

May 02, 2022

Version 1

Passaging of cells V.1

DOI

<https://dx.doi.org/10.17504/protocols.io.ewov1n3qkgr2/v1>

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Protocol Citation: Guido Krähenbühl 2022. Passaging of cells. **protocols.io**

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Protocol status: Working

We use this protocol and it's working

Created: May 02, 2022

Last Modified: May 02, 2022

Protocol Integer ID: 61793

Keywords: cells general lab procedure, cell passage, cell, procedure

Abstract

General Lab procedure, Cell passage

Before start

Warm up culture media, PBS and Trypsin in a  37 °C Waterbath



- 1 Inspect cells for confluency, when 70–90 % confluency is reached cells are ready for passage
- 2 Discard media and wash once with PBS
- 3 Add Trypsin to fully cover the surface of the culture vessel
- 4 Incubate for  00:05:00 at  37 °C 5m
- 4.1 Lightly tap the culture vessel and check for cell detachment. If cells are still attached prolong incubation
- 5 Add 2/3 of culture media to 1/3 of trypsin and transfer cells into a Falcon tube
- 6 Centrifuge at  300 rcf, Room temperature, 00:05:00 5m
- 7 Discard supernatant and resubstitute with  1 mL culture media
- 8 Count cells and adjust media volume for required cell density
- 9 Plate cells