



Mar 15, 2023

Phage infection and timed harvest of E. coli and B. subtilis cells

 [PubPub](#)

DOI

dx.doi.org/10.17504/protocols.io.j8nlkw2d5l5r/v1

Januka Athukoralage¹, Adair Borges¹

¹Arcadia Science

Arcadia Science



Arcadia Science

Arcadia Science

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.j8nlkw2d5l5r/v1>

Protocol Citation: Januka Athukoralage, Adair Borges 2023. Phage infection and timed harvest of E. coli and B. subtilis cells .
protocols.io <https://dx.doi.org/10.17504/protocols.io.j8nlkw2d5l5r/v1>

Manuscript citation:

Athukoralage J, Borges A, Reiter T. (2023). Challenges of isolating bacteriophage mRNA for chemical analysis.
<https://doi.org/10.57844/arcadia-j03a-mz33>



License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: February 22, 2023

Last Modified: January 30, 2024

Protocol Integer ID: 77401

Keywords: phage, bacteriophage, SPO1, T4, phages, bacteriophages, *E. coli*, *B. subtilis*, *Escherichia coli*, *Bacillus subtilis*, infection, infect, harvest, grow, produce, bacteriophage transcript, harvesting cells at different timepoint, harvest cell, harvesting cell, phage t4, infection, timed harvest, cell, enrichment for downstream analysis, rna

Abstract

This protocol outlines how to infect *E. coli* and *B. subtilis* with phage T4 and SPO1, respectively, and harvest cells at set time points.

Harvesting cells at different timepoints during infection allows you to evaluate when bacteriophage transcripts are most abundant in cells and maximize their enrichment for downstream analyses such as chemical analysis and RNA sequencing.

Materials

50 mL sterile Erlenmeyer flasks
5 mL sterile and capped test tubes
1.5 mL microcentrifuge tubes
Bechtop centrifuge capable of cooling
Shaking incubator
Shaking mixer
Liquid nitrogen and cryogenic container for safe handling
Freezer at -70 °C or above

Troubleshooting

Bacteriophage infection

- 1 Prepare 30 mL subcultures of *E. coli* and *B. subtilis* at 1:100 dilution (300 μ L in 30 mL) and grow to a target OD₆₀₀ of 0.4 in a shaking incubator (180 rpm). For both *E. coli* and *B. subtilis* this is ~2 h at 37 °C. Supplement the subculture media (LB) with 1 mM CaCl₂ and 1 mM MgCl₂.
- 2 Aliquot 500 μ L of cell culture into several 1.5 mL microcentrifuge tubes and inoculate with 50 μ L of phage at the correct dilution to reach your target MOI (see our protocol on MOI calculations, linked below).

Protocol



NAME

Calculating multiplicity of infection (MOI)

CREATED BY

Arcadia Science

[Preview](#)

- 3 Here, we are infecting *E. coli* with T4 and *B. subtilis* with SPO1. Be sure to simultaneously carry out a control experiment where you mock-infect 500 μ L *E. coli* and *B. subtilis* cultures with 50 μ L sterile SM buffer. Once inoculated, incubate the microcentrifuge tubes at 37 °C on a heated shaker block, shaking at 300 rpm.
- 4 Plate out phage and bacteria to calculate the actual MOI used in the experiment.

Cell harvest

- 5 Pre-cool a bench-top centrifuge to 4 °C and at desired time points, harvest cells by centrifuging at 5000 \times g for 3 min. For example, cells can be harvested at 5 min and 15 min in the hope of capturing both earlier and later infection stages. This timing will be highly phage-dependent!
- 6 Remove the supernatant from the pellet and immediately flash-freeze the pellets in liquid nitrogen. You must be speedy during this process to effectively halt phage infection. You can store bacterial pellets at -80 °C and use later for downstream applications such as RNA extraction.