

May 16, 2020 Version 1

Protocol for removing ssDNA from dsDNA or RNA Samples (NEB #M0568) V.1



DOI

dx.doi.org/10.17504/protocols.io.7ryhm7w

New England Biolabs¹

¹New England Biolabs

New England Biolabs (NEB)

Tech. support phone: +1(800)632-7799 email: info@neb.com



New England Biolabs

New England Biolabs

OPEN  ACCESS



DOI: dx.doi.org/10.17504/protocols.io.7ryhm7w

External link: <https://neb.com/protocols/2018/08/06/protocol-for-removing-ssdna-from-dsdna-or-rna-samples-m0568>

Protocol Citation: New England Biolabs 2020. Protocol for removing ssDNA from dsDNA or RNA Samples (NEB #M0568).

protocols.io <https://dx.doi.org/10.17504/protocols.io.7ryhm7w>

License: This is an open access protocol distributed under the terms of the **Creative Commons Attribution License**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: September 27, 2019

Last Modified: May 16, 2020

Protocol Integer ID: 28184

Abstract

The protocol described below will enable degradation of up to 20 pmol of a 25 nt ssDNA (~ 200 ng).

In order to degrade larger amounts of ssDNA or ssDNAs longer than 25 nt, we recommend adding more enzyme instead of extending the reaction time.

Users should note that ssDNAs longer than 25 nt may form secondary structures that hinder Thermolabile Exonuclease I activity.

Materials

MATERIALS

 NEBuffer 3.1 - 5.0 ml **New England Biolabs Catalog #B7203S**

 Thermolabile Exonuclease I **New England Biolabs Catalog #M0568**

 Nuclease-free Water **New England Biolabs Catalog #B1500**

Safety warnings

 Please see SDS (Safety Data Sheet) for hazards and safety warnings.

1 Prepare a  20 μL reaction as follows:



Sample containing ssDNA (up to 20 pmol of a 25-mer)	x μl
NEBuffer 3.1 (NEB #B7203)*	2 μl **
Thermolabile Exonuclease I	1 μl
Nuclease-free water	to 20 μl ***

*Most PCR buffers are compatible.

**No reaction buffer is necessary if enzyme is added to a PCR reaction.

***Scale larger reaction volumes proportionally.

2 Incubate at  37 $^{\circ}\text{C}$ for  00:04:00 .



3 Stop reaction by heat-inactivation at  80 $^{\circ}\text{C}$ for  00:01:00 .