

Feb 02, 2025

Version 2

Protocol for Gibson Assembly (NEBuilder® HiFi DNA Assembly) V.2

DOI

<https://dx.doi.org/10.17504/protocols.io.n92ld8m48v5b/v2>

Carolina Lopez¹

¹Washington University



Carolina Lopez

Washington University

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.n92ld8m48v5b/v2>

Protocol Citation: Carolina Lopez 2025. Protocol for Gibson Assembly (NEBuilder® HiFi DNA Assembly). protocols.io <https://dx.doi.org/10.17504/protocols.io.n92ld8m48v5b/v2> Version created by [Carolina Lopez](#)

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: February 02, 2025



Last Modified: February 02, 2025

Protocol Integer ID: 119446

Keywords: using gibson assembly, protocol for gibson assembly, gibson assembly, procedure for cloning, cloning, dna

Abstract

Procedure for cloning using Gibson Assembly NEBuilder[®] HiFi DNA Assembly Master Mix



Isolation of Purified Vector:

- 1 *Gibson Assembly technology uses homologous recombination to assemble adjacent DNA fragments that share end-terminal homology. The optimal length of the homologous fragment ends region depends on the number and length of the fragments in the assembly reaction.*

- 2 1. Digest Vector with Restriction Enzymes:

4h

Component	Volume (uL)
DNA Plasmid	X uL for 10 ug of Plasmid
10x Cutsmart Buffer	5
Enzyme 1	2.5
Enzyme 2	2.5
Nuclease Free H2O	40-X

2. Incubate for 03:00:00 at 37 °C .

3. Add 10 µL of 6x loading buffer to reaction

4. Make 1% low melt-agarose gel.

- a) Mix 1 g of Agar with 100 mL of TAE Buffer.

- b) Microwave to boil agarose and let cool until you can touch bottle, but gel is not solid.

- c) Add 1.5 µL of EtBr to agarose and pour into DNA gel mold with 10 well comb.

- d) Let gel solidify.

5. Load 60 µL of reaction into well of gel

6. Run gel for 00:45:00 at 120V.

7. Visualize band with UV light and cut out section of gel with band and place in

1.5 mL tube.

8. Purify Band from gel with QIAquick Gel Extraction kit (Qiagen, 28704)

- a) Weigh gel fragment in 1.5 mL tube (this will be volume with 100mg gel = 100uL)

- b) Add 3 volumes of Buffer QG to 1 volume gel.

- c) Incubate at 50 °C for 00:10:00 . Vortex every 2-3 min to help break up gel.

- d) Add 1 gel volume of isopropanol to the sample and mix.

- e) Place Qiaquick column into collection tube and add sample mixture to column.

- f) Let incubate for 00:01:00 .



- g) Centrifuge for  00:01:00 at max speed at Room temperature. Discard Flowthrough.
- h) Add  750 μL of PE buffer to column.
- i) Centrifuge for  00:01:00 at max speed at Room temperature. Discard Flowthrough.
- j) Centrifuge for  00:01:00 at max speed at Room temperature to dry column.
- k) Place column into new labeled  1.5 mL tube and add  35 μL of NF H₂O.
- l) Centrifuge for  00:01:00 at max speed at Room temperature to elute DNA.
- m) Measure DNA concentration with the Nanodrop.
- n) Vector Concentration:

Generation of PCR Product:

3 1. Set up PCR Reaction:

25m

	Volume (μL)
SpfI Fwd (10 μM)	1
NotI Rev (10 μM)	1
dNTPs	1
Plasmid (20ng)	1
Buffer	5
MgCl ₂	3
H ₂ O	38

2. Run PCR Conditions:

- a)  95 °C  00:02:00
- b)  95 °C  00:00:30
- c)  60 °C  00:00:30
- d)  72 °C  00:02:00
- e) Repeat step 2-4 35 times
- f)  72 °C  00:10:00
- g)  4 °C hold
3. After reaction, take  5 μL of product and add  1 μL of gel loading buffer and run on 1% agarose gel to make sure there is a correct PCR product.
4. Add  5 μL of Cutsmart buffer and  1 μL of DpnI to the reaction and incubate at  37 °C for  00:05:00 .

5. Purify Band from gel with QIAquick Gel Extraction kit (Qiagen, 28704)

- Add  135 μL of Buffer QG to  45 μL PCR reaction.
- Add  45 μL of isopropanol to the sample and mix.
- Place Qiaquick column into collection tube and add sample mixture to column.
- Let incubate for  00:01:00 .
- Centrifuge for  00:01:00 at max speed at Room temperature. Discard

Flowthrough.

- Add  750 μL of PE buffer to column.
- Centrifuge for  00:01:00 at max speed at Room temperature. Discard

Flowthrough.

- Centrifuge for  00:01:00 at max speed at Room temperature to dry column.
- Place column into new labeled  1.5 mL tube and add  35 μL of NF H₂O.
- Centrifuge for  00:01:00 at max speed at Room temperature to elute DNA.
- Measure DNA concentration with the Nanodrop.
- PCR Product Concentration:

Gibson Assembly with HIFI DNA Assembly Mix (NEB, E2621S).

2h 24m

4 1. Calculate the molar ratios of Vector and PCR product used

2h 24m

- <https://nebiocalculator.neb.com/#!/ligation>
- I usually use 1 vector: 2 PCR ratio

2. Mix

Component	Volume(uL)
Vector (50ng)	X
PCR Insert	Y
NEB builder	10 uL
H2O	10-X-Y
Total Volume	20uL

	A	B	C	D	E	F	G
	Component	Length of DNA (bp)	Molar ratio	ng of DNA	Volume of 50ng/ul solution		
	Vector	3015	1	50	1 ul		



	A	B	C	D	E	F	G
	PCR Fragment	500	2	16.58	0.33 ul		
	H2O				8.67 ul		

3. Incubate for  01:00:00 at  50 °C .

4. Transform Product into E. coli

a) Add  2 μL of product to  50 μL of TOP10 cells.

b) Incubate for  00:20:00 on ice.

c) Heat shock bacteria in  42 °C waterbath for  00:01:00 .

d) Incubate on Ice for  00:03:00 .

e) Add  100 μL of SOC media and shake in warm room for  01:00:00 .

f) Plate bacteria onto LB-Antibiotic Plate. And incubate overnight in  37 °C warm room.