, RNA purification, rna, residual contamination, precipitation protocol, poor nanodrop ratio

Abstract

If the RNA you have extracted is not pure and contains some residual contamination, as shown by poor Nanodrop ratios, you can reprecipitate the RNA, wash it and re-dissolve it to purify it.

Materials

MATERIALS

ethanol

Sodium Acetate Anhydrous Certified AR for Analysis Fisher Chemical **Fisher Scientific** **Catalog #**S/2120/53

Troubleshooting

1 Make Sodium Acetate 3M, pH 5.2.

2 Add 10% volume 3M sodium acetate pH 5.2 and 250% volume ethanol. So if your RNA solution is 100 ul, add 10 ul NaAc solution and 250 ul ethanol.

3 Mix well and put on dry ice for 30 min or -20 overnight

4 Centrifuge max speed 30 min at 4 C, remove supernatant

5 3 washes with 75% ethanol kept on dry ice. For example, add 900 ul 75% ethanol, centrifuge 5 min at max speed (~ 21,000 g), remove ethanol, three times.

6 10 min RT in hood drying

7 flick tubes for 1 minute

8 1 min at 80 C in heat-block

9 Resuspend in DEPC water

10 Assess the quality of the RNA using Nanodrop and Qubit.