

Oct 07, 2022

RNA extraction from E. coli

DOI

dx.doi.org/10.17504/protocols.io.j8nlkw44wl5r/v1

An.Huang¹

¹XJTLU



An.Huang

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN ACCESS



DOI: https://dx.doi.org/10.17504/protocols.io.j8nlkw44wl5r/v1

Protocol Citation: An. Huang 2022. RNA extraction from E. coli. protocols.io

https://dx.doi.org/10.17504/protocols.io.j8nlkw44wl5r/v1

License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: October 06, 2022



Last Modified: October 07, 2022

Protocol Integer ID: 70925

Keywords: simple rna extraction procedure, rna extraction, rna, qpcr, extraction

Disclaimer

DISCLAIMER - FOR INFORMATIONAL PURPOSES ONLY; USE AT YOUR OWN RISK

The protocol content here is for informational purposes only and does not constitute legal, medical, clinical, or safety advice, or otherwise; content added to <u>protocols.io</u> is not peer reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol should not substitute for independent professional judgment, advice, diagnosis, or treatment. Any action you take or refrain from taking using or relying upon the information presented here is strictly at your own risk. You agree that neither the Company nor any of the authors, contributors, administrators, or anyone else associated with <u>protocols.io</u>, can be held responsible for your use of the information contained in or linked to this protocol or any of our Sites/Apps and Services.

Abstract

RNA extraction is a fundamental step in multiple experiments, for example, qPCR. This protocol helps conduct a simple RNA extraction procedure.

Materials

Buffer LY (added 1% volume of dithiothreitol), Buffer RB, RNA Wash Buffer, DEPC-Treated ddH₂O, RNA Columns, DNA Clearance Column, Collection Tubes, 1.5 mL RNase-free microfuge tube, Lysozyme buffer (0.4 mg/mL)

Troubleshooting



Preparation for experiment

- Grow an overnight bacterial culture in the appropriate media at an appropriate temperature.
- In the following day, take 4 1 mL from overnight culture and add into 4 10 mL LB media. Grow until the OD600 reads at 0.6-1.0.

RNA extraction



10m

- Harvest 1.5 mL culture (< 5×10%) by centrifugation at 3.000 rpm, 00:10:00 for 10 min in a 1.5 mL microcentrifuge tube.
- 4 Discard all supernatant.

Note

You may use a pipette to remove the remaining liquid at the bottom of the tube.

- Resuspend the pellet in $\[\] \]$ freshly prepared Elution Buffer (10mM Tris-HCL pH 8.5) containing lysozyme (0.4 mg/mL lysozyme for Gram negative bacteria). Mix by tapping gently.
- 6 Incubate the resuspended pellet at room temperature for 3-5 min for Gram-negative bacteria.
- 7 Add 400 μL Buffer LY. Mix gently.
- Transfer the cleared lysate to a DNA Clearance column pre-inserted in a 2 mL Collection Tube. Centrifuge at 13.000 rpm, 00:02:00. Discard the DNA Clearance column and save the flow-through.

2m

9 Transfer flow-through to the RNA binding column. Add 0.5 volume 100% ethanol to the lysate.



Note

For example: 250 μ L 100% ethanol for 500 μ L.

10 Centrifuge at 13000 rpm, 00:01:00 . Discard the collection tube with the flow through and put the column back to a new collection tube.

1m

Add \sqsubseteq 500 μ L Buffer RB to the column and centrifuge at \bigcirc 13.000 rpm, 00:00:30 . Discard the flow-through.

30s

Add another Δ 500 μ L RNA Wash Buffer to the column and centrifuge at

13000 rpm, 00:00:30 . Discard the flow-through.

30s

Note

Ethanol should be first added into RNA Wash Buffer before use.

Add another \triangle 500 μ L RNA Wash Buffer to the column and centrifuge at \bigcirc 13000 rpm, 00:00:30 . Discard the flow-through and collection tube, put the column into a new collection tube.

30s

14 Centrifuge the column at 🚯 13000 rpm , with the lid open, for another 🚫 00:01:00 .

1m

Place the column to a RNase-free 1.5 mL tube, add 50-100 μ L DEPC treated ddH₂O to the column and centrifuge at 13000 rpm, 00:02:00.

2m

Note

The RNA is in the flow-through.



16