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Passage cells

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Protocol status: Working

We use this protocol and it's working

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Abstract

Passage Cells



- 1 Observe cells under microscope
- 2 Wipe outside surface of cell culture dish with ethanol cotton and bring it inside Biosafety Cabinet (BSC)
- 3 Bring medium, PBS, Trypsin-EDTA 2x to BSC
- 4 Bring autopipetter, disposable pipet (10ml, 2ml), one 15ml tube to BSC
- 5 Discard old medium
- 6 Rinse the bottom of dish with 10ml PBS
- 7 Add 1ml Trypsin-EDTA 2x to dish
- 8 Incubate the dish in CO2 Incubator 37oC 4min
- 9 Check under microscope (cell shape become rounded)
- 10 Neutralize trypsin-EDTA 2x by add 8ml medium
- 11 Suspend detached cells and transfer to 15ml tube
- 12 Centrifuge 1200 rpm, 4 min
- 13 Bring to BSC, discard medium



14 Add 20 ml DMEM and divided each 10 ml to dish (total 2 dish with @50% cells)