



Feb 06, 2023

Version 1

🌐 PCR protocol for gene COXI neo-caledonian freshwater micro-crustaceans V.1

DOI

<https://dx.doi.org/10.17504/protocols.io.261ge4y4jv47/v1>

coline.royaux¹

¹Université Pierre et Marie Curie (Paris VI)



Coline Royaux

Université Pierre et Marie Curie (Paris VI), Muséum National...

Create & collaborate more with a free account

Edit and publish protocols, collaborate in communities, share insights through comments, and track progress with run records.

Create free account

OPEN  ACCESS



DOI: <https://dx.doi.org/10.17504/protocols.io.261ge4y4jv47/v1>

Protocol Citation: coline.royaux 2023. PCR protocol for gene COXI neo-caledonian freshwater micro-crustaceans. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.261ge4y4jv47/v1>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working



Created: October 19, 2021

Last Modified: February 06, 2023

Protocol Integer ID: 54230

Keywords: pcr protocol for gene coxi, pcr protocol, cytochrome oxydase, pcr, gene coxi, gene, streptocephalus with several primer

Abstract

This PCR protocol has been used to amplify the Cytochrome Oxydase I gene of neo-caledonian genera *Boeckella*, *Lynceus*, *Eulimnadia* and *Streptocephalus* with several primers.



- 1 Prepare the mix for the PCR depending on the quantity of DNA samples you want to amplify

Note

The quantities per well and for 96 DNA samples (with a 4 volumes margin) are detailed in the sub-steps

Safety information

Be careful to take enough margin to avoid a lack of reagent and, if possible, do a positive and a negative control

- 1.1 For each well, mix  12.44 μL distilled water ,
 2 μL DNA buffer for Taq polymerase ,  1 μL DMSO ,  1 μL BSA ,
 0.8 μL DNTP and  0.12 μL Taq polymerase . This mix is hereafter named "intermediary mix".

Note

Very few micropipettes are able to dose such a small amount so you'll have to make your mix for several wells at a time.

So, for 96 DNA samples with a 4 volume margin (100 volumes), mix

 1244 μL distilled water ,  200 μL DNA buffer for Taq polymerase ,
 100 μL DMSO ,  100 μL BSA ,  80 μL DNTP and
 12 μL Taq polymerase .

- 1.2 Add your primers to the mix,  0.32 μL selected H primer  10 picomolar (pM) and
 0.32 μL selected L primer  10 picomolar (pM) for each well. This mix is hereafter named "final reagent mix".



Safety information

You may have to use different primers in your wells depending on the individual and the gene you want to amplify. If so, distribute the intermediary mix in as much tubes as primer pairs you want to use and add your primers in the proper quantity to make your final reagent mix. Don't forget to add margins in each mix for each different pair of primers you use.

Note

So, for 96 DNA samples with a 4 volume margin (100 volumes) and if you use the same primers for all 96 wells, add  32 µL selected H primer and

 32 µL selected L primer

- 2 In each well, deposit  18 µL of the final reagent mix and  2 µL genomic DNA unless for the negative control.
For the positive control, use the DNA of the species on which the primer has been designed.
- 3 Close the PCR plate with a heated aluminium foil and centrifuge it enough to make sure the reagents are all in the bottom of the well.
- 4 To amplify targeted DNA sequences, place your plate in the thermocycler, set and launch the cycle as described in the following sub-steps
 - 4.1  00:05:00 at  94 °C 5m
 - 4.2 40 cycles of the following steps : 2m
 -  00:00:30 at  94 °C
 -  00:00:30 at  50 °C
 -  00:01:00 at  72 °C
 - 4.3  00:08:00 at  72 °C 8m



4.4 Infinite timing at 20°C so the samples are well preserved and you have some time to pick it up

5 To know if your amplifications worked properly, you now have to proceed to a gel electrophoresis of your PCR products

5.1 To prepare the gel,

Safety information

This step is to be made under an extractor hood with gloves, a blouse and glasses as BET is highly mutagenic and carcinogenic

mix 0.8 mg agarose and 40 mL TAE buffer , heat the preparation in the microwave for a few moments until it is homogeneous (you can agitate the preparation). Wait until the preparation is cold enough to touch its container and add $0.8\text{ }\mu\text{L BET}$.

5.2 Pour the preparation in a mold and let the gel polymerize with a bar to form wells to deposit your PCR products

5.3 Deposit the gel in the electrophoresis tank

5.4 Fill each well with $2.5\text{ }\mu\text{L PCR product}$ and $0.5\text{ }\mu\text{L DNA loading dye (6x)}$, don't forget to save a well to deposit $1.2\text{ }\mu\text{L molecular weight marker}$

5.5 Start electrophoresis in TAE buffer for $00:15:00$ at 200 Volts

15m

5.6 Look at your gel under Ultra Violet light to see the results