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Purification of Total RNA from Cells Using Spin Technology

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Protocol status: Working

We use this protocol and it's working

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Abstract

Purification of Total RNA from Cells Using Spin Technology

Materials

Equipment:

Sterile,
RNase-free pipet tips
Microcentrifuge
Disposable gloves
TissueLyser
QIAshredder (cat#79656 Qiagen)

Reagent:

RNeasy mini kit
(cat#74106 Qiagen)
RNase
ZAP(cat#AM9780 Ambion)
 β -mercaptoethanol
96–100%
ethanol
PBS
0.25% trypsin

Troubleshooting



Method

- 1 Add 20 μ l β -ME per 1 ml Buffer RLT. Dispense in a fume hood. Buffer RLT containing β -ME can be stored at room temperature for up to 1 month.
- 2 Add volumes of ethanol (96–100%) as indicated on the bottle to concentrate buffer RPE to obtain a working Solution.
- 3 Harvest cells (do not use more than 1×10^7 cells) To lyse cells directly in the cell-culture vessel (up to 10cm diameter): Determine the number of cells. Completely aspirate the cell-culture medium, and proceed immediately to step 4. To trypsinize and collect cells (Cells grown in cell-culture flasks should always be trypsinized): Determine the number of cells. Detach cells from the dish or flask using trypsin, add medium containing serum to inactivate the trypsin, transfer the cells to an RNase-free centrifuge tube, and centrifuge at 1200rpm for 5 min. Completely aspirate the supernatant, and proceed to step 4.
- 4 Disrupt the cells by adding Buffer RLT. For direct lysis of cells: Add the appropriate volume of Buffer RLT as below to the cell-culture dish. Collect the lysate with cell scraper. Pipet the lysate into a microcentrifuge tube. Vortex or pipet to mix, and ensure that no cell clumps are visible before proceeding to step 5.

A	B
Dish diameter (cm)	Volume of Buffer RLT (μ l)
<6	350
6-10	600

For pelleted cells: Loosen the cell pellet thoroughly by flicking the tube. Add the appropriate volume of Buffer RLT as below. Pipet to mix, and proceed to step 5.

A	B
Number of pelleted cells	Volume of Buffer RLT (μ l)
< 5×10^6	350
$5 \times 10^6 - 1 \times 10^7$	600



- 5 Homogenize the lysate. Pipet the lysate directly into a QIAshredder spin column placed in a 2 ml collection tube, and centrifuge for 2 min at full speed.
- 6 Add 1 volume of 70% ethanol to the homogenized lysate, and mix well by pipetting.
- 7 Transfer up to 700 μ l of the sample, including any precipitate that may have formed, to an RNeasy spin column placed in a 2 ml collection tube. Close the lid gently, and centrifuge for 15 s at 10,000 rpm. Discard the flow-through *If the sample volume exceeds 700 μ l, centrifuge successive aliquots in the same RNeasy spin column. Discard the flow-through after each centrifugation.*
- 8 Add 700 μ l Buffer RW1 to the RNeasy spin column. Close the lid gently, and centrifuge for 15 s at 10,000 rpm to wash the spin column membrane. Discard the flow-through
- 9 Add 500 μ l Buffer RPE to the RNeasy spin column. Close the lid gently, and centrifuge for 15 s at 10,000 rpm to wash the spin column membrane. Discard the flow-through.
- 10 Repeat step 7.
- 11 Place the RNeasy spin column in a new 1.5 ml collection tube. Add 30–50 μ l RNase-free water directly to the spin column membrane. Close the lid gently, and centrifuge for 1 min at 10,000 rpm to elute the RNA.

Note:

- 12 Always wear gloves and spray with RNaseZap and spread solution all over gloves
- 13 Cell pellets can be stored at -70°C for later use or used directly in the procedure. Frozen cell pellets should be thawed slightly so that they can be dislodged by flicking the tube.
- 14 Homogenized cell lysates from step 5 can be stored at -70°C for several months. Frozen lysates should be incubated at 37°C in a water bath until completely thawed and salts are dissolved. Avoid prolonged incubation, which may compromise RNA integrity. If any insoluble material is visible, centrifuge for 5 min at 3000–5000 x g. Transfer supernatant to a new RNase-free glass or polypropylene tube, and continue with step 6.
- 15 Perform all steps of the procedure at room temperature. During the procedure, work quickly.



- 16 Perform all centrifugation steps at 20–25°C in a standard microcentrifuge. Ensure that the centrifuge does not cool below 20°C