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Version 3

## PCR Reaction Optimization V.3



Version 1 is forked from [WarmStart LAMP®](#)

DOI

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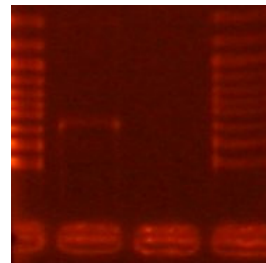
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**Protocol status:** Working

**We use this protocol and it's working**

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**Keywords:** pcr reaction optimization, nucleic acid amplification, thermo scientific pcr master mix kit, scientific pcr master mix kit

## Abstract

How to run nucleic acid amplification using the Thermo scientific PCR Master Mix kit.  
Each reaction produces 50  $\mu$ L.

For the original protocol, look at:  [PCR Master Mix Manual.pdf](#).

## Guidelines

Gloves must be worn at all times.

Use all precautions to avoid contamination when making reaction mixture.

Always pipette mix each reagent in aliquot before pipetting.

## Materials

### MATERIALS

 DNase/RNase free distilled water [Thermo Fisher Scientific Catalog #10977023](#)

- 70% ethanol solution in DI water
- RNAway
- Thermo Scientific Master Mix (2x)
- PCR primer mix (25  $\mu$ M)
- Target DNA or RNA
- RNase free water


## Troubleshooting



## Prepare Work Area



- 1  
Spray entire work area with 70% EtOH including pipettes, tip holder used for holding PCR tubes, and work surface. Wipe with a paper towel.
- 2  
Spray entire work area with RNAway.

## Gather Materials

- 3  
Take styrofoam container to Marley 527 (directly across from Marley 509) and fill halfway with ice.
- 4  
Set PCR tube holder on ice, and allow to cool for  00:03:00 .
- 5  
Transfer Master Mix, primers, RNase free water, and target tubes from freezer to PCR tube holder on ice.
- 6  
Allow reagents to thaw on ice
- 7  
Carefully obtain (2) 0.2 mL PCR tubes. Label one with "NTC" and the other "TARG". These will be your reaction vessels.

### Note

To avoid contamination when grabbing PCR tubes, only touch the outside of tubes. Avoid touching the inside of the caps of other tubes in this process. This is critical.

- 8  
Vortex mix all reagents for approximately  00:00:05 .
- 9  
Spin down all reagents for approximately  00:00:05 .



## Prepare Reaction

- 10 Add the following to your two tubes:

		Target	NTC
	PCR Master Mix	25 $\mu$ L	25 $\mu$ L
	Primer Mixture (25 $\mu$ M)	0.2 - 2.0 $\mu$ L	0.2 - 2.0 $\mu$ L
	Target	1 $\mu$ L	-
	Water	to 50 $\mu$ L	to 50 $\mu$ L
	Total	50 $\mu$ L	50 $\mu$ L

¶ Various primer concentrations are to be optimized. Start with 1.0  $\mu$ L (0.5  $\mu$ M).

- 11 Vortex mix the reaction mixture.

- 12 Spin down reaction mixture.

## Run PCR Reaction

- 13 Place reaction vessels into thermocycler.

- 14 Turn on thermocycler

- 15 Hit PROCEED to select a reaction cycle.

- 16 Scroll using the '<' and '>' keys to get to EPCR.  
Begin using the following program:

	Step	Temperature	Time	Number Cycles
	Denaturation	95°C	3 min	1



	Denaturation	95°C	30 s	30
	Annealing	58°C	30 s	
	Extension	72°C	60 s	
	Final Extension	72°C	10 min	1

17 Press PROCEED to begin