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Protocol status: Working
We use this protocol and it's working

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PBMC- 01b Isolation of human PBMC from Whole Blood V.4

Marco Cosentino¹, Elisa Storelli², Alessandra Luini¹, Massimiliano LM Legnaro³, Emanuela Rasini³, Marco Ferrari³, Franca Marino³

¹Center for Research in Medical Pharmacology, University of Insubria (Varese, Italy);

²Center for Research in Medical Pharmacology, University of Insubria;

³University of Insubria

Elisa Storelli

Center for Research in Medical Pharmacology, University of I...

ABSTRACT

Separation and purification of PBMC from FRESH BLOOD: list of published work using this protocol

Kustrimovic, N., Comi, C., Magistrelli, L., Rasini, E., Legnaro, M., Bombelli, R., Aleksic, I., Blandini, F., Minafra, B., Riboldazzi, G., Sturchio, A., Mauri, M., Bono, G., Marino, F., & Cosentino, M. (2018). Parkinson's disease patients have a complex phenotypic and functional Th1 bias: cross-sectional studies of CD4+ Th1/Th2/T17 and Treg in drug-naïve and drug-treated patients. *Journal of neuroinflammation*, 15(1), 205. <https://doi.org/10.1186/s12974-018-1248-8>

Kustrimovic, N., Rasini, E., Legnaro, M., Bombelli, R., Aleksic, I., Blandini, F., Comi, C., Mauri, M., Minafra, B., Riboldazzi, G., Sanchez-Guajardo, V., Marino, F., & Cosentino, M. (2016). Dopaminergic Receptors on CD4+ T Naive and Memory Lymphocytes Correlate with Motor Impairment in Patients with Parkinson's Disease. *Scientific reports*, 6, 33738. <https://doi.org/10.1038/srep33738>

Cosentino M., Ferrari M., Kustrimovic N., Rasini E., Marino F. (2015). Influence of dopamine receptor gene polymorphisms on circulating T lymphocytes: A pilot study in healthy subjects. *Human immunology*, 76, 10, 747-752. <https://doi.org/10.1016/j.humimm.2015.09.032>

Keywords: PBMC, Fresh Blood, Neuroimmune-Pharmacology, Parkinson's Disease, Cell isolation, Primary cell culture

MATERIALS

MATERIALS

☒ Ficoll Paque PLUS **Ge Healthcare Catalog #17144003-500 ml**

☒ Fetal Bovine Serum (FBS) **EuroClone Catalog #ECS0180L-500 ml**

☒ RPMI 1640 **EuroClone Catalog #ECM 0495L- 500 ml**

☒ Trypan Blue solution 0.4% **Sigma Aldrich Catalog #T8154- 100 ml**

☒ NaCl **Sigma Aldrich Catalog #S9625**

☒ Na₂HPO₄*7H₂O **Merck Serono GmbH Catalog #1.06574.1000**

☒ NaH₂PO₄ **Merck Serono GmbH Catalog #1.06346.0500**

☒ NH₄Cl **Merck Serono GmbH Catalog #1.01145.1000**

☒ KHCO₃ **Merck Serono GmbH Catalog #1.04854.500**

☒ EDTA **Sigma Aldrich Catalog #ED2SS**

☒ Acetic Acid 100% **Sigma Aldrich Catalog #A6283**

☒ Gentian violet 1% **Marco Viti Catalog #not available**

Instrumentation required:

- Laminar flow hood
- Autoclave

BEFORE START INSTRUCTIONS

If you need to obtain **PBMC for cell culture**, make sure you are using **sterile PBS, culture medium, filtered Lysis Buffer and sterile plastic disposables as well**. Moreover, work under laminar flow hood when you are processing samples. Otherwise, use non-sterile solutions and plastic disposables, and process samples in cell isolation laboratory.

ALL REAGENTS USED IN THIS PROTOCOL MUST BE AT ROOM TEMPERATURE!

- 1 Put the needed amount of blood sampl into a 50 mL conical tube.
- 2 Add an equal volume of **PBS 1X** and mix well.

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SOLUTION- 02 - Phosphate Buffered Saline (PBS)

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3 Place 3 mL of FICOLL in a 15 mL conical tube.

4 Carefully layer 12 mL of diluted blood on FICOLL with a glass Pasteur Pipette to a final volume of 15 ml as shown in the figure below.

- 5 Centrifuge samples 400 x g, 00:40:00 at room temperature (RT) without break.

Equipment

Allegra AVANTI 30

NAME

Centrifuge

TYPE

Beckman Coulter

BRAND

Beckman Italy

SKU

- 6 After centrifugation, take out the tubes carefully to not disturb the mononuclear cell layer that appears as a white, cloudy band between the plasma and FICOLL as shown in the figure below.

- 7 Carefully with a glass Pasteur pipette transfer the mononuclear lymphocyte cell layer to another 15 ml conical tube.

- 8 Wash the isolated PBMC with **PBS/FBS 2%** to a final volume of **10 mL** and centrifuge at **600 x g, 00:10:00** at RT.

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NAME

SOLUTION- 05 - Wash solution (PBS/FBS) for PBMC

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- 9 Remove supernatants, resuspend pellet in **1 mL** of **Lysis Buffer** and add another **9 mL** of **Lysis Buffer**. Immediately centrifuge tubes at **300 x g, 00:10:00** at RT.

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NAME

SOLUTION- 06 - Lysis Buffer

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- 10 Remove supernatant and resuspend pellet in **10 mL** of **PBS/FBS 2%** and centrifuge at **600 x g, 00:10:00** at RT.

Document
NAME SOLUTION- 05 - Wash solution (PBS/FBS) for PBMC
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- 11** Remove supernatant and resuspend the obtained pellet in **10 mL** of **RPMI/FBS 10%** for cell counting.

Document
NAME SOLUTION- 04 - Wash solution (RPMI/FBS) for PBMC
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- 12** **For manual cell count use Türk solution for checking purity.**

Follow protocol **CELL COUNT- 02**.

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NAME

SOLUTION- 08 - Türk solution

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13 OPTIONAL STEP

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For automatic cell count with Cellometer machine use Trypan Blue.

Follow protocol **CELL COUNT- 03**.

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SOLUTION- 09 - Trypan Blue solution

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14 If needed, check the purity of PBMC suspension by using morphological parameter of the flow cytometer.

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For this test 0.5×10^6 PBMC in 500 μ l of PBS are enough.

Equipment	
BD FACS Celesta	NAME
Flow Cytometer	TYPE
Becton Dickinson	BRAND
Milan Italy BD	SKU

15 Expected results

Expected result
<p>VIABILITY - The expected viability by Trypan Blue should be $\geq 90\%$.</p> <p>PURITY - The PBMC suspension obtained should contain at least 80% of lymphocytes, 10-15% of monocytes and few contaminant PMN cells ($\leq 5\%$) as confirmed by flow cytometry.</p> <p>YIELD - The expected amount of PBMCs should be $\pm 28,5 \times 10^6$ starting from 25 ml of fresh blood.</p>