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RNA Isolation for Tissue using TRIzol V.1

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Protocol status: In development














We are still developing and optimizing this protocol

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- 1 Add  1 mL TRIzol per  30 mg tissue and homogenize using handheld homogenizer.
- 2 Incubate at  25 °C for  00:05:00 to allow nucleoprotein complexes to dissociate.
- 3 Add  200 µL chloroform ( 20 % volume TRIzol) carefully and vortex to mix well.
- 4 Centrifuge at max speed for  00:15:00 at  4 °C .
- 5 Carefully remove the top aqueous phase and transfer to a new Eppendorf tube. The interphase and bottom organic phase can be saved for DNA and protein respectively.
- 6 Add  500 µL 100% isopropanol to the aqueous phase, mix by inversion and incubate at  -20 °C for a minimum of  02:00:00 .
- 7 Spin down at maximum speed for 30 mins to precipitate RNA.
- 8 Remove supernatant, and add  1 mL of  75 % volume ethanol to wash pellet.
- 9 Spin down at max speed for 15 minutes and remove supernatant.
- 10 Resuspend pellet in appropriate volume of nuclease free water.