



Oct 27, 2020

Protein expression and extraction of hard-to-produce proteins in the periplasmic space of *Escherichia coli*

 In 1 collection

DOI

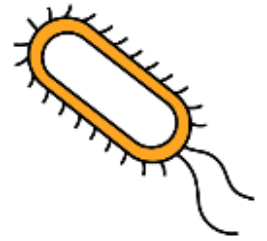
dx.doi.org/10.17504/protocols.io.bdr2i58e

Cristina Hernandez Rollan¹, Kristoffer Bach Falkenberg¹, Maja Rennig¹, Andreas Birk Bertelsen¹, Morten Norholm¹

¹Technical University of Denmark



Cristina Hernandez Rollan



Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.bdr2i58e>

Protocol Citation: Cristina Hernandez Rollan, Kristoffer Bach Falkenberg, Maja Rennig, Andreas Birk Bertelsen, Morten Norholm 2020. Protein expression and extraction of hard-to-produce proteins in the periplasmic space of *Escherichia coli*. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.bdr2i58e>

License: This is an open access protocol distributed under the terms of the **[Creative Commons Attribution License](#)**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited



Protocol status: Working

We use this protocol and it's working

Created: March 16, 2020

Last Modified: October 28, 2020

Protocol Integer ID: 34330

Keywords: LyGo, LPMO, Periplasmic expression, *E. coli*, protein expression, Periplasmic extraction, lytic polysaccharide monooxygenase, protein expression, proteins in the periplasmic space, protein, protein production, strain bl21, negative bacteria, t7 expression system,

Abstract

E. coli is a gram-negative bacteria used mainly in academia and in some industrial scenarios, as a protein production workhorse. This is due to its ease of manipulation and the range of genetic tools available.

This protocol describes how to express proteins in the periplasm *E. coli* with the strain BL21 (DE3) using a T7 expression system. Specifically, it describes a series of steps and tips to express "hard-to-express" proteins in *E. coli*, as for instance, LPMOs.

The protocol is adapted from Hemsworth, G. R., Henrissat, B., Davies, G. J., and Walton, P. H. (2014) Discovery and characterization of a new family of lytic polysaccharide monooxygenases. *Nat. Chem. Biol.*10, 122–126.

.

Guidelines

The periplasm of *E. coli* is often the preferred strategy to produce heterologous proteins in this bacterium as it provides the means for disulfide bond formation.

The choice of the signal peptide is of great importance to ensure correct and efficient translocation to the periplasm. In our lab, we routinely screen five different signal peptides: MalE^{SP}, OmpA^{SP}, PhoA^{SP}, DsBA^{SP}, and PelB^{SP}.



Materials

MATERIALS

⊗ BL21(DE3) Competent E.coli - 6×0.2 ml **New England Biolabs Catalog #C2527I**

⊗ IPTG **Bio Basic Inc. Catalog #IB0168.SIZE.100g**

⊗ EDTA **Fisher Scientific Catalog #16 004Y**

⊗ Kanamycin **Research Products International Corp (RPI) Catalog #K22000-25.0**

⊗ LB **Research Products International Corp (RPI) Catalog #L24400-2000.0**

⊗ Sucrose **Fisher Scientific Catalog #S25590B**

⊗ Trizma® base **Merck MilliporeSigma (Sigma-Aldrich) Catalog #93362**

Troubleshooting

Safety warnings



! This protocol describes the construction of GMO classified organisms. Make sure that the local GMO and safety legislations are respected.

Before start






Prepare a fresh transformation of your expression vector in *E. coli* BL21 DE3 cells.



Pre culture - Day 1

- 1 Pick a fresh colony of your BL21 (DE3) strain with your expression vector, and inoculate it in LB supplemented with relevant antibiotics. Grow the culture at  37 °C at 250 RPM shaking  Overnight . The volume of the overnight culture depends on the volume of the expression culture and should be at least 1/100 of the expression culture

Inoculation, Induction and expression - Day 2


- 2 Dilute the overnight culture 1:100 in fresh LB supplemented with relevant antibiotics
- 3 Grow the culture at  37 °C with 250 RPM shaking until an $OD_{600} = 0.5 - 0.6$
- 4 Move the culture into an incubator set to  18 °C with 180 RPM of shaking and grow the culture to $OD_{600} = 0.8 - 1.0$
- 5 Induce the expression by adding IPTG to a final concentration of  1 millimolar (mM)
- 6 Let the culture grow at  18 °C with 180 RPM shaking for  20:00:00

Note

Expression at low temperatures is recommended to enhance the solubility of some proteins.



Harvesting and periplasmic extraction - Day 3

10m

- 7 Spin the culture down at  8000 x g, 4°C, 00:20:00 and discard the supernatant

**Note**

Remove as much of the remaining liquid as possible from the centrifuge tube. This helps greatly in periplasmic extraction.



- 8 Resuspend the pellet in  3 mL of buffer TSE buffer (200 mM Tris-HCl pH 8, 500mM sucrose, 1mM EDTA) per gram of cells (this normalization can also be based on OD₆₀₀ where  12 µL TSE per OD Unit is added)


Note

Carefully resuspend the cells in the TSE buffer to avoid breaking the cells. A good tip is to use a sterile inoculation plastic loop to resuspend the pellet in the buffer before using a pipette tip.

- 9 Incubate the suspension at  Room temperature for  00:10:00

10m

- 10 Cold-shock the cells by adding  3 mL of ice-cold sterile MQ water per gram of cells (or alternatively,  12 µL of ice-cold water per every OD₆₀₀ Unit can be added).

- 11 Incubate the suspension  On ice for  00:10:00

- 12 Spin down the cells at  8000 x g, 4°C, 00:20:00 and collect the supernatant

Note

The supernatant contains the periplasmic extraction

- 13 Keep the extraction  On ice when working with it and at  4 °C for storage