

Oct 18, 2019

PCR Amplification of Desired Gene

DOI

dx.doi.org/10.17504/protocols.io.8fuhtnw

laem Dusseldorf¹

¹Heinrich-Heine Universität Düsseldorf



Igem Dusseldorf

OPEN ACCESS



DOI: dx.doi.org/10.17504/protocols.io.8fuhtnw

Protocol Citation: Igem Dusseldorf 2019. PCR Amplification of Desired Gene. protocols.io

https://dx.doi.org/10.17504/protocols.io.8fuhtnw

License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: October 18, 2019

Last Modified: October 18, 2019

Protocol Integer ID: 28884



Materials

water

Green HF buffer

dNTPS

F Primer

R Primer

Template DNA

Phusion DNA Polymerase

1% agarose gel

10000x GelRed nucleic acid stain

1x TAE buffer

Load DNA ladder



1 Pipette PCR components in the following order. Always set up at least two replicates.

2

# React ions	[μΙ]	Ther mocy cler Condi tions	Roun ds
Water	32.5	1. 98°C for 0:30 min	
Gree n HF Buffe r	10.0	2. 98°C for 0:10 min	5x
dNTP s (10 mM)	1.0	3. X°C for 0:20 min	
F Prime r (10 μΜ)	2.5	4. 72°C for X min	
R Prime r (10 µM)	2.5	5. 98°C for 0:10 min	30x
Temp late DNA (10 ng/ µL)	1.0	6. 72°C for X min	
Phusi on DNA Poly mera se	0.5	7. 72°C for 5:00 min	
Total	50.0	8. 4°C until use	



- 3 For step 3, use the annealing temperature of the gene-specific sequences of your primers.
 - For step 4 and 6, adjust the extension time based on the amplicon size (30 s / kb).
- 4 Prepare a 1 % agarose gel by dissolving agarose in 1x TAE buffer by microwaving (~ 3 min, swirling every min).
 - Add 2.5 µL of 10000x GelRed nucleic acid stain per 50 mL gel and mix well by swirling. When the conical flask is almost cool enough to touch (3-5 min after microwaving), pour slowly in gel tray and then insert the combs. Use wide-wells for large PCR reaction volumes.
 - Gel will be solidified after 45-60 min. Remove combs and add enough 1x TAE buffer to cover the gel.
- 5 Load 2-3 µl DNA ladder and whole PCR samplesinto the gel pockets. Connect lid (DNA will migrate to the + electrode). Turn on power supply and run at 80-110 V depending on the thickness and size of the gel. Examine gel after 30-60 min of electrophoresis and run until the marker reached the desired height.
 - Extract bands of the expected height from the gel with and clean up the fragments with an extraction kit.
- 6 Thermocycler Step 3: 5 cycles with annealing temperature that is equal to the forward or reverse primer with lower Tm. Has to be adjusted for every gene.
- 7 Termocycler Step 4: Adjust the elongation time with 30 s for each kb.
- 8 Termocycler Step 6: 30 cylces with 72 °C for elongation and adjusted extention time. Change time with 30 s per kb.
- 9 If not enough DNA was produced run another PCR after DNA extraction with the purified amplicon as a template and skip the 5 initial rounds.
- 10 Other optimisation: Increase time to 40 sec/kb