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Preparation of bacterial cell lysate for SDS-PAGE (2022 iGEM)



Forked from [Preparation of bacterial cell lysate for proteomics \(LC-MS\) by freeze and thaw cycles](#)

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Protocol status: Working

We use this protocol and it's working

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Abstract

Preparation of bacterial cell lysate for SDS-PAGE



Preparation of bacterial cell lysate for SDS-PAGE

- 1 Grow the cells overnight, to an OD₆₀₀ above 1. If IPTG induction needed, at it around OD₆₀₀ 0.2-0.3 (keep IPTG stock solution at -20 freezer).
- 2 Take one 1 ml of OD₆₀₀ = 1 from each sample. Correct the sample volume to obtain an equivalent size pellet. For us, as quantified using NanoCym950 nanoparticles (equivalent to the size of *E. coli*), 1 OD₆₀₀ equals to 10⁸ nanoparticles per mL. We estimate each well on SDS-PAGE contains proteins from 2*10⁶ bacteria. 16h 
- 3 Spin down the bacterial cells for 1 min at 13,000 rpm in a microfuge. 10m
- 4 Aspirate any trace of supernatant with a vacuum line avoiding touching the pellet, but removing all liquid. 1m
- 5 To each cell pellet, add 250 µL of distilled water, resuspend the pellet. Add 250 µL 2x SDS sample buffer (see below). Heat with a 95 degree heating block for 10 minutes. Store the samples at -20 degree freezer until SDS-PAGE. 1m

Receipt

- 6 2x SDS sample buffer, works for Tris-Cl or MOPS based gels
 - Glycerol 20 ml
 - SDS 4 g
 - 0.5 M Tris-HCl pH 6.8 stock solution 25 ml
 - Bromo Phenol Blue 1 mg; dissolve and volume to 95 ml
 - add 5% β-Mercaptoethanol before usage