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## 🌐 Pollen Acetolysis V.2

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**Protocol status:** Working

We use this protocol and it's working

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## Abstract

Custom pollen acetolysis procedure using a slightly modified version of standard acetolysis.

## Troubleshooting

## Sample Prep:

- 1 Anthers are placed into a 1.5mL tube and crushed.
- 2 Add glacial acetic acid and stir.
- 3 Centrifuge for 3min at 3700rpm to allow pollen to form a pellet at the bottom of the tube. If pollen sticks to the side of the tubes, rotate the tubes 180 degrees and re-centrifuge for 30s.
- 4 Draw out liquid with a pipette without disturbing the pellet. If pellet is disturbed, return the liquid to the tube, re-centrifuge and draw out again.

## Acetolysis:

- 5 Add 300uL 9:1 acetic anhydride: sulfuric acid acetolysis solution to samples and immediately transfer to heat block.
- 6 Incubate at 90°C for about 3-10min with caps open. Finished samples should appear brown in color

## Glacial Acetic Acid Wash:

- 7 Add 300uL of glacial acetic acid to all heated samples to stop the reaction.
- 8 Cap then vortex thoroughly. Rinse all glassware with glacial acetic acid and then water.
- 9 Spin at 3700 rpm for 3min
- 10 Draw out liquid using pipette.
- 11 Repeat glacial acetic acid wash one or two additional times

## Water Wash:

- 12 Add 300uL of water using a clean pipette tip.
- 13 Cap samples, vortex then spin again.
- 14 Draw water down to pollen pellet.
- 15 Repeat water wash one more time.

## Ethanol Wash:

- 16 Add 300uL of 50% ethanol to each sample.
- 17 Vortex thoroughly.
- 18 Spin at 3700rpm 3min.
- 19 Draw liquid out with pipette down to the pollen pellet.
- 20 Repeat procedure with 70% and 95% ethanol.

## Add Glycerin:

- 21 Add 3 drops of 1:1 glycerol/water to pollen residue.
- 22 Place on heat block at 25°C and leave overnight.

23

Mix samples, then mount on glass slide for viewing.

### Adopted From:

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- 27 Pollen processing reference manual (2014) Florida Institute of Technology.