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Version 2

Pollen Acetolysis V.2

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Protocol status: Working

We use this protocol and it's working

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Abstract

Custom pollen acetolysis procedure using a slightly modified version of standard acetolysis.

Troubleshooting



Sample Prep:

- 1 Anthers are placed into a 1.5mL tube and crushed.
- 2 Add glacial acetic acid and stir.
- 3 Centrifuge for 3min at 3700rpm to allow pollen to form a pellet at the bottom of the tube. If pollen sticks to the side of the tubes, rotate the tubes 180 degrees and re-centrifuge for 30s.
- 4 Draw out liquid with a pipette without disturbing the pellet. If pellet is disturbed, return the liquid to the tube, re-centrifuge and draw out again.

Acetolysis:

- 5 Add 300uL 9:1 acetic anhydride: sulfuric acid acetolysis solution to samples and immediately transfer to heat block.
- 6 Incubate at 90°C for about 3-10min with caps open. Finished samples should appear brown in color

Glacial Acetic Acid Wash:

- 7 Add 300uL of glacial acetic acid to all heated samples to stop the reaction.
- 8 Cap then vortex thoroughly. Rinse all glassware with glacial acetic acid and then water.
- 9 Spin at 3700 rpm for 3min
- 10 Draw out liquid using pipette.
- 11 Repeat glacial acetic acid wash one or two additional times



Water Wash:

12

Add 300uL of water using a clean pipette tip.

13

Cap samples, vortex then spin again.

14

Draw water down to pollen pellet.

15

Repeat water wash one more time.

Ethanol Wash:

16

Add 300uL of 50% ethanol to each sample.

17

Vortex thoroughly.

18

Spin at 3700rpm 3min.

19

Draw liquid out with pipette down to the pollen pellet.

20

Repeat procedure with 70% and 95% ethanol.

Add Glycerin:

21

Add 3 drops of 1:1 glycerol/water to pollen residue.

22

Place on heat block at 25°C and leave overnight.



23

Mix samples, then mount on glass slide for viewing.

Adopted From:

24

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