

Feb 22, 2019 Version 2

## PCR Reaction Optimization V.2

 Version 1 is forked from [WarmStart LAMP®](#)

DOI

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**Protocol status:** In development

We are still developing and optimizing this protocol

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## Abstract

How to run nucleic acid amplification using the Thermo scientific PCR Master Mix kit.  
Each reaction produces 50 µL.

For the original protocol, look at:  [PCR Master Mix Manual.pdf](#).

## Guidelines

Gloves must be worn at all times.  
Use all precautions to avoid contamination when making reaction mixture.  
Always pipette mix each reagent in aliquot before pipetting.

## Materials

### MATERIALS

 DNAse/RNAse free distilled water [Thermo Fisher Scientific Catalog #10977023](#)

- 70% ethanol solution in DI water
- RNAway
- Thermo Scientific Master Mix (2x)
- PCR primer mix (25 µM)
- Target DNA or RNA
- RNAse free water

## Prepare Work Area

- 1 Spray entire work area with 70% EtOH including pipettes, tip holder used for holding PCR tubes, and work surface. Wipe with a paper towel.
- 2 Spray entire work area with RNAway.

## Gather Materials

- 3 Take styrofoam container to Marley 527 (directly across from Marley 509) and fill halfway with ice.
- 4 Set PCR tube holder on ice, and allow to cool for  00:03:00 .
- 5 Transfer Master Mix, primers, RNase free water, and target tubes from freezer to PCR tube holder on ice.
- 6 Allow reagents to thaw on ice

## Gather Materials

- 7 Carefully obtain (2) 0.2 mL PCR tubes. Label one with "NTC" and the other "TARG". These will be your reaction vessels.

### Note

To avoid contamination when grabbing PCR tubes, only touch the outside of tubes. Avoid touching the inside of the caps of other tubes in this process. This is critical.

- 8 Vortex mix all reagents for approximately  00:00:05 .
- 9 Spin down all reagents for approximately  00:00:05 .

## Prepare Reaction

- 10 Add the following to your two tubes:

	Target	NTC
PCR Master Mix	25 µL	25 µL
Primer Mixture (25 µM)	0.2 - 2.0 µL	0.2 - 2.0 µL
Target	1 µL	-
Water	to 50 µL	to 50 µL
Total	50 µL	50 µL

Various primer concentrations are to be optimized. Start with 1.0 µL (0.5 µM).

- 11 Vortex mix the reaction mixture.

- 12 Spin down reaction mixture.

## Run LAMP Reaction

- 13 Place reaction vessels into thermocycler.

- 14 Turn on thermocycler

- 15 Hit PROCEED to select a reaction cycle.

- 16 Scroll using the '<' and '>' keys to get to PCR.

Begin using the following program:

Step	Temperature	Time	Number Cycles
Denaturation	95°C	3 min	1
Denaturation	95°C	30 s	30
Annealing	58°C	30 s	

	Extension	72°C	60 s	
	Final Extension	72°C	10 min	1

17 Press PROCEED to begin