

Sep 27, 2017

RNA extraction from Synechocystis sp. PCC 6803 with Trizol reagent

DOI

[dx.doi.org/10.17504/protocols.io.j3scqne](https://doi.org/10.17504/protocols.io.j3scqne)

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Protocol Citation: Dennis DD Dienst 2017. RNA extraction from Synechocystis sp. PCC 6803 with Trizol reagent. [protocols.io](#)
<https://dx.doi.org/10.17504/protocols.io.j3scqne>

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Protocol status: Working

Created: September 26, 2017

Last Modified: March 15, 2018

Protocol Integer ID: 8018

Keywords: rna extraction from synechocystis sp, rna extraction, synechocystis sp, pcc 6803 with trizol, synechocysti, rna, extraction

- 1 Label all required tubes and store on ice (or freezer, if not needed immediately)
- 2 Pre-cool all required centrifuges
- 3 Fill sterile 50 mL tube (Falcon) w/ ice and store in ice bath
- 4 Pour 20-25 mL cell culture (OD₇₅₀ < 1.0) to ice-filled tube (up to ~45 mL mark)
Note: avoid long transport of cell culture before harvest
- 5 spin down at 4000 - 5000 g and 4 ° C for 5 min
- 6 discard supernatant (w/ ice) into big beaker
Note: depending on strain/mutant some cells will get lost at this step
- 7 resuspend cell pellet in residual water (~ 1 mL)
- 8 transfer suspension into 2 mL (safe lock!) tubes (work on ice!)
- 9 spin down at ~13.000 g and 4° C for 15 sec
- 10 discard supernatant w/ pipet
Note: try to remove supernatant 'as quantitatively as possible'
- 11 resuspend pellet in 1 mL Trizol reagent
- 12 store at -20 °C or (better) -80 °C
- 13 incubate frozen samples at 65° C for 15 min under constant agitation
Note: if no shaking thermoblock is available, vortex once per minute

- 14 add 200 µL (ice cold) chloroform-isoamylalcohol (24:1) per 1 mL Trizol and vortex for 30 sec
- 15 spin down at ~11.000 g and 4° C for 10 min
- 16 transfer upper, aqueous phase to fresh 1.5 mL tube
- 17 add 1 Vol. (ice cold) phenol-chloroform-isoamylalcohol (25:24:1) Note: for RNA preparation phenol solution/mixtures should not be Tris-buffered
- 18 transfer upper, aqueous phase to fresh 1.5 mL tube
- 19 add 1 Vol. isopropanol (2-propanol), 10 µL 3 M Na-Acetat (pH 5.2) and 1 µL glycogen (RNA grade, Thermo)
- 20 incubate o/n at -20 for precipitation
- 21 spin down at ~13.000 g and 4° C for 30 min
- 22 remove supernatant and wash pellet w/ 70% EtOH (ice cold)
- 23 spin down at ~13.000 g and 4° C for 10 min
- 24 repeat steps 24 and 25
- 25 discard supernatant and air-dry RNA pellet for ~10 min
- 26 resuspend RNA Pellet in ~30 µL ultra-pure water and store at -20 or (better) -80°C